COMPENDIUM REPORT OF RED BLUFF DIVERSION DAMTROPARY JUVENILE ANADROMOUS FISH PRODUCTION INDICESTSOR YEA 2002-2012



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Compendium Report of Red Bluff Diversion Dam Rotaryp Juvenile Anadromous Fish Production Indices for Years 2002-2012

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Abstract.— Fall, late-fall, spring, and winter-run Chinookinsan (Oncorhynchus tshawytschaand Steelhead/Rainbow troutOncorhynchus myki)ss spawn in the Sacramento River and tributaries itiforaia's Central Valley upstream of Red Bluff Diversion Dam (RBDD) throughout the yearmpling of juvenile anadromous fish at RBDD allows for year-round quantitative optrotion and passage estimates of all runs of Chinook and mykiss. Incidental capture of Green Sturgeo Ac (penser medirostris) and various Lamprey species on petra spp. and Entosphenus tridentatus) has occurred throughout juvenile Chinook monitoria optivities since 1995. This compendium report addresses, in detail, juvenile admonous fish monitoring activities at RBDD for the period April 4, 2002 through Septen 80, 2013.

Sampling was conducted along a transect using to transect diameter rotary-screw traps attached via aircraft cable sedity to RBDD. Trap efficiency (i.e., the proportion of the juvenile salmonid population parss RBDD captured by traps) was modeled with percent of river discharge sample to develop a simple least-squares regression equation. Chinook and mykiss passage were estimated by employing the trap efficiency model. The ratio of fry to pre-sit/smolts passing RBDD was variable among years. Therefore, juvenile passage was attaitized to determine juvenile production by estimating a fry-equivalent Juventhe duction Index (JPI) for among-year comparisons. Catch per unit volume (CPUV) was as each index of relative abundance for Green Sturgeon and Lamprey species. Abiotita daollected or calculated throughout sample efforts included: water temperate flow, turbidity, and moon illuminosity (fraction of moon illuminated). The biatic variables were analyzed to determine if relationships existed throughout the gration periods of the anadromous species.

A trap efficiency model developed in 2000 to estienfish passage demonstrated improved correlation between 2002 a20d13 with the addition of 85 mark-recapture trials. The model'ssquared value improved greatly with the addition of numerous mark-recapture trials that used wild frize-class salmon over a variety of river discharge levels. Total passage estimates diring annual effort values with 90% confidence intervals (CI) are presented, by broedry for each run of Chinook. Fry and pre-smolt/smolt Chinook passage estimates with 90% are summarized annually by run in Appendix 1. Comparisons of relative variativithin and between runs of Chinook were performed by calculating Coefficients ariation (CV). Fall Chinook annual total passage estimates ranged between 6,26217 and 27,736,868 juveniles for brood years 2002-2012 (a) = 14,774,923, CV = 46.09% average, fall Chinook passage was composed of 74% fry and 26% pre-smolt/smott-slass fish (SD = 10.3). Late-fall

Chinook annual total passage estimates ranged between 159.99% average, late-fall Chinook passage was composed of 38% fry and 62% pre-simmolt/ssize-class fish (SD = 22.5). Winter Chinook annual total passage estimates raintogetween 848,976 and 8,363,106 juveniles for brood years 2002-2012 (\$\frac{1}{2}\$ = 3,763,652; = 73.2%). On average, winter Chinook passage was composed of 80% fry and 20% inputs/smolt size-class fish (SD = 11.2). Spring Chinook annual total passage estimater spring Chinook ranged between 158,966 and 626,925 juveniles for broodry 22002-2012 (\$\frac{1}{2}\$ = 364,508, CV = 45.0%). On average, spring Chinook passage was seen of 54% fry and 46% presmolt/smolt size-class fish (SD = 20.0). Annutal passage estimates for mykiss ranged between 56,798 and 151,694 juveniles for modal years 2002-2012 (\$\frac{1}{2}\$ = 116,272, CV = 25.7).

A significant relationship between the estimated unaber of adult females and fry-equivalent fall Chinook productestimates was detected (= 0.53, df = 10, P = 0.01). Recruits per female were calculated aantiped from 89 to 1,515 (ੀ = 749). Egg-to-fry survival estimates averaged 13f@%fall Chinook. A significant relationship between estimated number of femalesdafny-equivalent late-fall Chinook production estimates was detected² (\neq 0.67, df = 10,P = 0.002). Recruits per female were calculated and ranged from 47 to 243 (ੀ = 13 E)gg-to-fry survival estimates averaged 2.8% for late-fall Chinook. A significælationship between estimated number of females and fry-equivalent winter Chinopkoduction estimates was detected ($\hat{r} = 0.90$, df = 10,P < 0.001). Recruits per female were calculated ranged from 846 to 2,351 (ੀ = 1,349). Egg-to-fry surv**exti**mates averaged 26.4% for winter Chinook. No significant relationship between estiled number of females and fryequivalent spring Chinook production estimates where equivalent spring Chinook production estimates where experience is a spring chinook production estimate where experience experience is a spring chinook production estimate where experience experien 0.971). Recruits per female were calculated ambæd from 1,112 to 8,592 (ୀ = 3,122). Egg-to-fry survival estimates averaged 61.5% foringp Chinook. Spring Chinook juvenile to adult correlation values appear unreasble and well outside those found for other runs and from other studies.

Catch of Green Sturgeon was highly variable, nootmally distributed and ranged between 0 and 3,701 per year (median 93).1 Catch was primarily composed of recently emerged, post-exogenous feedarvae. The 10-year median capture total length averaged 27.3 mm (SD = 0.8) pen Sturgeon annual CPUV was typically very low and ranged from 0.0 to 20.1 feethft (\hat{j} = 2.5 fish/ac-ft, SD = 5.9). Data were positively skewed and median annual OPath 0.8 fish/ac-ft.

Lamprey species sampled included adult and jure Philoific Lamprey (Entosphenus tridentatu) sand to a much lesser extent River Lampteym (petra ayre) i and Pacific Brook Lampreya (mpetra pacifica). Unidentified lamprey ammocoetes and Pacific Lamprey composed 99.8% of all captures, 246% 75%, respectively. River Lamprey and Pacific Brook Lamprey composed the irreing 20.2%, combined. Lamprey captures occurred throughout the year between Octoband September. Lamprey ammocoete annual relative abundance ranged from 60.611.7 fish/ac-ft (1 = 6.8 fish/ac-ft, SD = 2.6). Overall, these data were medly distributed as median annual CPUV was 6.5 fish/ac-ft, similar to the mean value acific Lamprey macropthalmia

annual relative abundance was generally higher that mocoete relative abundance and ranged from 2.1 to 112.8 fish/ac-ft ($^{\circ}$ = 41isth fac-ft, SD = 34.7). Overall, Pacific Lamprey data was slightly positively skewed and innect PUV was 34.1 fish/ac-ft.

Tabular summaries of the abiotic conditions enothered during each annual capture period were summarized for each roufnsalmon, O. mykiss, Green Sturgeon and Lamprey species. The range of tempress experienced by Chinook fry and pre-smolt/smolts in the last 11 years of passaged RBDD have been within the optimal range of temperature tolerances for juveen@hinook survival. Green Sturgeon have likely benefitted from temperature managemeerfforts aimed at winter Chinook spawning and production, albeit less comprehens/ivel/amprey species have also likely benefitted from temperature management as temperature for early life stages of Lamprey in the mainstem Sacramento River appealmature been, on average, optimal in the last 11 years.

The relationship between river discharge, turbyditand fish passage are complex in the Upper Sacramento River whereap cand stream-type Chinook of various size-classes (i.e., runs, life stages agress) amigrate daily throughout the year. Fish passage increases often coincided with are insurer in turbidity which were sampled more effectively than increases in river discharge positive bias of fish passage estimates may result if the peak turbidity event swaampled following an un-sampled peak flow event. The importance of the first storement of the fall or winter period cannot be overstated. Smolt passage and juvenibent plrey passage increase exponentially and fry passage can be significant indufall storm events.

Rotary trap passage data indicated fry size-classister Chinook exhibit decreased nocturnal passage levels durindparound the full moon phase in the fall. Pre-smolt/smolt winter Chinook appeared less uenced by nighttime light levels and much more influenced by changes in dischargelse Spring, fall and late-fall Chinook fry exhibited varying degrees of decreases age during full moon periods, albeit storms and related hydrologic influx domieal tpeak migration periods.

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Introduction

The United States Fish and Wildlife Service (U\$FishSconducted direct monitoring of juvenile Chinook salmoon(corhynchus tshawytschap)ssage at Red Bluff Diversion Dam (RBDD; RM 243) on the SacramReinter, CA since 1994 (Johnson and Martin 1997). Martin et al. (2001) developed aptitative methodologies for indexing juvenile Chinook passage using rotaryvscoraps to assess the impacts of the RBDD Research Pumping Plant. Absolute abundarocalu (point and passage) estimates were needed to determine the level of importion the entrainment of salmonids and other fish community populations though experimental 'fish friendly' Archimedes and internal helical pumps (Borthwick Dorwin 2001). The original project objectives were met by 2000 and funding the project was discontinued.

In 2001, funding was secured through a CALFED Etay-Program grant for three years of annual monitoring operations to determitine effects of restoration activities in the Upper Sacramento River aimed primarily attent Chinook salmon. Through various amendments, extensions, and grant approximation CALFED Ecosystem Restoration Program, the State of California baseding source lasted until 2008. At this point, the State of California defaulted orethfunding agreement and internal USFWS funding sources through the Central Valleige Remprovement Act (CVPIA) bridged the gap for a period of time until Statentiang was restored. The US Bureau of Reclamation, the primary proponent of the Centralley Project (CVP) of which this project provides monitoring and abundance trencommation, has funded this project since 2010 due to regulatory requirements containwithin the Biological Opinion for the Operations and Criteria Plan for the CVP (NEURS).

Protection, restoration, and enhancement of anadious fish populations in the Sacramento River and its tributaries is an importanement of the CVPIA Section 3402. The CVPIA has a specific goal to double populations and anadromous fishes in the Central Valley of California. Juvenile salmonid production itoring is an important component authorized under Section 3406 (b)(160) IPIA and has funded many anadromous fish restoration actions which were ineth in the CVPIA Anadromous Fisheries Restoration Program (AFRP) Working PaperWS 1995), and Draft Restoration Plan (USFWS 1997; finalized in 2001).

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The National Marine Fisheries Service first list/Meinter-run Chinook salmon as threatened under threegency listing procedures for the ESA (16 U.S.C.R. 1531-1543) ugnsA4, 1989 (54 FR 32085). A proposed rule towaidter Chinook salmon to the list of threatened species beyond expirationthe emergency rule was published by the NMFS omcMal20, 1990 (55 FR 10260). Winter Chinook salmon were formally addedthe list of federally threatened species by finale on November 5, 1990 (55 FR 46515), and they were listed as a federally angered species on January 4, 1994 (59 FR 42400) cal habitat for winter Chinook salmon has been designated from Keswick (Palath 302) to the Golden Gate Bridge (58 FR 332/1/2; 1/6, 1993). Winter Chinook salmon have been listed as endangered untider CESA since September 22, 1989 (California 600 Regulations, Title XIV, Section 670.5). Their federal endangered status vessefirmed in June 2005 (70 FR 37160).

Since 2002, the USFWS rotary trap winter Chinometrijle production indices (JPI's) have primarily been used in support of problem estimates generated from carcass survey derived adult escapement data utsie on all Oceanic and Atmospheric Administration's (NOAA) Juvenile Producestimate Model. Martin et al. (2001) stated that RBDD was an ideal location to intro juvenile winter Chinook production because (1) the spawning grounds octanost exclusively above RBDD (Vogel and Marine 1991; Snider et al. 1997, USFOW\$)2(2) multiple traps could be attached to the dam and sample simultaneously as ransect, and (3) operation of the dam could control channel morphology and hydroidal characteristics of the sampling area providing for consistent sampling ditions for purposes of measuring juvenile fish passage.

Fall, late-fall, spring, and winter-run Chinooknaan and Steelhead/Rainbow Trout (Oncorhynchus myki)sspawn in the Sacramento River and tributariestreps of RBDD throughout the year resulting in year-roundejule salmonid passage (Moyle 2002). Sampling of juvenile anadromous fish at IRBID ws for year-round quantitative production and passage estimates of all runs of Othik and Steelhead/Rainbow trout. Timing and abundance data have been provided in time for fishery and water operations management purposes of the CVP since 2009, confidence intervals, indicating uncertainty in weekly passage mates, have been included in real-time bi-weekly reports to allow better managent of available water resources and to reduce impact of CVP operations on both fattendangered Species Act (ESA) listed and non-listed salmonid stocks. Currer key cramento River winter Chinook are ESA listed as endangered. Central Valley spriing Othand Central Valley Steelhead (hereafterO. mykiss) are listed as threatened within the Central Valley angered Species Unit.

Incidental capture of Green Sturgeofac (penser medirostr) sand various Lamprey species (Lampetra spp. and Entosphenus spas) occurred throughout juvenile Chinook monitoring activities at RBDD since 1995 (Gainels Maartin 2002). Although rotary traps were designed to capture outmigrating salmobs in olts, data from the incidental capture of sturgeon and lamprey species has become easingly relied upon for basic life-history information and as a measure of relatiabundance and species trend data. The Southern distinct population segment of the thornerican Green Sturgeon was proposed for listing as threatened under the Feders A on April 7, 2006 (FR 17757) which then took effect June 6, 2006. Pacific Laery (Entosphenus tridentatu) sare thought to be extirpated from at least 55% of the istorical habitat and have been recognized by the USFWS as a species needing are to conserve and restore these fish (Goodman and Reid 2012).

The objectives of this compendium report are to): \$\(\mathbb{U}\) mmarize the estimated abundance of all four runs of Chinook salmon \(\theta\) nthykiss passing RBDD for brood

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² Real-time biweekly reports located for download https://www.fws.gov/redbluff/rbdd biweekly final.htm

years (BY) 2002 through 2012, (2) estimate annelative abundance of Green Sturgeon and Lamprey species production for electronsecutive years, (3) define temporal patterns of abundance for all anadromopesies passing RBDD, (4) correlate juvenile salmon production with adult salmon escapent estimates, (5) perform exploratory data analyses of potential environmentavariates driving juvenile fish migration trends, and (6) describe various lifetonic attributes of anadromous juvenile fish produced in the Upper Sacramento River as inheited through long-term monitoring efforts at RBDD.

This compendium report addresses, in detail, owejuile anadromous fish monitoring activities at RBDD for the period April2002 through September 30, 2013. This report includes JPI's and relative abundarstienates for the 2002-2012 brood year emigration periods and will be submitted toetcalifornia Department of Fish and Wildlife to comply with contractual reporting requiments for Ecosystem Restoration Program Grant Agreement Number P0685507 and to Ltae Bureau of Reclamation who funded in part or in full the surveys from years 20through 2013 (Interagency Agreement No. R10PG20172).

Study Area

The Sacramento River originates in Northern Callidonear Mt. Shasta from the springs of Mt. Eddy (Hallock et al. 1961). It slowwith through 370 miles of the state draining numerous slopes of the coast, Klamathcades, and Sierra Nevada ranges and eventually reaches the Pacific Ocean via San scanday (Figure 1). Shasta Dam and its associated downstream flow regulating structukeswick Dam, have formed a complete barrier to upstream anadromous fish passaignce 1943 (Moffett 1949). The 59-river mile (RM) reach between Keswick Dam (RM) and RBDD (RM 243) supports areas of intact riparian vegetation and largely aims unobstructed. Within this reach, several major tributaries to the Sacramento upstreaf RBDD support various Chinook salmon spawning populations. These include Clearekand Cottonwood Creek (including Beegum Creek) on the west side of the samento River and Cow, Bear, Battle and Payne's Creek on the east side (Figureselow RBDD, the river encounters greater anthropogenic impacts as it flows souththe Sacramento-San Joaquin Delta. Impacts include, but are not limited to, channelian, water diversion, agricultural and municipal run-off, and loss of associated riparinagetation.

RBDD is located approximately 1.8 miles southefatsteocity of Red Bluff, California (Figure 1). The dam is 740-feet (ftd)ewaind composed of eleven, 60-ft wide fixed-wheel gates. Between gates are concretespateft in width. The USBR's dam operators were able to raise the RBDD gates allowform run-of-the-river conditions or lower them to impound and divert river flows intbet Tehama-Colusa and Corning canals. USBR operators generally raised the RBTDD fgom September 16 through May 14 and lowered them May 15 through Septemberduting the years 2002-2008. As of the spring of 2009, the RBDD gates were impeliolowered prior to June 15 and

were raised by the end of August or earlier (NMBSQ) in an effort to reduce the impact to spring Chinook salmon and Green Sturgesince the fall of 2011, the RBDD gates have been left in the raised position allogvimobstructed upstream and downstream passage of adult and juvenile anadromfosts. The RBDD has been replaced by a permanent pumping plant upstreamhost RBDD and the facilities have been relinquished to the Tehama Colusa Canal Aitthas of spring 2012. Mothballing of the RBDD infrastructure was scheduled to oxio 2014.

Methods

Sampling Gear—Sampling was conducted along a transect using &fu diameter rotary-screw traps (E.G. Solutions® Clissy Diregon) attached via aircraft cables directly to RBDD. The horizontal placero £rotary traps across the transect varied throughout the study but generally sampled hie river-margin (east and west river-margins) and mid-channel habitats simultans yu(Figure 2). Rotary traps were positioned within these spatial zones unless sampling equipment failed, river depths were insufficient (< 4-ft), or river hydrology reisted our ability to sample with all traps (water velocity < 2.0 ft/s).

Sampling Regimes-In general, rotary traps sampled continuouslyotinghout 24hour periods and samples were processed once dallyring periods of high fish abundance, elevated river flows, or heavy debrized traps were sampled multiple times per day, continuously, or at randomly pressed periods to reduce incidental mortality. When abundance of Chinook was very highb-sampling protocols were implemented to reduce listed species take and ienial mortality in accordance with National Marine Fisheries Service (NMFS) Sectican (110(A) research permit terms and conditions. The specific sub-sampling protocollemented was contingent upon the number of Chinook captured or the probability of sessfully sampling various river conditions. Initially, rotary trap cones were stturally modified to only sample onehalf of the normal volume of water entering the centering the centering and Poytress 2004). If further reductions in capture were needed, the nuembof traps sampled was reduced from four to three. During storm events and as sobed elevated river discharge levels, each 24-hour sampling period was divided into fousix non-overlapping strata and one or two strata was randomly selected for samplif Martin et al 2001). Estimates were extrapolated to un-sampled strata by dividinated by the strata-selection probability (i.e.,P = 0.25 or 0.17). If further reductions in efforter needed or river conditions were intolerable, sampling was discontid or not conducted. When days or weeks were unable to be sampled, mean daily agas sestimates were imputed for missed days based on weekly or monthly mean dailineates (i.e., interpolated).

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Daily Passage Estimat(\mathfrak{B}_{a}).—The following procedures and formulae were used to derive daily and weekly estimates of total number unmarked Chinook and mykiss passing RBDD. We define as catch at trap(i = 1,...t) on dayd (d = 1,...,n),

and X_{di} as volume sampled at trap(i = 1, t) on dayd (d = 1, ...n). Daily salmonid catch and water volume sampled were expressed as:

$$C_d = \int_{i=1}^t C_{di}$$

and,

$$X_d = \int_{i=1}^t X_{di}$$

The %Q was estimated from the ratio of water volume sate (X_d) to river discharge (Q_d) on dayd.

$$\%\hat{Q}_d = \frac{X_d}{Q_d}$$

Total salmonid passage was estimated on dd ady = 1,...,n) by

$$\hat{P}_d = \frac{C_d}{\hat{T}_d}$$

where,

5.
$$\hat{T}_d = (a)(\%\hat{Q}_d) + b$$

and, T_d = estimated trap efficiency on daty

Weekly Passage).—opulaS on totals for numbers of Chinook andhykiss passing RBDD each week were derived from where there are days within the week:

$$\hat{P} = \frac{N}{n} \int_{d=1}^{n} \hat{P}_{d}$$

Estimated Variance-

7.
$$Var(\hat{P}) = (1 - \frac{n}{N}) \frac{N^2}{n} s_{\hat{p}_d}^2 + \frac{N}{n} \int_{d=1}^n Var(\hat{P}_d) + 2 \int_{i*j}^n Cov(\hat{P}_i, \hat{P}_j)$$

The first term in eq. 7 is associated with sampbifiglays within the week.

8.
$$s_{\hat{P}_d}^2 = \frac{(\hat{P}_d - \hat{\overline{P}})^2}{n-1}$$

The second term in eq. 7 is associated with $estingaP_d$ within the day.

9.
$$Var(\hat{P}_d) = \frac{\hat{P}_d(1 - \hat{T}_d)}{\hat{T}_d} + Var(\hat{T}_d) \frac{\hat{P}_d(1 - \hat{T}_d) + \hat{P}_d^2 \hat{T}_d}{\hat{T}_d^3}$$

where,

10.
$$Var(\hat{T}_d)$$
 = error variance of the trap efficiency model

The third term in eq. 7 is associated with estimgtboth P_i and P_j with the same trap efficiency model.

11.
$$Cov(\hat{P}_i, \hat{P}_j) = \frac{Cov(\hat{T}_i, \hat{T}_j)\hat{P}_i\hat{P}_j}{\hat{T}_i\hat{T}_j}$$

where,

12.
$$Cov(\hat{T}_i, \hat{T}_i) = Var(\hat{*}) + x_i Cov(\hat{*}, \hat{*}) + x_i Cov(\hat{*}, \hat{*}) + x_i x_i Var(\hat{*})$$

for some $\hat{T}_i = \hat{*} + \hat{*}x_i$

Confidence intervals (CI) were constructed aroundsing eq. 13.

13.
$$P \pm t_{*/2,n-1} \sqrt{Var(\hat{P})}$$

Annual JI's were estimated by summin gacross weeks.

$$JPI = \int_{week=1}^{52} \hat{P}$$

Fry-Equivalent Chinook Production EstimateThe ratio of Chinook fry (<46 mm FL) to pre-smolt/smolts (>45 mm FL) passing RBDD/sriable among years. Therefore, we standardized juvenile production stylerating a fry-equivalent JI for among-year comparisons. Fry-equivalent JI's westimated by the summation of fry JI and a weighted (1.7:1) pre-smolt/smolt JI (inverse value of 59% fry-to-presmolt/smolt survival; Hallock undated). Rottrap JI's could then be directly compared to determine variability in production breaten years.

Relative Abundance.Catch per unit volume (CUV; Gaines and Martin 2) Was used as an index of relative abundance (RA) foe 68 turgeon and Lamprey species at RBDD.

$$RA_{dt} = \frac{C_{dt}}{V_{dt}}$$

 RA_t = relative abundance on dalyby trapt (catch/acre-foot),

C_{dt} = number of fish captured on dayby trapt, and V_{dt} = volume of water sampled on dayby trapt.

The volume of water sampled (1)/was estimated for each trap as the product of one half the cross sectional area (wetted portion) **be**tcone, water velocity (ft/s) directly in front of the cone at a depth of 2-feet, cone modifi (multiplied by 0.5) or not (multiplied by 1.0), and duration of sampling.

Exploratory Data Analyses The sampling of four runs of Chino Ok, mykiss, Green Sturgeon, and Lamprey occurred over 11 years as variety of environmental conditions. Abiotic data collected or calculate dotughout sample efforts included water temperature, flow, turbidity, and moon illuminisity (fraction of moon illuminated). The abiotic factors were analyzed tetermine if patterns or trends existed throughout the migration periods of the icars species. Additional statistical analyses were performed, when applicable, and addit methods are noted within the results section for species-specific data treaddalyzed.

Results

Sampling Effort—Annual sampling effort varied throughout the 1day period of reporting. The reasons for less than 100% effarted by time of year and run sampled due to numerous factors. These factors can be grantized as either intentional or unintentional decreases in effort. Intentional decreases in effort were primarily due to ESA Section 10(a)1(A) take and incidental mortantitys, the desire to decrease potential impacts to ESA listed fish or hatcherly assed production groups, or when staffing levels were not appropriate for the condits encountered. Unintentional decreases in effort were due primarily to storminity and related debris flows or conditions considered too dangerous to sample. it and related debris flows or conditions considered too dangerous to sample. it and related debris flows or conditions considered too dangerous to sample. It also due to operational requirements imposed by USBR operators (e.g., linguager raising of the dam gates).

Annual sample effort was assigned a value of \$1500 on sampling four traps 365 days a year. Annual sample effort values by salchespecies and run are described in Table 1. Overall, annual sample effort for all restricts combined ranged from 0.53 to

0.91 (f) = 0.80, SD = 0.10) following annual jue estailmonid brood year cycles. The lowest values corresponded to the year 2002 whem stang did not begin until mid-April of the year. The highest value correspont the the year 2007 when flow events were mild, staffing levels were optimal, and permeistrictions did not dictate major sampling effort reductions (Table 1).

Mark-Recapture Trials—Trap efficiency estimates were calculated by conting mark-recapture trials (Volkhardt et al. 2007) usungmarked salmon collected from daily trap samples. Trials were conducted when tratch values allowed the release of 1,000 fish per trial, generally, as well as whearffiring and river conditions would allow. Mark-recapture trials were also employed to validately trap efficiency estimates by comparing actual with predicted (modeled) estimates was especially important during peak salmon outmigration periods.

The number of trials conducted each calendar **yeaged** from 0 in 2010 to 21 in 2004 ($\hat{I} = 7.7$) and totaled 85 trials between 2002 2013 (Table 2). Trials were conducted with four rotary traps (N = 74) or three traps (N = 11). Some trials were conducted with cones modified to sample half the uroe of water (N = 25) or mixed (N = 1), but primarily unmodified and sampling full **eff(N** = 59). Trap efficiencies were tested with the RBDD gates raised ($\hat{I} = 7.7$) and lowered (N = 13) during the years when RBDD was in operation (Table 2).

Trials were conducted through a variety of flowdarap effort conditions representing actual sampling conditions detecter throughout various fish migration periods (Table 2). Estimates of the percentage well water volume sampled by traps (%Q) ranged from 0.72 to 6.87% ($^{\circ}$ = 3.10, SD = 1.Bf)ciency estimates for the 85 trials ranged from 0.34 to 5.48% ($^{\circ}$ = 2.37%, SD1).0

Released fish groups ranged from 340 to 5,143/iddials (f) = 1,598) and recaptured fish numbers ranged from 7 to 119 (f) per trial. Trials were conducted predominantly with fry size-class (<46 mm fork lend) gnaturally produced fall Chinook (67%) and to a lesser extent winter Chinook (2276) als were conducted in some years using unmarked pre-smolt/smolts (11%) following transport to leman National Fish Hatchery Fall Chinook production releases ring spring, as conditions and staffing levels allowed (Table 2).

Average fork lengths of release groups in the fixe-class had fork lengths ranging from 35.5 to 57.1 mm ($^{\circ}$ l = 37.2 mm). Recapture of fengths ranged from 34.6 to 62.4 mm ($^{\circ}$ l = 37.3 mm). Average fork lengths of fishare of in the pre-smolt/smolt size-class ranged from 68.7 to 81.2 mm ($^{\circ}$ l = 75.3 mme) calculate of the lengths ranged from 61.3 to 80.2 mm ($^{\circ}$ l = 75.3 mm; Table 2). A pairtest was performed on the average

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⁵ Coleman National Fish Hatchery is located upstreafintBDD on Battle Creek a tributary to the Sacratione Fall Chinook production fish (~12 million per year) were adipostic ped (i.e., marked) in varying proportions of the years of study between 0 and 25%. Unmarked fish were included in some iefficy trials as they could not be distinguished in actually produced fish.

release and recaptured fish lengths for all triants indicated no significant difference between the released and recaptured fish sizes (0.759, df = 83, t = -0.308)

Trap Efficiency Modeling-Between 1998 and 2000, Martin et al. (2001) developed a trap efficiency model for the RBDD ryotæpping operation by conducting 58 mark-recapture trials (one trial excluded duezero efficiency value). These data were used as the basis of the trap efficiency modelalculate daily passage estimates. The model was further developed between 2002 and 20th the addition of 85 mark-recapture trials. Trap efficiency was positively related to (%Q), with higher efficiencies occurring as the relative percental electrical evolume sampled by rotary traps increased. Trap efficiency was inversely teel to river discharge Q), as river discharge increased, trap efficiency decreased.

As mark-recapture trials were conducted, the tefficiency model was typically updated one time each year. The newest model was jied on July 1 of each year, the beginning of the annual winter Chinook juvenile dotoyear period. Between 2002 and 2013 nine different models were utilized. The sipectates and model parameters with P-values used throughout the reporting period assedict chronologically below the groups of mark-recapture trials incorporated intermodels in Table 2. The net result over the 11-year period was stabilization and imperment of the trap efficiency model with the addition of 85 mark-recapture trials. v@all, theP-values indicated a high level of significance for the paramet@Q in all years (P< 0.001). The model's-squared value dropped in the first few years and then imped greatly with the addition of numerous naturally produced fry size-class mark-peter trials over a variety of river discharge levels (Table 2; Figure 3).

Chinook Capture Fork Length AnalyseShinook run assignment based on length-at-date (LAD) criteria was originally developed from rowth data in the Upper Sacramento River at the Tehama Colusa Fish Fausility fall Chinook production records from 1972 through 1981 (Fisher 1992). Stimeate of apparent growth rate was originally developed from fall Chinook < 90 Filmas fish migrated or were depleted from the spawning channels by this sizish(€r 1992). Johnson et al. (1992) further developed (extrapolated) the data to predict for fish ≥90 mm and ≤250 mm FL. The data was further refined by Frank FishtereoCalifornia Department of Fish and Game, whereby estimated growth curves were proced for all runs based on adult timing, water temperatures, and juvenile emergenticeing and growth (Brown and Greene 1992). The growth curves were fitted totale of daily growth increments (i.e., fork length at age in days) by the California Depart of Water Resources in the early

1990's (Brown and Greene 1992; Greene 1992). dllowing fork length data encompassed fish sampled by rotary traps using the tables up to 180 mm FL, as fish were rarely captured above this length (i.e., extre outliers).

Fall Chinook sampled from brood years 2002-2012 weavily weighted to the fry size-class category (<46mm). On average, 75 f all fish sampled as fall could be described as fry (SD = 6.9) with 71.0% of the tegasturing less than 40 mm FL (Figure 4a). The remaining 24.3% (SD = 6.9) were attributed pre-smolt/smolt category (>45 mm) with fish between 70 and 89 mm compositing of that value. Overall, fall Chinook were sampled between 30 and 134 mm annuality trivial numbers below or above this range (Figure 4b). Fall Chinook should be growth, on average, between December and March, followed by a significant in length in April, followed by more moderate and variable growth through Novem (Figure 4c). The growth pattern exhibited by fall Chinook appears strongly influent by the duration of the fall Chinook spawning period and the LAD criteria. Beginning of the fall Chinook instead of fall of fall of fall Chinook during the fitted weeks of April.

Late-fall Chinook sampled from brood years 2002220ere not heavily weighted to the fry size-class category (<46mm). On aver249% of all fish sampled as late-fall could be described as fry (SD = 12.8) with 96.31% eofry measuring less than 40 mm FL (Figure 5a). The remaining 75.1% (SD = 12.8) atteiteuted to the pre-smolt/smolt category (>45 mm) with fish between 70 and 89 mmposing 48.3% of that value. Overall, late-fall Chinook were sampled betweem26 180 mm annually (Figure 5b). Late-fall Chinook showed little growth, on averagetween April and May, followed by a significant increase in length in June and Jollowed by more moderate and variable growth between late-September and February (Figure The growth pattern exhibited by late-fall Chinook appears modestly up ficed by the LAD criteria. Beginning on July 1, newly emerged fry were cliests winter Chinook instead of latefall Chinook slightly increasing the median for note of late-fall Chinook during the first few weeks of July. In mid-September and to a leextent in late-December, the overall fork length distribution for late-fall Cloick increases from one week to the next and was likely a result of decreased sampling effore to RBDD gate operations and initial winter storms.

Winter Chinook sampled from brood years 2002-20/42e heavily weighted to the fry size-class category (<46mm). On average,% of all fish sampled as winter could be described as fry (SD = 8.8) with 92.86% coffry measuring less than 40 mm FL (Figure 6a). The remaining 22.1% (SD = 8.8) where the used to the pre-smolt/smolt category (>45 mm) with fish between 46 and 69 mm posing 85.3% of that value. Overall, winter Chinook were sampled between 27 462 mm annually (Figure 6b). Winter Chinook showed little growth, on average whose July and October, followed by a significant increase in length in mid-October by more moderate growth through December. The growth pattern was then hyghariable between January and

April (Figure 6c). The growth pattern exhibited winter Chinook appears moderately influenced by the LAD criteria. Beginning on Oeto166, newly emerged fry were classified as spring Chinook instead of winter Othich thereby significantly increasing the median fork length of winter Chinook during these two weeks of October.

Spring Chinook sampled from brood years 2002-202122 slightly weighted to the fry size-class category (<46mm). On average, 56f2216 fish sampled as spring could be described as fry (SD = 19.6) with 90.0% of the feasuring less than 40 mm FL (Figure 7a). The remaining 41.4% (SD = 19.6) attributed to the pre-smolt/smolt category (>45 mm) with fish between 70 and 89 mmposing 69.2% of that value. Overall, spring Chinook were sampled between 28 tatal mm annually (Figure 7b). Spring Chinook showed moderate growth, on average ween October and mid-December, followed by more consistent increasing with through May (Figure 7c). Spring Chinook disappear from the catch typically the with sporadic capture of large smolts in July of some years. The growth patternile ted by spring Chinook appears moderately influenced by the LAD criteria. Begignon December 1, newly emerged fry were classified as fall Chinook instead of the specific followed by resulting in positive size-class bias for spring Chinook.

O. mykiss Capture Size Analyses ollowing the conventions used by Gaines and Martin (2002) size categorization for mykiss followed a slightly different pattern than Chinook and was organized by fork length as fry (n4h), sub-yearling (41–138 mm), and yearling (>138 mm). Moyle (2002) described acento RiverO. mykiss populations as highly variable, but typically relagh140-150 mm FL in their first year. The focus of our data reporting is age-0 and the focus of our size-class analyses was primarily < 139mm and secondarily < 200 mm for the region of the region of

O. mykiss sampled from calendar years 2002-2012 were heavelighted towards the 41-80 mm size-class (79.2%; Figure 8a) which the the sub-yearling category (Figure 8b). On average, a modest 8.2% could the coaized as fry (Table 3). Overall, O. mykiss yearling and estimated age-2 fish were annual type at at rates of 2.4% and 0.6%, respectively (Table 3). There was little at was detected within any size-class between categories, yet variance in weekly captumes high throughout the year (Figure 8c). The variable life-history strate of 9. mykiss resident and anadromous forms was evident from our size-class capture datageneral, newly emerged fry occurred in early-April and increased in size to be able to be a second cohort of either resident trout or summer steelhe was sampled which demonstrated a secondary growth pattern through December (Figure) 8

O. mykiss CAMP Program Life-Stage Comparisonsmykiss capture patterns appeared to be different than that of Chinook salmas relatively few. mykiss were captured as fry (1 = 8.3%) and the majority weren steed as sub-yearlings (1 = 88.7%;

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⁶ Summer steelhead are believed to be extirpated eithe construction of dams blocked access to heathwhabitat (Moyle 2002).

Table 3; Figure 8b). Fry capture was highest 0220nd 2006 (11.2% and 17.5%) although these years sampled the first and third fet O. mykiss of the 11 years, respectively. Yearling and age-2 capture was getlydow averaging only 3.0%.

Life stage classification of fry was uniform the bout all years (f = 6.8%, SD = 2.6%) and did not vary greatly in 2002 and 2006 intrast to age classification. arr and silvery-parr accounted for 91.5% of the mykiss handled at RBDD although there was a large difference between the two categories,0% and 17.5% respectively. Annual variability in parr and silvery-parr classifions (SD = 15.5 and 16.8) seemed to change after 2005 and was likely due to a protected nge or interpretation of morphological characteristics by field staff. Juiles showing signs of anadromy (i.e., smolts) made up only 1.6% of individuals sampled.

O. mykiss Weight-Length AnalysisLog₁₀ transformedO. mykiss weight-length data showed a strong overall relationship betwebe two variables (r= 0.942, Table 4). The annual slope coefficients for the 11-yeariod varied slightly, ranging from 2.858 to 3.052. The variability in growth was **not**hsidered significant as the 95% CI annual slope coefficients encompassed the slopeficient of the overall mean (Table 4). Typical of most weight-length models (ope afficient 2007), the variability about the regression increased with the overall lengththeef fish (Figure 9).

Salmonid Passage-assage estimates for the four runs of Chinooknee calculated weekly as fry and pre-smolt/smolt passage he sum of the weekly fry and pre-smolt/smolt passage values equal the weekly passage values. Confidence intervals (CI) were calculated at the 90% levelaforuns for weekly passage estimates. Weekly CI values were summed to obtain the annulation of the annual passage estimate (i.e., summed weekly passage estimate) gative CI values were set to zero and result in some years CI's being asymmetrical rad the annual passage estimate. Annual passage estimates (i.e., total passage estimate), by brood year, with CI's and annual effort values are presented for Chinook with ables 5a-5d and graphically in Figures 10, 12, 14, and 16. Fry and pre-smolt/scoloinook passage estimates with 90% CI's summarized annually by run can be four prendix 1 (Tables A1-A8). Comparisons of relative variation within and betweens of Chinook were performed by calculating Coefficients of Variation (Sokal Rooth If 1995) of passage estimates.

Fall Chinook annual passage estimates ranged leet we627,261 and 27,736,868 juveniles for brood years 2002-2012 (\$\daggerapsilon = 14,774,928 = 46.2%; Table 5a). On average, fall Chinook passage was composed of 74% fry alkdp26-smolt/smolt size-class fish (SD = 10.3). roportions as low as 56% and as laisy 187% fry were detected (Table 5a). Annual effort values resulted in interpolations between 9 and 60% of annual passage estimates (\$\daggerapsilon = 28\%). In general, the effect of partner front on CI width indicated greater spread of CI's with decreasing effort (Figure 10).

On average, weekly fall passage equated to 5% to fannual fall Chinook passage between mid-January and early March (Figure 1 Weekly passage varied considerably during this period with some weeks page totals accounting for >25% of annual passage values. Between BY 2002 and 25% of average annual passage occurred by the end of March, signifying January to March as the greatest period of migration. A second, albeit much diminished, to passage occurred between late April and May of each year due to the releasenmarked fall Chinook production fish from Coleman National Fish Hatchery. The stee fould not be distinguished from wild fish due to fractional marking processes that ied over the 11-year period from 0 to 25%. Overall, fall passage was complete by ether of July each year with sporadic small pulses of smolts through November (Figure).11b

Late-fall Chinook annual passage estimates rangebolden 91,995 and 2,559,519 juveniles for brood years 2002-2012 ($\hat{l} = 447,7 \text{V1}$; C159.9%; Table 5b). On average, late-fall Chinook passage was composed of 38%nth/62% pre-smolt/smolt size-class fish (SD = 22.5). roportions as low as 11% and as 72% fry were detected (Table 5b). Annual effort values resulted in interpolation between 9 and 56% of annual passage estimates ($\hat{l} = 31\%$). The effect of areflect on CI width indicated greater spread of CI's with decreasing effort due to hateh releases, in general (Figure 12).

On average, weekly late-fall passage started allowaged held at ≤5% of total annual passage between April and May (Figure 106e)ekly passage varied considerably during this period with some weeks \$100 passage totals accounting for >35% of annual passage values. A second, similar magnituothe of passage occurred between July and August in most years. A third, albeitiolismed, mode occurred during October and November with passage accounting fotou \$25% of the annual run in some years. Between BY 2002 and 2012, 75% of \$200 passage occurred by mid-September, signifying April through September the greatest period of migration. Overall, late-fall passage was complete by the cefnole cember each year with sporadic small pulses of smolts through February (Figure).13b

Winter Chinook annual passage estimates ranged **betw**848,976 and 8,363,106 juveniles for brood years 2002-2012 ($^{\circ}$ = 3,763,862;= 73.2%; Table 5c). On average, winter Chinook passage was composed of 80% fry29f%d pre-smolt/smolt size-class fish (SD = 11.2). roportions as low as 53% and as 90% fry were detected (Table 5c). Annual effort values resulted in interpolation between 8 and 42% of annual passage estimates ($^{\circ}$ = 18%). The effect of areffcat on CI width indicated greater spread of CI's with decreasing effort due to subplaining measures during peak migration periods (i.e., take or impact reduction), general (Figure 14).

On average, weekly winter passage increased densities through September to a peak into early October. Weekly passage varied integrably during August through December with some weeks' passage totals accounting 20% of annual passage values. Between BY 2002 and 2012, 75% of average apassage occurred by mid-

October. Weekly passage between October and De**cerind**licated wide variability over the 11-year period, yet the trend showed strathereases followed by a second increase or mode of winter passage in November landember (Figure 15a). Overall, winter passage was 99% complete by the defrollecember each year with sporadic pulses of smolts through March that contited minimally to the annual total winter passage estimate (Figure 15b).

Spring Chinook annual passage estimates rangeddeetw 458,966 and 626,925 juveniles for brood years 2002-2012 (=364,508=C45.0%; Table 5d). On average, spring Chinook passage was composed of 54% fry Chinoper-smolt/smolt size-class fish (SD = 20.0). roportions as low as 24% and the as 91% fry were detected (Table 5d). Annual effort values resulted in interpolation between 1 and 49% of annual passage estimates (=29%). The effect of are float on CI width indicated a slightly greater spread of CI's with decreasing effort does to be sampling during winter storm events, in general (Figure 16).

On average, weekly spring passage started abraptlyheld at roughly 5% of total annual passage between mid-October and mid-Noven Ibigrure 17a). Weekly passage varied somewhat during this period with some weets sage totals accounting for up to 20% of annual passage values. A second, inederas gnitude mode of passage occurred during December in most years with a single k accounting for nearly 50% of the annual passage estimate. Between BY 20622912, 75% of average annual passage occurred by mid-April, signifying Octoberugh April as the greatest period of migration. A third mode of similar magnitude tetsecond mode occurred during April and May with passage accounting for up to 45% efahnual run in some years. This could be characterized as an erroneous increasepring passage. Unmarked fall production fish exceeded the size-class for fall and therefore fell within the spring run category using LAD criteria. Between 200720112, on average, 4.3% of the marked fall production fish fell within the springen size-class using LAD criteria. Assumedly, a similar proportion of the unmarked fiveere added into the spring-run passage estimates as they could not be distinguishern naturally produced fish. Overall, spring Chinook passage was complete be the best was each year (Figure 17b).

O. mykiss passage estimates were generated using trap efficy estimates calculated using the Chinook-based trap efficiemcydel. Caution should be exercised when interpreting the following results as Chinoxod O. mykiss trap efficiency values likely differ, perhaps greatly. Irrespective of the magnitude of passage estimates based on Chinook efficiency trials, then ds in abundance remain plausible due to the standardization of effort and catch. Ilke Chinook,O. mykiss were not attributed to a fry or pre-smolt/smolt category and assage estimates with 90% CI's were calculated that included all size-classes life stages combined.

Annual passage estimates **to**rmykiss ranged between 56,798 and 151,694 juveniles for calendar years 2002-2012 (↑ = 116,**2**₹2 = 25.7%; Table 5e). Annual

effort values resulted in interpolations of betweenand 56% of annual passage estimates (at = 22%). The effect of annual effortQd width indicated a slightly greater spread of Cl's with decreasing effort, in geneFagure 18).

On average, weeklyO. mykiss passage was low (<5% on average) from April through July of each year with some variability.11 years of sampling only once did passage exceed 10% of annual passage during thesters. Weekly passage between July and August increased to peak values ranging fr% to nearly 25% (Figure 19a). Between 2002 and 2012, 75% of average annual passagurred by mid-August. Weekly passage generally declined between SepteraberOctober. Overall,O. mykiss passage was negligible between December and thewforlg February each year (Figure 19b).

Fry-Equivalen Chinook Production Estimates Juvenile Chinook passage values were standardized tory-equivalent production estimates for within- and between-year comparisons. As noted above, the various runs weare pled with oftentimes considerable variability in fry to pre-smolt/smottios over the 11-year sample period (Table 5a-5d). By multiplying 1.7 to all fish sterd pin the pre-smolt/smolt category (>45mm) within each run, annual Chinook production over the RBDD transect could be estimated. These standardized production estimates ld then be compared to adult escapement estimates calculated from the Califor interval Valley Chinook opulation Report (Azat 2013) or carcass survey data in tise of winter Chinook (USFWS 2006-2011 and 2013). Moreover, by comparing production the number of adult Chinook females each year (by run) and estimating fecundate from CNFH and Livingston Stone National Fish Hatchery (LSNFH) hatchery ption underviced, estimated recruits per female and egg-to-fry survival estimates were grated.

Fall Chinook fry-equivalent production estimates wheen 2002 and 2012 ranged from 7,554,574 to 30,624,209 (from 7,262,473, CAB.2%). Lower and upper 90% CI's were generated for each week, summed annually, award aged between 6,670,475 and 30,707,529 (Table 6a).

Adult fall Chinook escapement estimates above Rendernstem Sacramento River plus tributaries reported) estimated escapentheetween 12,908 and 458,772 (f) = 93,661) for the same years. Fall Chinook carcasses data collected by California Department of Fish and Wildlife (CDFW) providedual female:male sex ratio estimates averaging 0.46:0.54 (D. Killam, unpublishtata). A significant relationship between estimated number of females and fry-equival fall Chinook production estimates was detected? (= 0.53, df = 10?, = 0.01; Figure 20a). Recruits per female were calculated ranging from 89 to 1,515 (f) = 749) suming an average female fecundity value of 5,407, based on fall Chinookuspiag records from CNFH between 2008 and 2012 (K. Brown, unpublished data), restuit ean egg-to-fry survival estimate averaging 13.9% for fall Chinook (Table 6a).

Late-fall Chinook fry-equivalent production esttersabetween 2002 and 2012 ranged from 116,188 to 4,041,505 (↑ = 669,939, CN69=8%). Lower and upper 90% Cl's were generated for each week, summed annually, averaged between 222,044 and 1,236,432 (Table 6b).

Adult late-fall Chinook escapement estimates ab@@DD estimated escapement between 2,931 and 36,220 (\$\frac{1}{2}\$ = 9,108) for the samersy. Late-fall Chinook annual female:male sex ratio estimates relied on an asstimmpof the average ratio found for fall Chinook (i.e., 0.46:0.54). A significant trie haship between estimated number of females and fry-equivalent late-fall Chinook protitions estimates was detected (\$\frac{2}{2}\$ (\$\frac{1}{2}\$ = 10,P = 0.002; Figure 20b). Recruits per female were utanted ranging from 47 to 243 (\$\frac{1}{2}\$ = 131). Assuming an average female in the fall Chinook spawning records from CNFH between 2000 2012 (K. Brown, unpublished data) resulted in an egg-to-fry survival averaging 2.8% for late-fall Chinook (Table 6b).

Winter Chinook fry-equivalent production estimates ween 2002 and 2012 ranged from 996,621 to 8,943,194 (↑ = 4,152,547,=070.1%). Lower and upper 90% Cl's were generated for each week, summed annually, averaged between 2,265,220 and 6,124,494 (Table 6c).

Adult winter Chinook escapement estimates above IRIQUSFWS/CDFW carcass survey data; available attps://www.fws.gov/redbluff/he_reports.asp) estimated escapement between 824 and 17,205 (and 17,205) for the same syes Winter Chinook annual female: male sex ratio estimates were estimated dgrifhe annual carcass surveys (Table 6c). A highly significant relationship beetwoestimated number of females and fry-equivalent winter Chinook production estimates detected (r= 0.90, df = 10,P < 0.001; Figure 20c). Recruits per female were utalled ranging from 846 to 2,351 (all = 1,349). Annual female fecundity values were esterdated on winter Chinook spawning records from LSNFH between 2008 and 2005 EVVS Annual ropagation Reports; available attps://www.fws.gov/redbluff/he_reports.asp) and resulted in an egg-to-fry survival estimate averaging 26.4% for winter Chinotopable 6c).

Spring Chinook fry-equivalent production estimables ween 2002 and 2012 ranged from 207,793 to 747,026 (f) = 471,527, CV0.9%). Lower and upper 90% CI's were generated for each week, summed annually, award aged between 199,365 and 792,668 (Table 6d).

Adult spring Chinook escapement estimates above Rainstem Sacramento River plus tributaries reported) estimated escaperineetween 77 and 399 (f) = 195) for the same years. Spring Chinook annual female: steekeratio estimates relied on an assumption of the average ratio found for fall Chrik (i.e., 0.46:0.54). No significant relationship between estimated number of femalesdary-equivalent spring Chinook production estimates was detected (# 0.00, df = 10? = 0.971; Figure 20d). Recruits

per female were calculated ranging from 1,112 to 92, (f) = 3,122). Assuming an average female fecundity value of 5,078, based voeraging of 5 years of fall and latefall Chinook spawning records from CNFH and 10 systawinter Chinook spawning records from LSNFH, resulted in an egg-to-fry salveistimate averaging 61.5% for spring Chinook (Table 6d).

Green Sturgeon Data-Capture of young of the year sturgeon occurred warlly between calendar years 2002 and 2012, except in 82000 atch was highly variable, not normally distributed, and ranged between 0 and 31,700 year (median = 193; Table 7). Sturgeon sampled by rotary traps could be positived entified as Green Sturgeon in the field above total length of 46 mm. At this size, lateral sessitivere fully developed and could be counted to distinguish between White (penser transmontant) and Green Sturgeon (Moyle 2002). Of 2,912 sturgeon measured the field, 99.14% were less than 46 mm. In all years, except 2007 and 2008, subplemental and/or juvenile sturgeon rotary trap catch (up to 50% in some years re supplied to UC Davis for genetic research and all were determined to be @returgeon (See Israel et al. 2004; Israel and May 2010). We therefore assumed altogram captured in rotary traps were Green Sturgeon based on the results of genetic years. Moreover, Green Sturgeon were the only confirmed spawning Acipenserids send patr or above the RBDD transect between 2008 and 2012 during sturgeon spawning eyer (oytress et al. 2009-2013).

Green Sturgeon catch was primarily composed of metry emerged, post-exogenous feeding larvae with a 10-year median wasptotal length averaging 27.3 mm (SD = 0.8; Table 7). Sturgeon were sampled exert v18 and 188 mm, but those sampled above 40 mm were considered outliers (N = Table 7; Figure 21a).

The temporal pattern of Green Sturgeon capturescured, on average, between May 1 and August 28 of each year. Green Sturgepture trends indicated annual variability, but on average 50% were sampled by the of June each year and nearly 100% by the end of July (Figure 21b), with outlies, juveniles) captured in August, September and as late as November (e.g., 188 mrin 50)me years.

Relative abundance of Green Sturgeon was measured tch per estimated water volume sampled (CUV in ac-ft) through rotating cones and summed daily. Daily values were summed annually to produce each annual index of abundance. Absolute abundance estimates, via trap efficientials, could not be calculated due to low numbers of sturgeon sampled on a daily basis take fragile nature of newly emerged exogenous feeding larvae.

Green Sturgeon annual CUV was typically low aamged from 0.0 to 20.1 fish/ac-ft ($^{\circ}$ = 2.5 fish/ac-ft, SD = 5.9). taDvarere positively skewed and median annual CUV was 0.8 fish/ac-ft. Relative abundatise ibution data were highly influenced by samples collected in 2011 that equalate two orders of magnitude higher

than any other year's index (Figure 21c). Overvalitiability in CUV between years was relatively high as the CV was 236% for the eleveen-period (Table 7).

Lamprey Species DataCapture of multiple lamprey species occurred bastow water year (WY; October - September) 2003 and 20/03/. 2002 was excluded from analyses as less than 50% of the entire year wansplad. Lamprey species sampled included adult and juvenile acific Lamprey and acmuch lesser extent River Lamprey (Lampetra ayres) and acific Brook Lamprey (Lampetra pacifica) Unidentified lamprey ammocoetes and acific Lamprey (L) composed 99.80% all captures, 24% and 75%, respectively. River Lamprey and acific Brook Larenypcombined, composed the remaining 0.2% of all captures. Annual catch, the name relative abundance information for River and acific Brook Lamprey cape found in Appendix 1 (Tables A9 and A10) and are not discussed further due to very capture rates.

Annual catch of ammocoetes was relatively stable ranged between 385 and 1,415 individuals per year ($\hat{l} = 757$, median = 6757bJe 8a). The catch coefficient of variation for ammocoetes was 38.5%. Minimum Tlarofprey ammocoetes was 14 mm and maximum TL was 191. Over the eleven completely sampled, the average minimum and maximum TL's were 32 and 164 mm, retispely ($\hat{l} = 105$, SD = 4.7; Figure 22a).

Annual catch of L macropthalmia and a small frienct of adults was variable and ranged between 204 and 5,252 individuals per year (2,335, median = 2,747; Table 8b). The catch coefficient of variation for L was 5.3%. Minimum TL of L was 72 mm and maximum TL was 834. Over the eleven years leadynth a verage minimum and maximum TL's were 88 and 665 mm, respectively 1(50 = SD = 37.3; Figure 23a).

Lamprey captures occurred throughout the year betwo October and September. Ammocoete capture trends indicated annual variabilibut on average 25% were sampled by the end of January, 50% were sampled by the end of May and 100% by the end ptember (Figure 22b). Transformed L (macropthalmia and adult) capture trds indicated a different pattern of capture and annual variability compared to ammetes. On average, 5% were sampled through October, 50% were sampled throughbet through February, 90% by the beginning of Aprihval 100% by the end of September (Figure 23b).

Relative abundance of ammocoetes and L were merestuas CUV through individual rotary trap cones and summed daily. In produce were summed annually to produce each year's annual index of abundance. On the summed annually to employing mark-recapture methods could not be chatted due to the sporadic capture of adequate numbers of juveniles (e.g., > 1,000 individuals) that would be needed for mark-recapture trials. Moreover, emphasis was pethon conducting Chinook mark-recapture trials at times of pronounced lamprey ablance.

Ammocoete annual relative abundance ranged fro6nt8.11.7 fish/ac-ft (f = 6.8 fish/ac-ft, SD = 2.6; Figure 22c). Overall, amretecdata were normally distributed as median CUV was 6.5 fish/ac-ft, similar to the meanure. Variability in CUV between years was modest and the coefficient of variations w89% for the eleven-year period (Table 8a).

L annual relative abundance was generally hightern ammocoete relative abundance and ranged from 2.1 to 112.8 fish/acft(41.0 fish/ac-ft, SD = 34.7; Figure 23c). Overall, L data was slightly positively skeed and median CUV was 34.1 fish/ac-ft. Variability in CUV between years was moderated the coefficient of variation was 85% for the eleven-year period (Table 8b).

Abiotic Conditions—Tabular summaries of the abiotic conditions threatre encountered during each annual capture period wavenmarized for each run of salmon,O. mykiss, Green Sturgeon and Lamprey species. Tabular switesmassociated with each species annual captures are located briesa9a-9f and include: dates of capture, peak daily water temperature, peak dailyer discharge levels and mean daily turbidity values. A series of exploratory plotsmaring the above daily environmental data variables plus an index of moon illuminosity revgenerated for fry and pre-smolt Chinook daily passage estimates for visual analyster Chinook fry and presmolt/smolt plots are included in Appendix 2 (FigsuA1-A23) for reference.

Annual environmental covariate data for fall Chrikosalmon can be found in Table 9a. Results presented below describe data averagged 11 brood years. Fall Chinook were sampled over a period of 250 to 273 days peary($^{\circ}$ 1 = 264 days, SD = 7). Water temperatures ranged from 45 to 62 °F ($^{\circ}$ 1 = 55°F= \$DD). Sacramento River discharge ranged from 5,605 to 72,027 CFS ($^{\circ}$ 1 = 14,844 CF=\$5,\$PD2). Turbidity values ranged from 1.5 to 298.7 NTU ($^{\circ}$ 1 = 14.4 NTU, SD = 6.3).

Annual environmental covariate data for late-fallinook salmon can be found in Table 9b. Results presented below describe datagged over 11 brood years. Late-fall Chinook were sampled over a period of 27036 days per year (1 = 300 days, SD = 24). Water temperatures ranged from 46 to 62 °F (56°F, SD = 0.7). Sacramento River discharge ranged from 5,536 to 67,520 CFS (1 £012,5S, SD = 2,829). Turbidity values ranged from 1.4 to 272.0 NTU (1 = 11.3 ISTU; 6.2).

Annual environmental covariate data for winter the salmon can be found in Table 9c. Results presented below describe datagred over 11 brood years. Winter Chinook were sampled over a period of 207 to 27% squer year (1 = 250 days, SD = 20). Water temperatures ranged from 46 to 61 °F (1 = 75.5°D = 0.8). Sacramento River discharge ranged from 5,349 to 66,800 CFS (1 5210,95, SD = 3,767). Turbidity values ranged from 1.3 to 290.2 NTU (1 = 12.5 NSTU-, 5.1).

Annual environmental covariate data for springrobk salmon can be found in Table 9d. Results presented below describe datagged over 11 brood years. Spring Chinook were sampled over a period of 221 to 25% squar year ($\hat{}$ = 232 days, SD = 9). Water temperatures ranged from 46 to 62 °F ($\hat{}$ = 75.35D = 0.6). Sacramento River discharge ranged from 5,349 to 68,720 CFS ($\hat{}$ 7016,85, SD = 6,116). Turbidity values ranged from 1.4 to 305.9 NTU ($\hat{}$ = 16.0 NSDU; 7.0).

Annual environmental covariate data for mykiss can be found in Table 9e. Results presented below describe data averaged 60 earlendar years 0. mykiss were sampled over a period of 331 to 363 days per year (349 days, SD = 12). Water temperatures ranged from 46 to 63 °F ($^{\circ}$ = 56°F= \$08). Sacramento River discharge ranged from 5,333 to 67,610 CFS ($^{\circ}$ = 12,519 CF=\$3,551). Turbidity values ranged from 1.4 to 263.7 NTU ($^{\circ}$ = 11.4 NTU, SD = 4.1).

Annual environmental covariate data for Green § team can be found in Table 9f. Results presented below describe data averaged 6 fearlendar years. Green Sturgeon were sampled over a period of 56 to 15 fs table year (1 = 88 days, SD = 27). Water temperatures ranged from 55 to 61 °F (1 = 75 SCD = 0.9). Sacramento River discharge ranged from 9,639 to 23,538 CFS (1 & 31 C, FS, SD = 2,181). Turbidity values ranged from 2.4 to 93.9 NTU (1 = 8.5 NTU; SCD).

Due to the large amount of variability and lackarformal distribution, all environmental covariate CUV data analyses for Greaturgeon were performed using natural log transformed data (Sokal and Rohlf 1995) vironmental covariates were regressed against the natural log of daily CUV instead for Green Sturgeon in a linear regression setting (Figure 24). Maximum daily water perature was the only variable found to be significantly related to Green Sturger trative abundance, albeit the relationship explained ~5% of the variability ardurally relative abundance abundance of 15, P < 0.001).

Annual environmental covariate data for Lampspp. can be found in Table 9g. Results presented below describe data averaged of water years. Lamprey were sampled over a period of 358 to 364 days per year (362 days, SD = 2). Water temperatures ranged from 46 to 63 °F (1 = 56°F, 507). Sacramento River discharge ranged from 5,347 to 68,873 CFS (1 = 12,595 CFS4,5107). Turbidity values ranged from 1.2 to 306.8 NTU (1 = 11.9 NTU, SD = 4.4).

Due to the variability and lack of a normal distintion, all environmental covariate CUV data analyses for Lamprespp. were performed using natural log transformed data. Environmental covariates were regressed registine natural log of daily CUV data for Lampreyspp. in a linear and multiple regression setting. follow independent variables appear to contribute to predicting Lampsepp. relative abundance and were significantly related to abundance level (0.223, df = 1999), <0.001). Individual variable linear regression analyses indicated tuitle; water temperature, discharge,

and full moon illuminosity were correlated in dending order of magnitude (Figure 25). None of the covariates tested explained midnan ~16% of the variability associated with daily CUV data.

Discussion

Trap Efficiency Modeling-Over the past 11 years, annual mark-recaptural stri added 85 data points to the RBDD rotary trap efficity linear regression model (Figure 3). Explanation of the variability associated with efficiency and Q, in terms of the associated r-squared value, was reduced for that few years and then steadily increased in more recent years. The reduction dues, in part, to more precisted calculations over the initial model when diversion RBDD were not subtracted from daily river discharge values. Diversions wealther to be removed from the total discharge (Q) passing the transect as these data became a leitabreal-time starting in 2002.

The addition of a multitude of fry size-class striaver a variety of discharge levels greatly increased the accuracy of trap efficiens time ates. Fry size-class fish are the predominant size-class sampled at RBDD (i.e. arfallwinter Chinook) thereby making them the best representatives for use in mark-return trials. The original trap efficiency model developed by Martin et al. (200 fm) ployed primarily hatchery-raised smolts, as these fish were all that were available arge quantities and permitted for use in experiments to develop the initial model of the primary fish size-class sample RBDD rotary traps. Roper and Scarnecchia (1996) and Whitton et al. (2008) fosign difficant differences in trap efficiency when conducting paired mark-recapturials using hatchery and wild caught fish. The most recent years of RBDD data supposition cept.

While a simple linear regression model has worked over the years for our real-time data output needs, analysis of the data withing model, other possible covariates, and other more advanced modeling techniques has been ranted. Analysis incorporating additional potential explanatory variables was conducted using a generalized additive model technique (GAM; Hastige Taibshirani 1990). From this analysis, variables including turbidity, fish sized run, water temperature, weather condition, lunar phase, and river depth were exploin addition to%Q. The result was that only %Q and weather were found to be significant model lexipatory variables (= 0.68; df = 141,P < 0.01). The weather variable needs focused testing day ducting more mark-recapture trials under a variety of weathconditions to determine the applicability or mechanism of this variable. The Conditions to interpolate missed sample days.

At minimum, an update to the 142 trial linear trefficiency model (Figure 3) needs to be implemented for future passage estimateculations. The update will

include the removal of hatchery fish triaNs=(23) used as surrogates for natural stocks. Removal of all RBDD "gates in" mark-recapturestr(NsI=31) due to the cessation of RBDD dam operations since 2011 (NMFS 2009) isvalsanted.

The loss of annual maintenance and RBDD gate introgve perations at the rotary trap sample site (Figure 1) will allow the river other's geometry to change more frequently due to natural flow driven substrate traport mechanisms. RBDD operations of the past virtually "reset" the sampsite to facilitate pumping during the gates-out period and improve fish passage at the fadders during the gates-in period. As the sample site's channel configuration is abbotion fluctuate in the absence of dam operations, the overall effect could be differing to efficiency values in relation to flow compared to previous years' data. Annual mark-petrate trials will be needed to evaluate this phenomenon, which has been observed the uncontrolled channel sampling locations (e.g., Clear Creek; Greenwald explanation). The use of a GAM model may also be of benefit in this situation as it cobble constructed and employed annually to account for wide variation in annual trap efficiency values; albeit at the expense of being able to produce real-time data summaries.

A linear model that also removed the remaining-2002 trials (N=16) which estimated%Q in a less precise manner, would result in the mestresentative trap efficiency model. A post-RBDD wild Chinook model to type would incorporate 72 mark-recapture trials with a high degree of signafice (N=72, f = 0.669, F = 141.8, < 0.001) and be most representative of current sample onditions in terms of fish size-class and environmental conditions.

Chinook Capture Size Analyse@verall capture of Chinook salmon by RBDD rotary traps was heavily weighted towards fry sizes less than 40mm in fork length. All four runs' greatest proportion of fish were food in this size-class, albeit in a range of proportions from 24% for late-fall (Figure 5b) toeo 72% for winter run (Figure 6b). The capture size-class results fit well with the ratory strategies of 'stream' and 'ocean type' as noted in Moyle (2002) for late-fapting and fall/winter Chinook, respectively. The question of size selectivity apprture bias of rotary traps, a passive sampling gear (Hubert 1996), comes into questione row dealing with two very different migration strategies.

A two sample t-test was performed to evaluate threatential for size-class bias by comparing fry (fall and winter Chinook) size-class efficiency values (N=43) to presmolt/smolt (fall) trap efficiency values (± 10) between similar river discharge conditions. The t-test results did not indicateyasignificant difference between the mean efficiency values (± 10). Interestingly, the mean efficiency and standard deviation of the values were identife ± 10 . SD = 0.01) between groups. We recommend further study of the relation between pre-smolt/smolt size-class and trap efficiency to determine if edition or bias may exist between or among Chinook runs. Additional sampling effort to the lation of the capture

substantially more pre-smolts in the numbers requirfor efficiency trials in the Sacramento River to further test this potential satisfies molting salmonids also appear to succumb to stress induced mortality at a much greatete than fry, particularly in warmer water conditions due to relatively high respion levels, adding to the difficulty in testing this potential bias.

O. mykiss Life-Stage and GrowthCatch of O. mykiss was scattered throughout the year with multiple modes in abundance of predoately sub-yearling parr and silvery-parr occurring in early May and August.mykiss fry (<41 mm) made up 17.5% of the total O. mykiss catch in 2006 and was 2.4 standard deviations ftben11-year mean. In contrast, yolk-sac fry, made up only 9of the O. mykiss catch in 2006 and varied less than 1 standard deviation from the tearymean (Table 3). Elevated spring discharge resulted in poor sampling conditions whiteduced sampling effort, possibly scoured redds, and ultimately resulted in low over mykiss catch in 2006. Regardless of the cause of low catch rates, inlikely the migration patterns of O. mykiss changed in 2006 and the variability in age-classibility on was likely due to our sampling effort in that year.

The small percentage of mykiss smolts that showed signs of anadromy were generally migrating during March through June which consistent with outmigrating smolts found in Battle, Mill, and Deer Creeks (stommand Merrick 2012; Colby and Brown 2013). Interpretation of O. mykissdata collected at the RBDD was complicated as a robust resident (non-anadromous) populations exathroughout the Upper Sacramento River and its' tributaries. opulations anadromous and resident. mykisslife history forms are often sympatric and may intered (Zimmerman and Reeves 2000; Docker and Heath 2003), thereby regular abilities to separate the anadromous and non-anadromous components of the examples. Donahue and Null (2013) conducted research using otolith Strontiual (2011) and ratios to determine whether O. mykiss returning to a hatchery were progeny of anadromous sesident females. A similar analysis could be conducted using juve in important separation in spawn timing exists between anadromous and resident forms of pokiss coexisting within the Upper Sacramento River basin.

Linear regression equations developed using weighth data obtained from 0. mykiss showed a strong correlation between the two valies $(\hat{r}^2 = 0.942)$. The annual slope coefficient varied slightly between 2.858 and 52. Carlander (1969) suggested that slopes less than 3.0 might indicate a crowdestunted population. However, permit restrictions may have introduced bias intercresults as we were unable to anesthetize and weigh fish >200 mm thereby reducting slope of the regression compared to that of a complete analysis of the plation.

Sample Effort Influence on Passage Estimates ampling effort had profound effects on the precision of passage estimates ampling effort had profound effects on the precision of passage estimates ampling effort had profound effects on the precision of passage estimates ampling effort had profound effects on the precision of passage estimates ampling effort had profound effects on the precision of passage estimates ampling effort had profound effects on the precision of passage estimates ampling effort had profound effects on the precision of passage estimates ampling effort had profound effects on the precision of passage estimates ampling effort had profound effects on the precision of passage estimates ampling effort had profound effects on the precision of passage estimates ampling effort had profound effects on the precision of passage estimates amplified effects of the precision of the passage estimates amplified effects of the passage estimates and the passage estimates amplified effects of the passage estimates amplified effects of the passage estimates and the passage estimates amplified effects of the passage estimates amplified effects estimated effects of the passage estimates amplified effects of

14, 16, and 18). In general, as sampling effortrelessed, variance within weekly passage estimates increased and the width of comfort intervals subsequently increased. This effect was most prominent where the fives reduced during peak periods of outmigration or for long periods of tin(the 1 week) when sharp increases or decreases in fish abundance occurred. Unfortungated impling of outmigrant Chinook on a large river system such as the Sacramentor Rivinevariably subject to discharge events that are insurmountable for variable period stime.

Logistical factors including staffing and permitirestrictions can also have significant effects on the precision of estimates or example, a comparison of BY 2002 and BY 2005 winter Chinook passage with equival fort values (0.64) shows less precision of BY 2002 passage estimates over BY (270) to 5c). The basis of the relatively low effort in 2002 was capture restrictis prompted by ESA Section 10(a)(1)(A) NMFS permits for endangered winter Orbitin Moreover, staff levels were initially low as the program was reinstated aftenearly two-year hiatus and substantial sub-sampling measures (i.e., standardized sub-siample repeated weeks) had to be taken during record abundance levels. The netoeffees that sampling of fry, the predominant size-class of ocean type Chinook (MaQ@2; Figure 6a/b), was reduced in terms of the number of days each week and houfre ach night sampled during the peak emigration period. The overall net effect 120% wider Cl's about the 2002 estimate (i.e., less precision) compared to BY 2005s was due to interpolation of 45% of the fry data which comprised 90% of the 2000 ual estimate. In contrast, BY 2005 sampled 90% of the fry data which comprise \$200 the annual estimate. Effort was reduced 36% in 2005 as a result of winter stownereby sampling ceased for 3 straight weeks due to high river discharge level be effect of that lost sampling time in January did little to reduce the precision of the 2005 estimate as it was during a period when a mere fraction of a percent of totalssage for winter Chinook typically occurs (Figure 15). The impact to the BY 2605 hinookpassage estimate, on the other hand, was very wide Cl's about the estimate to the lowest effort of all 11 years during a critical time period for that rub stmigration (Table 5a, Figure 11).

In summary, the precision of passage estimates vary widely for numerous reasons within runs and among years. Inter-annyarilability in environmental conditions will always be a factor when attemptitogsample a riverine environment. Making good sampling decisions with knowledge efstpecies of interest and riverine conditions coupled with tenacity to sample critical code of outmigration (Volkhardt et al. 2007) are key to generating passage estimates an acceptable level of precision. Applying effort throughout each period of interesteds to be balanced between the value of data collected, an acceptable level of precision required of the data, the cost to attain the required precision, the impact samplimary have to a particular species, and the feasibility to appropriately sample the species interest.

Chinook Passage VariabilityJuvenile Chinook passage by one to four runsroccu every single day of the year in varying proportion RBDD. The sources and degree of

variability of juvenile Chinook passage are asrdeves the life-history and migration strategies of the runs they encompass. The magneton run-specific adult spawners appears to have the greatest influence on the oldermagnitude of juvenile Chinook passage and associated variability.

In recent decades, fall Chinook adults consistent/minated the Upper Sacramento River spawning salmon populations (AM) is 2006, Azat 2013). Throughout the past decade, we witnessed a 'coltaps the Sacramento River fall Chinook adult population and accordingly tracked in juvenile passage (Figure 10). Lindley et al. (2009) analyzed the freshwated marine components of fall Chinook outmigrants from BY 2004 and 2005 through treturn as adults in 2007 and 2008. They indicated BY 2004 and 2005 juveniles untered poor marine conditions upon ocean entry in the spring of 2005 and 2006 whitesulted in the marked decline in fall Chinook adult abundance starting in 2007.

Juvenile fall Chinook had the greatest mean anpassage value (14,774,923) of the four runs sampled at RBDD (Table 5a). Fallb@hipassage also exhibited the second smallest degree of variability with a C\\$60\2\%. Notably, fall Chinook annual production by the CNFH averages 12 million juven\bar{a}esimilar value to the mean passage value of unmarked fall Chinook all Chinook production fish from CNFH contributed heavily to the relative stability of thannual returning fall Chinook adult population (Williams 2006) and, consequently, julæpassage estimates over the past eleven years (i.e., basis of fall Chinook population

Temporal abundance patterns of fall Chinook intelidae primary passage of juveniles occurs between late December and Marchuffe 11a/b). Over half the run passed RBDD by mid-February, yet this varied once 11-year period by +/- one month. Fall run passage on the American River (William) Clear Creek (Earley et al. 2013a) and Stanislaus River (yper and Justice 2006) iliforaia generally subsides to low values by the end of March. This would be consisted the ocean type migration strategy as noted by Moyle (2002). The remaining fun smolts and subsequent 'jump' in abundance in April to May was a result of the mantked proportion of the CNFH production releases. Reduced variability in we plays age was observed in the final 20% of annual fall Chinook passage (Figure 11b).

Spring Chinook had the lowest average passage valla64,000 juveniles and the lowest CV of 45% (Table 5d). The low value of spathinook passage at RBDD can be attributed to a relatively small number of adults as vning primarily in Battle and Clear Creeks (Figure 1). Some extant populations appear habit Beegum Creek, a tributary to Cottonwood Creek (CDFG 2001), and in the main stacramento River (Killam 2009, Azat 2013). Of particular interest with respect the accuracy of spring Chinook

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⁷ Fall Chinook passages estimates do not includentiaeked proportion (0-25%) of CNFH production fishmarked fish of hatchery origin are included in annual passage estimates thredroccurrence is evidenced by increased passages primarily in May through June of each calendar year (Figure 11b).

juvenile passage at RBDD is the annual spawn tiofiagult spring Chinook and expected juvenile emergence timing. USFWS rotappoing operations on Battle and Clear Creeks between 2003 and 2012 have not predietmergence (i.e., through temperature unit analyses; Beacham and Murray 1990) sampled juvenile spring Chinook prior to November of each year. On averational first spring Chinook juvenile migrants from Battle and Clear Creeks were samplering the week of November 26 each year (USFWS, unpublished data). As a result; criteria used to identify juvenile spring Chinook at RBDD are noticeably inaccuratishas ampled prior to late November were not sampled upstream in primary protion areas at that time of year.

Simulating a removal of all LAD spring run betweetober 16 and November 25 of each year sampled would resultdecreased spring run passage estimates by 19%, on average (range 2.6 to 44.2%). The effects of reingoin correctly assigned fry annually did not indicate a statistically significant differce between annual estimates (pairled test, N = 11,P < 0.001). When incorrectly assigned fry are reendpythe slightly more accurate simulated spring Chinook annual passagues are main within the 90% CI of standard estimates.

Furthering the simulation by adding the weekly @betr through November spring Chinook estimated passage to the winter Chinooksage estimates (i.e., late spawning or emerging winter run most likely candidate; see FUVS 2013), had minimal effect on the magnitude of winter Chinook passage. The agree hacrease to winter Chinook passage was a mere 2.6% (range 0.6 to 8.8%) and passage remained within the 90% CI of the annual winter Chinook estimates li years.

Winter Chinook average annual juvenile passagetheasecond highest of the four runs estimated at 3,763,362 (Table 5c). The the annual estimates was 73.2%; higher than fall or spring, but moderately dispertseOverall, passage in years 2002. 2003, 2005, and 2006 surpassed the highest prewale of winter Chinook passage since juvenile monitoring began in 1995 (Gaines Madtin 2002). Similar to fall Chinook, winter Chinook adult escapement and subset juvenile passage began a marked decline in 2007 (Figure 16). Juvenile wildleinook have been determined to enter the ocean during March and April of each respriyeer et al. 2013) . Overall, it is believed that juvenile winter Chinook suffered theme fate as juvenile fall Chinook with poor marine conditions upon ocean entry in the ting of 2005 and 2006. Winter Chinook juvenile cohort replacement rates droppedlow 1.0 starting with BY 2007, similar to adult fall run as noted in Lindley et (2009). The lowest passage estimate between 2002 and 2012 for winter Chinook occurre 2011 at 848,976. Not until 2014 will we know if adult or juvenile cohort replacenterates will improve to a value of 1.0 or greater. Winter Chinook passage estimates betwBY 1999 to BY 2002 (Gaines and oytress 2003) indicate that replacement rates carry substantially and replacement rates of 3.0 or greater have been estimated betweenenile cohorts.

Late-fall Chinook passage averaged 447,711 jueseficit the 11-year period and exhibited the greatest amount of variability with CaV of 159.9%. Late-fall Chinook juvenile passage estimates are likely affected Asip criteria similar to spring Chinook in terms of potential for overestimation. The variable associated with weekly late-fall passage shows a decrease in median abundance byethicaning of June each year which may be more representative of actual late-farhergence. Additionally, as demonstrated by Figures 13 a/b, the late-fall mtgra starts abruptly unlike for fall and winter Chinook which follow a more bell-shaped parts in abundance (See Figures 11a/b and 15 a/b). It was highly likely that early ergent late-fall fry were, in fact, late emerging fall Chinook. Run specific genetic morinity (Banks et al. 2000, Banks and Jacobsen 2004) could assist in determining the ritage of the error in run assignment.

Sampling effort during mid-April to mid-May, tharty late-fall run emergent period, was also typically low in an effort to rectumpacts to CNFH fall Chinook production fish caught in rotary traps. Within prapredation of fry by CNFH production smolts could also negatively bias late-fall juvæpiloduction estimates. Sub-sampling of portions of the day and night (≤25% of each perioduction estimates. Sub-sampling staffing in some years which can reduce potentias During all other years, multiple sample days were typically sacrificed to allow prejakCNFH production fish to recede ultimately reducing the accuracy of late-fall pagesæstimates.

Fry-Equivalent Chinook Production Estimatesstimation and analyses of the productivity of salmon runs in the Upper SacrameRtover basin can provide valuable information to a variety of interests. ManagementCalifornia's complex water resources for agriculture, municipal, commerciald accological uses is an increasingly controversial and complex endeavor. Knowledgenefetffects of manipulating water storage and river processes on the productivity new Sacramento River fish populations can only benefit fishery and water operations maerogin an attempt to balance the competing demands on the system. Reducing uncetytasissociated with threatened and/or endangered fish population dynamics by enying knowledge of the abundance, migration timing, and variability of steopopulations over time can then inform the decision making processes guiding managed of water and fishery resources into the future.

Fall Chinook fry-equivalent juvenile productiodities (FEJI; Table 6a) indicate a significant and moderate correlation with fall Chrink escapement estimates (Figure 20a). Approximately 53% of the variation associateth fall FEJI 's was attributed to the estimated number of females in the system ab ReDD each year (Figure 20a). The CV of estimated fall run females was greater than 's indicating wide dispersion of contributors to the juvenile population over the en-year period. Conversely, the CV of FEJI 's was relatively low valued at 43%. Fruntbre, recruits per female and similarly egg-to-fry survival demonstrated moderlater waverage values of 749 and

13.9%, respectively, when compared to the estimatedues for winter Chinook (Table 6a).

As noted in Kocik and Taylor (1987), factors impiproduction are typically a combination of biotic and abiotic factors. The scens of variability relating to fall FEJIs are directly and indirectly related to adult abundae, but abundance alone does not explain the low CV in fall run juvenile productions.simple, albeit incorrect, conclusion might be that adult escapement of fall Chinook comes years exceeds the useable spawning area of the system (Bovee 1982, Connat. 2001) or optimal spawning efficiency (Wales and Coots 1955). Upon closemenation of the likely origin(s) of juvenile production, the data indicate substantial riability in the distribution of fall run adults between the mainstem Sacramento River aiteutaries, including Clear Creek and Battle Creek, between years. roportions oftoening adults within the mainstem and Battle Creek have demonstrated high degreexadability (Figure 26). The overwhelming return of fall run to Battle Creek2002 resulted in the lowest value of fall Chinook recruits per femal (= 89) which was outside two standard deviations of the average (Table 6a). The number of adults return to the CNFH clearly overwhelmed the capacity of Battle Creek to prodimeeniles. Sub-optimal wetted useable spawning area (Bovee 1982), red superintipos McNeil 1968, Heard 1978), and female stress resulting in egg retention (Nea953, Foerster 1968) were likely just some of the factors that reduced the overall produity of the 2002 fall Chinook adults returning to the Upper Sacramento River.

In years when estimates of fall Chinook productivere at their highest in terms of recruits/females (Table 6a), the proportions supring in the mainstem and combined tributaries were closest to 50:50. Further exantion indicates that when contributions from the Battle and Clear Creeks aucted for equal proportions (i.e., 25% each), peak values of ~1,500 recruits/femalesevestimated to have been produced resulting in the highest net spawning coeffice (Wales and Coots 1955). Optimal natural juvenile fall Chinook production ues in the Upper Sacramento River system could result under some conditions if intetion of restoration projects on Battle and Clear Creeks integrate with mitigation in the control of the control o the mainstem Sacramento River. The effect of extent hatchery fall Chinook production on Battle Creek irrespective of natufial production in the Sacramento and Chinook-bearing tributaries should be considered for the evaluation as was noted in Williams (2006). The effects of restoration of alereek appear to be providing production benefits on stream and basin wide scales nagement prerogatives and actions related to the CV affect both factors, to trying degrees, and decisions should be prioritized to attain optimal results for bothsheries and water operations.

Late-fall Chinook FEJIs indicated high variabile V = 170%; Table 6b), but a strong correlation with escapement estimate (0.67; Figure 20b). The magnitude of late-fall FEJIs were consistently an order of matgrde less than FEJIs of fall Chinook. One exception was 2002, which increased the Ctheoeleven-year period by 100%

(Table 6b). The fall and late-fall adult Chinoskapement values of 2001 and 2002 were high compared to the other 10 years of data(#2013). A large run of late spawning fall run may also have contributed to take number of juvenile fish falling within the late-fall size-class according to LAiDeria, but the adult estimate could have suffered similar inaccuracies in run assignmentrial/bility in CV values of anadromous fish was described by Rothchild and Dinardo (1987)eing inversely related to the number of years included within the time series byses. While 2002 appears to be an outlier in this data set, it is likely with moreages of data collection and analyses the CV associated with late-fall production would be more meaning with other runs of Chinook.

The stream-type migration strategy noted by Mo(2002) and our size classification method categorized the majority attel-fall outmigrants as smolts (ୀ = 62%) which inflated the late-fall FEJIs greatlytimes (Table 5b, Table 6b). Recruits per female and similarly egg-to-fry survival had lows@Md the lowest average values of 131 and 2.8%, respectively, in comparison to others (Table 6b). This was unexpected as this metric does not appear to appeal to a run that was sampled primarily as smolts (f) = 62%) over eleven years relativer, fry-equivalent calculations based on a static fry-to-smolt survival estimate 9% (Hallock undated) was unlikely to be an accurate constant for late-fall Chinook asats calculated from hatchery-based fall Chinook survival data. The fact that corriedas with adult escapement were determined to be significant and moderately strongs unexpected given the vagaries of sampling late-fall Chinook smolts and the usthefstatic 59% survival estimate inversely applied to the majority of the run sample Additionally, difficulties with performing carcass surveys for late-fall Chinook toulow visibility, winter flow events or logistical issues (Killam 2009 and 2012) tylpicasult in sub-optimal sampling conditions and, assumedly, would reduce the accurate adult estimate.

Overall, production of late-fall Chinook appears and the run has been characterized by some as vulnerable to extinct long/le et al. 2008, Katz et al. 2012). Greater attention to the relatively low abundance/els and juvenile rearing habitat needs of this genetically distinct run (Banks et 2000, Garza et al. 2007, Smith et al. 2009) with its unique over-summering, relatively dofreshwater residency (Randall et al. 1987) and large size-at-outmigration strate of late-fall inook have likely allowed them to persist in the Upper Sacramento River system as those upy a distinct ecological niche. Juvenile monitoring of this run could benefit great confidence in the accuracy of run assignment of juveniles was examined using nonal expension techniques (Harvey and Stroble 2013).

Comparisons between winter Chinook adults and julgeproduction began early using data generated by this monitoring projectartin et al. (2001) demonstrated a strong relationship with only 5 years of data. Thenual analyses of the winter FEJI and adult estimates continually indicated a strong tionship with the addition of each

year's data (See Gaines and oytress 2003, oytress Carrillo 2008, oytress and Carrillo 2012). The analysis of the most recentred for data continues to indicate a strong relationship between the two variables evaluates evaluated an order of magnitude.

Winter Chinook FEJIs indicated mild variability (= 67%; Table 6c) and a very strong level of significance and correlation withmale adult escapement estimates (r =0.90; Figure 20c). Intensive adult and juvenite intoring for this ESA listed endangered species coupled with superlative sangetion adultions, in most years, appears to have resulted in very high quality inflation regarding the status and trends in adult and juvenile population abundance.

Egg-to-fry survival estimates generated from arlnwiater Chinook data indicate a range of values between 15 and 49% (Table & fat)rs Aglance, this appeared counterintuitive based on the highly regulated Samento River system (e.g., flow and water temperatures) that typically exists during the interior Chinook spawning period. The average egg-to-fry survival estimate of 26% insiderably higher than that determined from other studies on acific salmonid = 15%; e.g., Wales and Coots 1955) but was consistent with highly regulated attiquaystems (Groot and Margolis 1991). A very low CV of 38% also appeared consistent a regulated system. Recruits per female, similarly, indicated a low CV of 36% three second highest average value of 1,349 (Table 6c).

Natural log transformed adult female estimated uneficed juvenile production and a significant relationship was determined accougator roughly half of the variability associated with egg-to-fry survival rates=(r0.51,df = 10,P = 0.012). Densities of winter Chinook spawners are much lower currentlarthin the years estimated following the completion of Shasta Dam (USFWS 2006)mpletion of the reengineered Anderson-Cottonwood Irrigation Distfish ladders in 2001 resulted in greater access and subsequently a greater conceintra spawners in the uppermost reaches accessible to anadromous fish (USFWS 20006-2Competition for optimal spawning habitat can result in lower juvenile produn if sub-optimal wetted useable spawning area (Bovee 1982), red superimposition New 1968, Heard 1978), and female stress resulting in egg retention (Neave31950erster 1968) occur to varying degrees. Low resolution carcass recovery data, (reach specific) indicate an abundance of spawners utilizing the uppermost @rimiles of the Sacramento River (USFWS 2006-2011) even as seemingly suitable habitabeen made available for approximately 20+ river miles downstream of thentenus at Keswick Dam (RM 302). Geist et al. (2002) studied physiochemical characties affecting redd site selection preferences by Chinook and different growth and elepment rates have been attributed to different segments within the same/eir (Wells and McNeil 1970). High resolution redd surveys or spawning area mappingpleging a GIS spatial analytical framework (Earley et al. 2013b) may shed light to tariability associated with winter Chinook spawning habitat over a variety of adult mathance levels. Analyses of these

types of data could result in less uncertainty other annual specific density dependent mechanisms affecting juvenile production and previtil rection for future restoration activities for winter Chinook.

Spring run Chinook FEJIs were the lowest of call fruns monitored and indicated the lowest variability (CV = 41%; Table 6d). Nationship with female adult escapement estimates was detected (0.00; Figure 20d) and may be attributed substantially to measurement error (Sokal and Robes). Estimates of recruits per female averaged 3,122 and the egg-to-fry survivatable averaged 61.5%. These values appear unreasonable outside of a hatchery environmend well above those found for other runs (this report) and other studies (e.g.ales and Coots 1955, Groot and Margolis 1991). Individual annual estimates varientlerately (CV= 70.8%) and nearly half appeared highly unlikely, with some valueseexting the number of eggs deposited by spawners (Table 6d).

Spring Chinook juvenile fish production estimate RBDD were the least accurate and currently constitute 2.1%, on average, of total Chinook production above RBDD. Mainstem Sacramento River spawner estimateged from a low of 0 to a high of 370 between 2002 and 2012. Annual indexes pingp Chinook adult abundance above RBDD during the same years constitute 2.70% extipated in the Sacramento River system (Azat 2013). Give nelatively sporadic and low adult abundance levels, vagaries of using LAD criter deam usal CNFH fall Chinook production releases with fractional mark rates, nedationship could be found between adult escapement and spring Chinook FEJIs where net ting to use methods to correct for these inaccuracies. The effects of inaccurating run assignment did not appear to affect the FEJIs of other runs (e.g., winter fall run) and therefore were not considered biologically significant. Genetic morning of fry in the fall after emergence from tributaries where emergence and migration detacollected (e.g., Earley et al. 2013a) may allow for more accurate estimation of the trontributions of this run to the Upper Sacramento River outmigrant population.

Green Sturgeon Capture DynamicsRotary traps were originally constructed to sample outmigrating salmonid smolts, but have beffective in sampling a variety of downstream migrating fish (Volkhardt et al. 200 Potary traps sampling at RBDD have been effective at monitoring temporal and spatients in relative abundance of Green Sturgeon since 1995 (Gaines and Martin 2002).

Annual adult Green Sturgeon aggregations were roless behind the RBDD when gates were lowered each spring (Brown 2007). Gsetergeon larvae were captured in 2012 (Table 7), the first year the RBDD gates wet dowered as it was replaced by a permanent pumping plant (NMFS 2009). Spawning determined to have occurred in multiple locations as fasts 20 river miles upstream of RBDD (Poytress 2009-2013). The location of the RBDD rotary traps has been irrone to be within the Green

Sturgeon spawning grounds as eggs were sampledthyingelow the RBDD and upstream of the RBDD traps in multiple years (Resystret al. 2009, 2010, 2012).

Total length distribution data from Green Sturgeconlections at RBDD indicate a narrow and consistent size-class of larvae (Figure). These data are consistent with laboratory-based studies conducted by Kynard e(2005) on the behavior of early life intervals of Klamath River Green Sturgeon. Their sdetermined that larvae migrated during two distinct periods (i.e., two-step migrant). The first migration of newly exogenous feeding larvae was determined to be attraIndispersion from production areas. The second migration (of juveniles) to vivetering areas occurred in the fall some 180 days after hatching, on average. Our youtrap data suggest we are sampling exclusively the initial redistribution of larvae from egg incubation and hatching areas.

Benthic D-net sampling conducted by Poytress.e(2010-2011) targeted the lowest portion of the water column (inverse of rotatraps) and consistently captured Green Sturgeon larvae of the same size-class ampdeal distribution pattern as rotary traps. D-net samples were collected between Magt early-August (See Figure 21b for corresponding RST data only) downstream of spawairegs in years 2008-2011; even as no larvae were collected by rotary traps in 2002 roae were sampled by both methods primarily in the thalweg and in river veities >/= 1.3 ft/sec. Conversely, zero juveniles were collected with benthic D-nets in a pilot syu(Poytress et al. 2013) targeting this life-stage and habitat type in thertithos during the fall period. Rotary traps have collected a few sporadic juveniles (cogtliers; Figure 21a) over the entire sample record of the project. These data indidated Green Sturgeon juveniles are no longer utilizing our sampling region or more likesing a different habitat type (Hayes et al. 1996). Accordingly, rotary traps appeable a relatively ineffective gear type for sampling the secondary juvenile sturgeon migration.

Protections afforded to ESA listed southern distipropulation segment of Green Sturgeon (since 2006), limited quantities of lar, valued the small size at capture have not allowed their drift distances (Auer and Bake02), rates (Braaten et al. 2008), or rotary trap efficiencies to be calculated for threitial dispersion migration of Sacramento River Green Sturgeon at RBDD. Redutivedance indices for Green Sturgeon were highly variable, typically low valued 1.0 fish/ac-ft sampled (Table 7), and contained one extraordinarily strong year-classigure 21c). As noted by Allen and Hightower (2010), variations in recruitment by orslepf magnitude between years is common among fish stocks. Moreover, strong and kweer classes greatly influence adult fish populations. Green sturgeon relative advance indices should not be interpreted as recruitment to the adult population but should be viewed as a production metric influencing recruitment (e.g., e.g.) year class strength). Alternately,

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⁸ Rotary traps generally require a minimum waterowith of 1.2 ft/sec to operate properly. D-netsnspaled velocities ranging from 1.3 – 6.6 ft/sec. RST sampled velocities ranging in 1.3 – 6.3 ft/sec.

Green Sturgeon larvae relative abundance indicestobe viewed as an indirect metric for adult spawning population densities stream of RBDD if genetic monitoring were conducted consistently (Israel and May 2010).

Lamprey Capture Dynamies Similar to Green Sturgeon, rotary trap sampliong Chinook salmon has provided the additional benefitapturing out-migrating lamprey ammocoetes and juveniles. Greater attention tostancestor of the earliest vertebrates (Moyle 2002) has recently been paidlteyUSFWS since it was petitioned for listing under the ESA in 2003 (Nawa et al. 2008) though not listed due to inadequate data on the species' range and thretate,USFWS has engaged in a strategy to collaborativelyconserve and restore Pacific Lamprey throughout the tive range. Through the formation and development of the Pacifiamprey Conservation Initiative, an assessment of Lamprey populations in Califdrais are cently been completed (Goodman and Reid 2012). The assessment noted thrat prey species had been extirpated from at least 55% of their historical threat north of Point Conception, CA by 1985. Long-term monitoring data sets including REDD rotary trap data, utilizing temporal and spatial distribution patterns as weed size-class and relative abundance levels of lamprey, can aid in the assessment arms envation of this ecologically vital species (Close et al. 2002).

Variability in annual size-class total length ritisations was typically minor for both lamprey life stages sampled (Figure 22a agdrei23a). Ammocoetes were slightly smaller than macropthalmia and slightly representable in their annual average length distributions valued at 110 mm TL (CV= 4.76%) le 8a). Pacific Lamprey macropthalmia were the dominant life stage samplered the median size at capture was consistently near 125 mm TL (CV= 1.6%; TalpleA8dbults, typically noted as outliers, were encountered in much lower frequenciend were considered upstream migrants inadvertently captured when the RBDD gartese lowered as they sought upstream passage around the partial migration bearri

Temporal distribution patterns indicated that ammonetes and macropthalmia migrate past RBDD year-round. Ammocoetes, on an energy americal ambient regularly throughout the year (Figure 22b), whereas macropthia moved, en masse, episodically between November and March (Figure) 23 these data are consistent with studies of macropthalmia in the Columbia River systems noted by Close et al. (1995) and Kostow (2002).

Relative abundance indices of ammocoetes (Fig2o) 2aried little between years and little overall when compared with macropthalm(Figure 23c). Macropthalmia abundance indices varied considerably between years between years between years abundance indices varied considerably between years leads be so. On average, macropthalmia relative abundance was six times that mmocoetes indicating metamorphosis and redistribution to different habits from those used for rearing by ammocoetes (Goodman and Reid 2012). Difference includes abundance CV's of the two life stages likely indicates differences attachability (Hubert and Fabrizio 2007)

or habitat use (Hayes et al. 1996), variable migratrigger effects, or variability in sampling effort that often occurred during periodsmacropthalmia migration.

Water Temperature and Juvenile Fish Dynami8sight variation within and among salmonid runs (including mykiss) and years was noted for water temperatures found at RBDD (Tables 9a-e). Nonetheless, Uppearsento River salmonids were subjected to a relatively wide 20 degree range after temperatures. Temperatures were recorded between 44 and 64 degrees with therage being 55 degrees each year. As summarized in Vogel and Marine (1991), thing of temperatures experienced by Chinook fry and pre-smolt/smoltship last 11 years of passage at RBDD have been within the optimal range of thertoledrances for survival.

Sacramento River water temperatures below Shasts Wick dams can be managed at certain times of the year under some ditions through discharge management to provide selective withdrawal at subgreed intakes (USBR 1991 & 1994, Vermeyen 1997). Ambient air temperatures typic by gulate river water temperatures during winter and early spring periods while stor by flood control operations are preeminent. The water temperatures recorded durthe last 11 years appear to have been favorable for extant spring run spawners, anothe so for fall and late-fall run Chinook and mykiss spawner and outmigrant populations.

The most vulnerable Chinook run to temperature rangement operations conducted by the USBR is winter Chinook (NMFS 20109)) perature management of the Sacramento River via Shasta/Keswick release the SBR for winter Chinook appeared to be effective during the last 11 years and enced by the relatively favorable and stable egg-to-fry survival estima (1886) be 6c). Moreover, temperature management of the upper 50 river miles of the Sarce ato River aimed at winter Chinook resulted in benefits to over-summering that Chinook pre-smolts and a relatively small proportion of fall Chinook smolts.

Temperature management during the summertime ainae winter Chinook may have indirectly favored the resident form of fmykiss. As noted by Lieberman et al. (2001), altering the thermal regime and foodbwatructure by way of temperature management likely affects the proportion of anadrouss to resident forms in large rivers. Lamprey species have likely benefitted from tempera management as temperatures for early life stages of lamprey in the last 11 years (Table 9g).

Green Sturgeon have likely benefitted from temptera management efforts aimed at winter Chinook spawning and production betalless comprehensively. Van Ennennaam et al. (2005) determined Green Sturgeggndevelopment temperatures to be optimal between 57.0 and 63.5° F. Mayfield Deth (2004) determined optimal temperatures for larval development to be between 50.5° and 66.2° F. Temperatures recorded at RBDD during larval capture periods and 58.3° F and were generally

within sub-optimal (lower end) to optimal rangesable 9f). A weak negative relationship between Green Sturgeon CPUV and wteteperatures was detected in our analysis indicating greater capture rates at the water temperatures (Figure 24d). The slightly sub-optimal temperatures might resultarvae migrating from incubation areas prematurely. Conversely, the optimal thermal ironment of the lab-based migration data from Kynard et al. (2005) resulted/ery similar migration timing between the lab and larval captures in rotary trapserms of days post hatch (Poytress et al. 2013). Sacramento River Green Sturgeonateanopear to be following their natural life-history migration patterns as opposted being coerced from their incubation areas due to sub-optimal water temperats at RBDD. This may not be true for larvae migrating some 20 miles upstream where effects of temperature management may have a more pronounced negatived Green Sturgeon larvae (Poytress et al. 2013). Temperature managemen Cloinook may also have the indirect negative effect of redirecting the spawgihabitat of Green Sturgeon adults by 20 river miles. A habitat comparison study on teleative value of the upper 20 river miles of the Sacramento River versus 20 lower rivibes of habitat currently benefitting Green Sturgeon adult spawners and from temperature management efforts should be conducted.

River Discharge, Turbidity, and Juvenile Fishr Digsa—Volkhardt et al. (2007) stated that "flow" (i.e., discharge) was a dominate tor in juvenile trapping operations. Trapping efficiency and migration rates are affected flow and the RBDD rotary trap passage data reflect these statements well. Explory plots demonstrating fry (Appendix 2, Figures A1-A11) and pre-smolt/smonth of the etfs of environmental variables on fish migration. Turbidity was plotted, but not laded in the final plots presented as the effects could not be deciphered from discharde the edity scale of analyses.

The effects of river discharge on turbidity and utlant fish passage are complex in the Upper Sacramento River where ocean and stretape Chinook of various size-classes (i.e., runs, life stages and ages) mightaits throughout the year. Decreases in discharge in the Shasta/Keswick dam regulated Streem to River, typical of late summer to early winter periods, appear to coinciwith relatively clear water conditions and low turbidity (e.g., ~ 1.5 NTU) BDRD. Fall or early winter freshets and winter rain-driven storm events result in highlyrizable increases in discharge levels and turbidity measures in terms of the magnitude and altion depending upon the source(s) of run-off.

A course scale analyses of fish passage and disvertange and turbidity measurements during storm events typically indiscate pattern that fish passage increases with simultaneous increases in both vote isa. Inspection of Chinook passage on a daily time step typically demonstrate a reductin fish passage a day prior to a storm or rain-event during periods of stable riverscharge. As storms produced increases in run-off or discharge from tributarpuints outside of the Shasta/Keswick

dam complex, mean daily turbidity typically incredsand fish passage began to increase. When storm related increases in discensing inished, turbidity diminished, but Chinook passage often increased greatly for 24 hours after the peak flow event.

One problem confounding the results of storm aisth spassage observations and analyses was that sampling during large storm rff/rdischarge events often ceased due to safety concerns, concerns for fish impactsionply due to the inability to sample the river when woody debris stop rotary traps from perating properly. In some years, storm events resulted in discharge levels too great ample effectively or damaged traps which resulted in numerous days or weeks armsled afterwards. The results are typically negative bias in passage estimates is day owing the peak discharge or concurrent turbidity events are un-sampled. Alterely, the direction of bias can be positive depending on time of year, interpolation ethods, sample effort during extended storm periods, or fish developmental stage

A fine scale, hourly analysis of fish passager, discharge and turbidity during storm events indicated a more intricate relations the variables. As a comparison, two separate storm events (December 5200 November 2012) were analyzed (Figure 27a/b). In 2005, 24-hour samples conducted prior to and after the peak flow period which was missed due to arbitisty to sample the river as it more than quintupled in discharge (i.e., 7,000 CFS to 000 CFS). During this storm event, sampling was conducted following the peak of rich planned 24-hour sample had to be cut short due to the huge influx of fry and strout sage that occurred during the turbidity increase (i.e., from 10's to 1,000's preduct) and the need to reduce the potential impact to listed winter Chinook.

During a November 2012 storm event, a different stagy was employed to collect data more effectively throughout the storm period for this event, we randomly sampled portions of the day and night in an attent manage the huge influx of fish anticipated to occur during the year's first storment. Between 11/17/12 and 11/23/12, the project was able to collect 7-randoms lelected samples that occurred throughout the first major river stage increase (Figure 27b). Samples were collected during increases and decreases in river stage. Programwere also collected prior to, during, and following a substantial increase in the first pre-smolt/smolt Chinook and juvenile lamprey fish passage increased expondential he peak period of fish capture occurred following the peak in river stage and degree and peak periods of turbidity measurements taken at RBDD. Captures at the beginning of the next stage increase (Figure 27b).

Overall, it appears that flow and turbidity are portant drivers for fish passage. The RBDD rotary trap data indicate that increased it often results in greater fish

passage than increases in river discharge or stategree which often occur as part of water management operations at Shasta Dam. The visuoisables generally increase sequentially with discharge increases followed utrybit increases (Figure 27a/b). Fish passage increases often coincide with the increase in turbidity which can often be sampled more effectively than increases in rives charge and may result in positive bias of juvenile fish passage estimates if the please lidity event is sampled compared to the peak flow event.

The importance of the first storm event of thelfat winter period cannot be overstated. Chinook smolt and juvenile lampreyspage increased exponentially and fry passage can be significant if first storms oracufall Chinook begin to emerge. Fishery and water operations managers should be awante importance of the first Sacramento River stage increases following the semand fall Sacramento River flow regulation period. The redistribution of winter and ver-summering fall and late-fall Chinook smolts, or more generally, all anadromouve juile fish migrating from the Upper Sacramento River to the lower river and Sacrato San-Joaquin Delta with the first storm events of each water year should be importated into management plans for Delta operations.

Moon Illuminosity and Juvenile Fish Dynamides noted in Hubert and Fabrizio (2007), species and life stages within species betwelffering behaviors and therefore catchability in response to light levels. Gained Martin (2002) determined that Chinook passage occurred primarily during noctuperalods except when turbidity levels and discharge increased with storm eventsther analyses of the effects of moon phase and ambient light levels in a statisticam ework may be warranted for Chinook salmon as trends were detected based one rotations. Rotary trap passage data indicated winter Chinook fry exhibit decrease exturnal passage levels during and around the full moon phase in the fall (Appendix Egures A1-A11). Pre-smolt/smolt winter Chinook appeared less influenced by nightetilight levels and much more influenced by changes in discharge levels (Appendix Grizgures A12-A23). A similar phenomenon was noted by Reimers (1971) for juvefaille Chinook in Edson Creek, Oregon. Alternately, more data concerning nighteticloud cover may further clarify the behavior associated with moon illuminosity and provided to the provided time of the provided time.

Spring, fall and late-fall Chinook fry exhibitedying degrees of decreased passage during full moon periods, albeit storms ædted hydrologic influx dominated peak migration periodsO. mykisselative abundance was not analyzed with respect to moon illuminosity. Lamprey CPUV regression analysticated a significant, but nearly imperceptible relationship (Figure 25a) likely doethe fact that lamprey are captured throughout the year under nearly all conditionsreen Sturgeon regression analysis

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⁹ Juvenile Green Sturgeon have been captured sp**orally** during the first flow events along with lar**ge**mbers of Pacific Lamprey juveniles and ammocoetes.

indicated no significant linear relationship betweenoon illuminosity and relative abundance (Figure 24a). Migration of age-0 Greengeon larvae has been determined to occur during nocturnal hours (Kynetcal. 2005) primarily between 21:00 and 02:00 using D-nets (Poytress et al. 2010) was presumed to be similar for rotary traps as periodic diel sampling events have collected sturgeon during daytime sample periods.

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Literature Cited

- Allen, M. S. and J.E. Hightower. 2010. Fish PtipoulDynamics:Mortality, Growth and Recruitment. Pages 43-769W.A. Hubert and M. C. Quist, editors. Inland Fishser Management, 3d edition. American Fisheries Society, Bethesda,yMand.
- Auer, N.A., and E.A. Baker. 2002. Duration antiofrifarval lake sturgeon in the Sturgeon River, Michigan. Journal of Applied Icollogy 18:557-564.
- Azat, J. 2013. GrandTab 2013.04.18. Californitrat Valley Chinook Population Database Report. California Department of Fish Wildlife. http://www.calfish.org/tabid/213/Default.aspx
- Banks, M.A., Rashbrook, V.K., Calvaetta, M.J.,, DeAn, and D. Hedgecock. 2000. Analysis of microsatellite DNA resolves geneticosture and diversity of chinook salmon (Oncorhynchus tshawytsc)him California's Central Valley. Canadian Journal of Fisheries and Aquatic Sciences 57:915-927.
- Banks M.A. and D.P. Jacobson. 2004. Which geneticens and GSI methods are more appropriate for defining marine distribution and gration of salmon? North Pacific Anadromous Fish Commission Technical Note 5, 39-42.
- Beacham, T.D. and C.B. Murray. 1990. Temperations, Size, and Development of Embryos and Alevins of Five Species of Pacifico Balkn Comparative Analysis. CTransactions of the American Fisheries Socie 9,61927-945.
- Borthwick, S. M. and R. R. Corwin. 2011. Fishaientent by Archimedes lifts and an internal helical pump at Red Bluff Research Pumplagt, Upper Sacramento River, California: February 1997 May 2000. Red Blufferen Pumping Plant Report Series, Volume 13. U.S. Bureau of Reclamation Ruff CA.
- Bovee KD. 1982. A guide to stream habitat analysisg the instream flow incremental methodology. Washington, DC:U.S. Fish and Wildlife ice, FWS/OBS-82/26.
- Braaten, P. J., Fuller, D.B., Holte, L.D., Lott, Riste, W., Brandt, T.F. and R.G. Legare. 2008. Drift Dynamics of Larval Pallid Sturgeon Simolvelnose Sturgeon in a Natural Side Channel of the Upper Missouri River, MontalNarth American Journal of Fisheries Management. 28:808-826.
- Brown, K. 2007. Evidence of spawning by greengestun, Acipenser medirostrish, the Upper Sacramento River, California. EnvironmeBitalbgy of Fishes 79:297-303.

- Brown, R. L. and S. Greene. 1992. Biological Ansent: Effects of the Central Valley Project and State Water Project Delta Operation Vanter-Run Chinook Salmon. California Department of Water Resources.
- California Department of Fish and Game (CDFG)1.29pring-run Chinook Salmon. Annual Report Prepared for the Fish and Game Cosimnis Habitat Conservation Division, Native Anadromous Fish and Watershed BrailMarch, 2001.
- Carlander, K. D. 1969. Handbook of freshwater figsbieology. Volume One. The Iowa State University Press, Ames.
- Close, D. A., M. S. Fitzpatrick, H. W. Li, B. Parkelatch, and G. James. 1995. Status report of the Pacific lamprey (ampetra tridentata) in the Columbia River Basin. (Project No. 94í026, Contract No. 95BI9067). PrepareU.S. Department of Energy, Bonneville Power Administration, Portlandegon. 35 pp.
- Close, D. A., M. S. Fitzpatrick, and H. W. Li. 2002 ecological and cultural importance of a species and risk of extinction, Pacific langer Fesheries 27(7):19-25.
- Colby, D. J., and M. R. Brown. 2013. Juvenileosaidhmonitoring in Battle Creek, California, November 2010 through June 2011. Us8.afnd Wildlife Service, Red Bluff Fish and Wildlife Office, Red Bluff, Califann
- Connor, W. P., Garcia, A. P., Connor, A. H., Gatto., Groves, P, A, and Chandler, J.A. 2001. Estimating the carrying capacity of the Sentativer for fall chinook salmon redds. Northwest Science. 75: 363-371.
- Docker, M. F., and D. D. Heath. 2003. Genetic avisage between sympatric anadromous steelhead and freshwater resident raintage tin British Columbia, Canada. Conservation Genetics 4:227–231.
- Donohoe, C. J., and R. Null. 2013. Migratory **h**istod maternal origin of rainbow trout (Oncorhynchus mykiss) returning to Colemantion Fish Hatchery in 2008. Institute of Marine Sciences, University of Canta Cruz, Santa Cruz, California.
- Earley, J. T., D. J. Colby, and M. R. Brown. 2019 enile salmonid monitoring in Clear Creek, California, from October 2010 through Septem 2011. U.S. Fish and Wildlife Service, Red Bluff Fish and Wildlife Office, Reft, California.
- Earley, L. A., S.L. Giovannetti, and M.R. Browh3l20Fall Chinook Salmon Redd Mapping for the Clear Creek Restoration Project),822012. U.S. Fish and Wildlife Service, Red Bluff Fish and Wildlife Office, Reft. 80alifornia.

- Fisher, F.W. 1992. (DRAFT) Chinook Salmon, Oncours Tshawytscha, Growth and Occurrence in the Sacramento-San Joaquin RiverSysthland Fisheries Division of California Department of Fish and Game. June, 1992.
- Foerster, R. E. 1968. The sockeye salmon orhynchus nerka Fisheries Research Board of Canada Bulletin 162.
- Gaines, P.D. and C. D. Martin. 2002. Abundand Seasonal, Spatial and Diel Distribution Patterns of Juvenile Salmonids Pasting Red Bluff Diversion Dam, Sacramento River. Red Bluff Research Pumping Reprirt Series, Volume 14, U.S. Fish and Wildlife Service, Red Bluff, CA.
- Gaines, P.D. and W.R. Poytress. 2003. Brood2002 winter Chinook juvenile production indices with comparisons to adult escane U.S. Fish and Wildlife Service report to California Bay-Delta Authori8an Francisco, CA.
- Gaines, P.D. and W.R. Poytress. 2004. Brood2962 winter Chinook juvenile production indices with comparisons to adult escaneant. U.S. Fish and Wildlife Service report to California Bay-Delta Authori8an Francisco, CA.
- Garza, J.C., Blankenship, S.M. Lemaire, C., and arier. 2007. Genetic population structure of Chinook salmonn (ncorhynchus tshawytschim California's Central Valley. Draft Final Report for CalFed Project "prefinensive Evaluation of Population Structure and Diversity for Central Walchinook Salmon". 82pp.
- Geist, D.R., T.P. Hanrahan, E.V. Arntzen, G.A. Malli, C.J. Murray, and Y.J. Chien. 2002. Physiochemical characteristics of the hypogrzone affect redd site selection by chum salmon and fall Chinook salmothen Columbia River. North American Journal of Fisheries Management 22: 10085.1
- Goodman, D.H. and S.B. Reid. 2012. Pacific Lar(Epreto)sphenus tridentati)s Assessment and Template for Conservation Measur@alifornia. U.S. Fish and Wildlife Service, Arcata, California. 117 pp.
- Greene, S. 1992. Daily fork-length table from aday. Frank Fisher, California Department of Fish and Game. California Department/Water Resources, Environmental Services Department, Sacramento.
- Greenwald, G. M., J.T. Earley, and M.R. Brown3.200venile salmonid monitoring in Clear Creek, California, from July 2001 to July 2005FWS Report. U.S. Fish and Wildlife Service, Red Bluff Fish and Wildlife OffRed Bluff, California.
- Groot, C. and L.Margolis. 1991. Pacific Salmfert-Listories. UBC Press, Vancouver, B.C.

- Hallock, R.J. Undated. The status of inlandtatabind factors adversely impacting salmon resources. Anadromous Fisheries Progratifion Department of Fish and Game, Red Bluff, CA.
- Hallock, R.J., W.F. Van Woert, and L. Shapoloo 1. 12n Evaluation of Stocking Hatchery-reared Steelhead Rainbow Tros all (no gairdnerii gairdne) iin the Sacramento River System. California Departments and Game. Fish Bulletin 114. 74 p.
- Harvey, B. and C. Stroble. 2013 Comparison oftigerersus Delta Model Length-at-Daterun assignments for juvenile Chinook salmostate and federal south Delta salvage facilities. California Department of Waterources. Submitted to Interagency Ecological Program for the San Frantag/Delta Estuary. Technical Report 88, March 2013.
- Hastie, T.J. and Tibshirani, R.J (1999) eralized Additive Modelsondon: Chapman and Hall.
- Hayes. D. B., C. Paolo Ferreri, and W. M. Tay 1966.1Active Fish Capture Methods. Pages 193-2260 B.R. Murphy and D. W. Willis, editors. Fisheries Interior, 2nd edition. American Fisheries Society, Bethesday 14ad.
- Heard. W. R. 1978. Probable case of streambedseveling-1967 pink salmon, Oncorhynchus gorbuschspawners and survival of their progeny in SaStrerek, southeastern Alaska. U.S. National Marine Fisherevice Fishery Bulletin 76:569-582.
- Hubert, W. A. 1996. Passive capture technique ges 157-1928 B. R. Murphy and D. W. Willis, editors. Fisheries technique ged edition. American Fisheries Society, Bethesda, Maryland.
- Hubert, W. A. and M.C. Fabrizio. 2007. Relativendance and catch per unit effort. Pages 279-326 C.S. Guy and M.L. Brown, editors. Analysis alertimetation of freshwater fisheries data. American Fisheries Sp.cBethesda, Maryland.
- Israel, J.A., J.F. Cordes, M.A. Blumberg, and B. 20204. Geographic patterns of genetic differentiation among collections of gressurgeon. North American Journal of Fisheries Management 24:922-931.
- Israel, J.A. and B. May. 2010. Indirect genestiionates of breeding population size in the polyploidy green sturgeor (ipenser medirostr) Molecular Ecology 19, 1058-1070.

- Johnson, R. R. D.C. Weigand and F. W. Fisher. 12822 of growth data to determine the spatial and temporal distribution of four rups juvenile chinook salmon in the Sacramento River, California. Report No. AFF182R25. U. S. Fish and Wildlife Service, Northern Central Valley Fishery Resouffice ORed Bluff, CA.
- Johnson, R. R. and C. D Martin. 1997. Abundantcessonal, spatial and diel distribution patterns of juvenile salmonids passingd Bluff Diversion Dam, Sacramento River, July 1994 June 1995. RedRestarch Pumping Plant Report Series, Volume 2. U. S. Fish and Wildlife Seriese. Bluff, CA.
- Johnson M. R. and K. Merrick. 2012. Juvenile daidh Monitoring Using Rotary Screw Traps in Deer Creek and Mill Creek, Tehama Co Catifornia, Summary Report: 1994-2010. RBFO Technical Report No. 04-2012.
- Katz, J., Moyle, P. B., Quinones, R.M., IsraeladdAS.E. Purdy. 2012. Impending extinction of salmon, steelhead and trout (Salmærið) in California. Environmental Biology of Fish. Published online January 2012.
- Killam, D. 2009. Chinook Salmon Populationship Utpper Sacramento River Basin 2008. Revised 1-11-2010. Northern Region-Department Fish and Game, Sacramento River Salmon and Steelhead Assessmejet: Prechnical Report No. 09-1.
- Killam, D. 2012. Chinook Salmon Populationship Utpper Sacramento River Basin 2011. Northern Region-Department of Fish and Gabaeramento River Salmon and Steelhead Assessment Project Technical Report 3N 2012.
- Kocik, J.F. and W.W. Taylor. 1987. Effect of Fidl Winter Instream Flow on Year-Class Strength of Pacific Salmon Evolutionarily Adapte Etarly Fry Outmigration: A Great Lakes Perspective. American Fisheries Society Styling 1:430-440.
- Kostow, K. 2002. Oregon lamprey: natural histoatus and analysis of management issues. Oregon Department of Fish and Wildlifey albs, Oregon. 112 pp.
- Kynard, B., E. Parker, and T. Parker. 2005. Burharvearly life intervals of Klamath River green sturgeor cipenser medirostrisvith a note on body color. Environmental Biology of Fishes 72:85-97.
- Lieberman, D. M., M. J. Horn, S. Duffy. 2001ects for a temperature control device on nutrients, POM, and plankton in the tailwaters by Shasta Lake, California. Hydrobiologia 452:191–202.
- Lindley, S. T., C. B. Grimes, M. S. Mohr, W. & Stein, J. T. Anderson, L.W. Botsford, , D. L. Bottom, C. A. Busack, Colker, J. Ferguson, J. C. Garza,

- A. M. Grover, D. G. Hankin, R. G. Kope, P. WohaksLow, R. B. MacFarlane, K. Moore, M. Palmer-Zwahlen, F. B. Schwing, th, Schrittracy, R. Webb, B. K. Wells, T. H. Williams. 2009. What cause Sacramento River Fall Chinook stock collapse? Pre-publication report to the Pacifishery Management Council.
- Martin, C.D., P.D. Gaines and R.R. Johnson. **253** imating the abundance of Sacramento River juvenile winter Chinook salmoln witmparisons to adult escapement. Red Bluff Research Pumping Plant **Repo**es, Volume 5. U.S. Fish and Wildlife Service, Red Bluff, CA.
- Mayfield, R.B. and J.J. Cech. 2004. Temperaturets of green sturgeon bioenergetics. Transactions of the American Fisheries Society 9633970.
- McNeil, W. J. 1968. Migration and distribution porfix salmon spawners in Sashin Creek in 1965, and survival of their progeny. U.S. Firsch Wildlife Service Fishery Bulletin 66:575-586.
- Meeuwig, M. H., J. M. Bayer, and J. G. Seelye.. **2670** fcts of temperature on survival and development of early life stage Pacific and temperature on survival of the American Fisheries Society 134:19-27.
- Moffett, J.W. 1949. The First Four Years of **Kinlg**non Maintenance Below Shasta Dam, Sacramento River, California, California **Diepent** of Fish and Game 35(2): 77-102.
- Moyle, P. B. 2002. Inland fishes of California press. Berkeley, California.
- Moyle, P.B., J.A. Israel, and S.E. Purdy. Sabbeethead, and Trout in California in California, Status of an Emblematic Fauna. Report missioned by California Trout, 2008. Center for Watershed Sciences, Unityeof California, Davis. Davis, CA.
- Mundie, J.H. and R.E. Traber. 1983. Movements of salmor Onchorhynchus kisutch fingerlings in a stream following marking with talvistain. Canadian Journal of Fisheries and Aquatic Science 40:1318-1319.
- National Marine Fisheries Service (NMFS). 2009o@cal Opinion on the Long-term Central Valley Project and State Water Project @tipens Criteria and Plan.

 NOAA (National Oceanic and Atmospheric Administrational Marine Fisheries Service, Southwest Fisheries Service Central Beach, California.
- Nawa, R. K., J. E. Vaile, P. Lind, T. M. K NadanāndicKay, C. Elkins, B. Bakke, J. Miller, W.Wood, K. Beardslee, and D. Wales. 2003 etition for rules to list:

- Pacific lamprey (Lampetratridentata); river lamprey (Lampetra ayreşi western brook lamprey (Lampetra richardso); iand Kern brook lamprey (mpetra hubbsi) as threatened or endangered under the Endang@pdcies Act. January 23, 2003.
- Neave, F. 1953. Principles affecting the size in and chum salmon populations in British Columbia. Journal of Fisheries Researe Industry Canada. 9:450-491.
- Pope, K. L., C. G. Kruse. 2007. Condition. Pa@est7412n C. S. Guy and M. L. Brown, editors. Analysis and interpretation of freshwaftesheries data. American Fisheries Society, Bethesda, Maryland.
- Poytress, W.R., and F. D. Carrillo. 2008. Broad 2006 winter Chinook juvenile production indices with comparisons to juvenile production estimates derived from adult escapement. Report of U.S. Fish and Wil Stepvice report to California Bay-Delta Authority and California Department of Fishd & Game, Sacramento, CA.
- Poytress, W.R., and F. D. Carrillo. 2012. Broad 2@10 winter Chinook juvenile production indices with comparisons to juvenile procession estimates derived from adult escapement. Report of U.S. Fish and Wildlifevice report to California Department of Fish and Game and US Bureau of Retitamn
- Poytress, W.R., J.J. Gruber, D.A. Trachtenbarg J. And Van Eenennaam. 2009. 2008 Upper Sacramento River Green Sturgeon Spawning at labid Larval Migration Surveys. Annual Report of U.S. Fish and Wildling to US Bureau of Reclamation, Red Bluff, CA.
- Poytress, W.R., J.J. Gruber, and J.P. Van Eenenn 2016. 2009 Upper Sacramento River Green Sturgeon Spawning Habitat and Larvarally Surveys. Annual Report of U.S. Fish and Wildlife Service to U. & Bull Buof Reclamation, Red Bluff, CA.
- Poytress, W.R., J.J. Gruber, and J.P. Van Eenen 22021 Ph. 2010 Upper Sacramento River Green Sturgeon Spawning Habitat and Larvaral Vibra Surveys. Annual Report of U.S. Fish and Wildlife Service to U.S. Reclamation, Red Bluff, CA.
- Poytress, W.R., and F. D. Carrillo. 2012. Bread-2010 winter Chinook juvenile production indices with comparisons to juvenile production estimates derived from adult escapement. Report of U.S. Fish and Wildervice report to California Department of Fish and Game and US Bureau of Retitation
- Poytress, W.R., J.J. Gruber, and J.P. Van Eenenn 2012. 2011 Upper Sacramento River Green Sturgeon Spawning Habitat and Larvar William Surveys. Annual Report of U.S. Fish and Wildlife Service to U.S. Bure Reof amation, Red Bluff, CA.

- Poytress, W.R., J.J. Gruber, C.E. Praetorius, Rantah Eenennaam. 2013. 2012 Upper Sacramento River Green Sturgeon Spawning Habitah aung-of-the-Year Migration Surveys. Annual Report of U.S. Fish aindiffer Service to U.S. Bureau of Reclamation, Red Bluff, CA.
- Pyper, B. and C. Justice. 2006. Analyses of tracew trap sampling of migrating juvenile Chinook salmon in the Stanislaus River 6-12005. Cramer Fish Sciences, Gresham, Oregon.
- Pyper, B., T. Garrison., S. Cramer, P.L. Brandes. Jacobsen., and M. A. Banks. 2013. Absolute abundance estimates of juvenile spring-and winter-run Chinook salmon at Chipps Island. Cramer Fish Sciences Techeipart Ror U.S. Fish and Wildlife Service, Lodi, CA. 89 pp.
- Randall, R. G., Healey, M.C., and J.B. Demps@r7.. 1/9ariability in Length of Freshwater Residence of Salmon, Trout, and Chameri& Fisheries Society Symposium 1:27-41.
- Reimers, P.E. 1971. The Length of Residence entitle. Fall Chinook Salmon in Sixes River, Oregon. Doctoral Thesis submitted to Oregon University.
- Roper, B and D. L. Scarnecchia. 1996. A comparistrap efficiencies for wild and hatchery age-0 Chinook salmon. North Americannal of Fisheries Management 16:214-217.
- Rothchild, B. J. and G.T. DiNardo. 1987. Comparts Recruitment Variability and Life History Data among Marine and Anadromous Fishesser Rean Fisheries Society Symposium 1:531-546.
- Smith, C.T., LaGranve, A.R., and W. R. Ardreroipe Cation with M.A. Banks and D.P. Jacobsen. 2009. Run Composition of Chinook salah Rued Bluff Diversion Dam during gates-in operations: A comparison of pheroity and genetic assignment to run type. U.S. Fish and Wildlife Service, Abern Fish Technology Center, Longview, WA. CY 2007 Report prepared for U.Sea Burf Reclamation-Mid Pacific Region, Red Bluff, CA.
- Snider, B., B. Reavis, and S. Hamelburg, S. **Srbtill**, and E. Kohler. 1997. 1996 Upper Sacramento River winter-run Chinook salmon esca**ptersuer**vey. California Department of Fish and Game, Environmental Ser**Direis**ion, Sacramento, CA.
- Sokal, R. R. and F. J. Rohlf. 1995. Biometryritheiples and practice of statistics in biological research, dedition. W. H. Freeman and Company.

- United States Bureau of Reclamation. 1991. Phanneiport and final environmental statement: Shasta Outflow Temperature Control. U,SMBiR-Pacific Region. Shasta County, California.
- United States Bureau of Reclamation. 1994. SæmænBasin Fish Habitat Improvement Study Final Environmental AssessmenBBR, Mid-Pacific Region.
- United States Fish and Wildlife Service (USFW99)5. 1Working Paper on Restoration Needs. Habitat Restoration Actions to Double Natt Production of Anadromous Fish in the Central Valley of California, Vol. 2ti 6n 9. May, 1995. Prepared for the US Fish and Wildlife Service under the direction Anadromous Fish Restoration Program Core Group. Stockton, CA.
- United States Fish and Wildlife Service (USFW95)7. 1Comprehensive Assessment and Monitoring Program (CAMP) Implementation Plan. Mante 1997. Prepared by Central Valley Fish and Wildlife Restoration Program (CAMP) Experience of the Program of the P
- United States Fish and Wildlife Service (USFWS). 279 in all Restoration Plan for the Anadromous Fish Restoration Program. A plan toeins natural production of anadromous fish in the Central Valley of Califor in pared for the Secretary of the Interior by the United States Fish and Wildlife where with the assistance from the Anadromous Fish and Restoration Program Core Quoder authority of the Central Valley Project Improvement Act.
- United States Fish and Wildlife Service (USFW05)6. 2Upper Sacramento River winter Chinook salmon carcass survey 2005 annual republif WS, Red Bluff Fish and Wildlife Office, Red Bluff, California.
- United States Fish and Wildlife Service (USFW05)7. 2Upper Sacramento River winter Chinook salmon carcass survey 2006 annual republif FWS, Red Bluff Fish and Wildlife Office, Red Bluff, California.
- United States Fish and Wildlife Service (USFW05)8. 2Upper Sacramento River winter Chinook salmon carcass survey 2007 annual republif WS, Red Bluff Fish and Wildlife Office, Red Bluff, California.
- United States Fish and Wildlife Service (USFW050)9. 2Upper Sacramento River winter Chinook salmon carcass survey 2008 annual republif WS, Red Bluff Fish and Wildlife Office, Red Bluff, California.

- United States Fish and Wildlife Service (USFW03)0. 2Upper Sacramento River winter Chinook salmon carcass survey 2009 annual republif WS, Red Bluff Fish and Wildlife Office, Red Bluff, California.
- United States Fish and Wildlife Service (USFW03)1. 2Upper Sacramento River winter Chinook salmon carcass survey 2010 annual republic WS, Red Bluff Fish and Wildlife Office, Red Bluff, California.
- United States Fish and Wildlife Service (USFW03)3. 2Upper Sacramento River winter Chinook salmon carcass survey 2012 annual reputations, Red Bluff Fish and Wildlife Office, Red Bluff, California.
- Van Eenennaam, J.P., J. Linares-Casenave, Xabers J. Doroshov. 2005. Effect of incubation temperature on green sturgeon embry Asipenser medirostris Environmental Biology of Fishes 72:145-154.
- Vermeyn, T. B. 1997. Use of Temperature Control Reservoir Release Water Temperatures. Report R-97-09, United States artment of the Interior, Bureau of Reclamation. Water Resources Reseabdratery, Technical Services Center. Denver, Colorado.
- Vogel, D.A. and K.R. Marine. 1991. Guide to Usperamento River Chinook salmon life history. CH2M Hill for the U.S. Bureau oflateation Central Valley Project, Redding, CA.
- Volkhardt, G. C., S.L. Johnson, B.A. Miller, Tckelson, and D. E. Seiler. 2007. Rotary screw traps and inclined plane screen traps. Pages266 in D. H. Johnson, B. M. Shrier, J.S. O'Neil, J. A. Knutzen, X. AugerAt, O'Neil and T. N. Pearsons. Salmonid field protocols handbook: techniques fascessing status and trends in salmon and trout populations. American Fisheriesi ty, Bethesda, Maryland.
- Wales, J.H., and M. Coots. 1955. Efficiency of cliki salmon spawning in Fall Creek, California. Transactions of American Fisherie & So & 4:137-149.
- Wells, R. A. and W. J. McNeil. 1970. Effect of tryuzf spawning bed on growth and development of pink salmon embryos and alevins. Fish and Wildlife Service Special Scientific Report Fisheries 616.
- Whitton, K. S., D. J. Colby, J. M. Newton, and Markwn. 2008. Juvenile salmonid monitoring in Battle Creek, California, Novemb@02 through June 2008. USFWS Report. U.S. Fish and Wildlife Service, Red Blstffand Wildlife Office, Red Bluff, California.

- Williams, J. G. 2006. Central Valley Salmonrspective on Chinook and Steelhead in the Central Valley of California. San Francistua Esand Watershed Science. Volume 4, Issue 3, Article 2.
- Zabel, R. W. and S. Achord. 2004. Relating size emiles to survival within and among populations of Chinook salmon. Ecology \$850. 795-806.
- Zimmerman C.E., and G. H., Reeves. 2000. Poppusatucture of sympatric anadromous and nonanadromous Oncorhynchus mykkislence from spawning surveys and otolith microchemistry. Canadian Jolunfiæ isheries and Aquatic Sciences 57:2152–2162.

Tables

Table 1. Summary of annual RBDD rotary trap satisfied by run and species for the period April 2002 through September 2013, by broged (BY).

BY	Fall	Late-Fall	Winter	Spring	O. mykiss
2002	0.76	0.57	0.64	0.75	0.53
2003	0.81	0.76	0.81	0.81	0.76
2004	0.85	0.88	0.84	0.85	0.83
2005	0.56	0.73	0.64	0.57	0.83
2006	0.90	0.70	0.83	0.89	0.59
2007	0.88	0.90	0.89	0.89	0.91
2008	0.79	0.89	0.87	0.85	0.89
2009	0.84	0.72	0.75	0.79	0.76
2010	0.75	0.86	0.81	0.77	0.85
2011	0.87	0.77	0.82	0.86	0.76
2012	0.85	0.89	0.89	0.86	0.86
Min	0.56	0.57	0.64	0.57	0.53
Max	0.90	0.90	0.89	0.89	0.91
Mean	0.81	0.79	0.80	0.81	0.78
SD	0.094	0.104	0.088	0.091	0.122
CV	11.7%	13.2%	10.9%	11.3%	15.6%

Table 2. Summary of mark-recapture experiments ducted by RBDD rotary trap project between 2002 2013. Summaries include trap effort data, fish release and recapture up sizes (N) and mean fork lengths (FL), percentage of rivecturing sampled (%Q) and estimated trap efficiency for each trial respective. Model data below each trial period indictates model was employed, total trials incorporated into model and linear respective.

		# Traps	Traps		Releas	e Group	Recap	ture Group	_	
Date	Run	Sampling	Modified	RBDD Gates	N	FL (mm)	N	FL (mm)	%Q	%TE
6/26/2002	Fal ¹	4	Yes	Lowered	805	68.7	8	61.3	1.58	0.99
8/6/2002	Falĺ	4	Yes	Lowered	743	69.7	16	80.2	1.66	2.15
8/20/2002	Falĺ	3	Yes	Lowered	340	76.5	7	77.7	1.41	2.06
Model	Employed	#Trials	Slope	Intercept	Р	R ²				
7/1/2002 -	6/30/2003	61	0.00792	0.00003205	<0.0001	0.394				

		# Traps	Traps		Releas	se Group	Recap	ture Group	_	
Date	Run	Sampling	Modified	RBDD Gates	N	FL (mm)	N	FL (mm)	%Q	%TE
1/28/2003	Fall	4	Yes	Raised	5,143	36.8	33	37.0	0.75	0.64
2/5/2003	Fall	4	Yes	Raised	2,942	36.7	10	37.9	1.36	0.34
2/10/2003	Fall	4	Yes	Raised	3,106	37.8	29	37.9	1.59	0.93
2/21/2003	Fall	3	Yes	Raised	3,256	37.4	15	37.3	0.72	0.46
2/26/2003	Fall	4	Yes	Raised	2,019	37.0	22	37.2	1.14	1.09
3/1/2003	Fall	4	No	Raised	1,456	37.0	31	37.0	3.31	2.13
3/4/2003	Fall	4	No	Raised	1,168	37.1	28	37.4	3.76	2.40
3/7/2003	Fall	4	No	Raised	1,053	37.4	22	36.6	3.58	2.09
3/20/2003	Fall	3	No	Raised	1,067	38.2	17	38.3	2.83	1.59
9/2/2003	Winter	4	No	Lowered	1,119	37.1	14	36.1	2.03	1.25
9/5/2003	Winter	3	No	Lowered	1,283	36.7	26	37.2	2.52	2.03
9/8/2003	Winter	3	No	Lowered	1,197	37.3	30	37.1	2.57	2.51
9/23/2003	Winter	3	No	Raised	1,012	35.5	18	35.6	2.20	1.78

9/27/2003	Winter	4	No	Raised	1,017	36.9	28	36.6	2.93	2.75
10/1/2003	Winter	4	No	Raised	1,064	37.6	20	36.7	3.09	1.88
10/6/2003	Winter	4	No	Raised	999	37.2	22	36.8	2.82	2.20
10/10/2003	Winter	4	No	Raised	1,017	38.1	16	38.3	3.06	1.57
10/15/2003	Winter	4	No	Raised	1,209	38.0	26	37.6	2.98	2.15
Model	Employed	#Trials	Slope	Intercept	Р	R ²				
7/1/2003 -	6/30/2004	79	0.00752	0.00046251	<0.0001	0.426				

-		# Traps	Traps		Releas	e Group	Recapt	ure Group		
Date	Run	Sampling	Modified	RBDD Gates	N	FL (mm)	N	FL (mm)	%Q	%TE
1/18/2004	Fall	4	Yes	Raised	2,074	37.1	26	37.1	1.52	1.25
1/24/2004	Fall	4	Yes	Raised	2,018	38.4	36	37.4	1.79	1.78
1/31/2004	Fall	4	Yes	Raised	2,024	37.7	33	37.6	1.61	1.63
2/6/2004	Fall	4	Yes	Raised	1,999	37.9	31	38.0	1.61	1.55
2/9/2004	Fall	4	Yes	Raised	2,017	37.8	27	37.0	1.69	1.34
2/13/2004	Fall	4	Yes	Raised	2,009	37.2	31	38.3	1.87	1.54
3/14/2004	Fall	3	No	Raised	1,401	38.3	18	39.6	1.98	1.28
3/23/2004	Fall	3	No	Raised	815	38.8	15	39.1	2.50	1.84
4/28/2004	Falí¹	4	Yes	Raised	1,304	72.9	33	71.7	1.94	2.53
5/4/2004	Fall ¹	4	No	Raised	814	75.5	18	75.1	3.35	2.21
5/18/2004	Falĺ	4	No	Lowered	867	80.2	10	75.1	3.20	1.15
5/26/2004	Falí ^l	4	No	Lowered	1,096	81.2	27	80.2	2.83	2.46
6/2/2004	Fall ¹	4	No	Lowered	888	76.2	28	77.2	2.77	3.15
6/15/2004	Falí¹	4	No	Lowered	691	76.4	12	79.1	2.17	1.74
8/31/2004	Winter	4	No	Lowered	1,096	36.5	41	36.0	3.00	3.74
9/3/2004	Winter	4	No	Lowered	1,153	36.6	50	35.6	3.23	4.34
9/17/2004	Winter	4	No	Raised	1,023	36.0	14	35.4	2.52	1.37

9/20/2004	Winter	4	No	Raised	1,017	35.8	21	35.4	2.48	2.06
9/23/2004	Winter	4	No	Raised	2,006	36.0	31	35.1	2.62	1.55
9/27/2004	Winter	4	No	Raised	1,918	36.1	36	36.1	2.77	1.88
10/1/2004	Winter	4	No	Raised	1,682	36.4	24	36.0	3.11	1.43
Model	Employed	#Trials	Slope	Intercept	Р	R²		•		
7/1/2004 -	6/30/2006	99	0.007464	0.00087452	<0.0001	0.385				

		# Traps	Traps		Releas	se Group	Recap	ture Group		
Date	Run	Sampling	Modified	RBDD Gates	N	FL (mm)	N	FL (mm)	%Q	%TE
1/23/2005	Fall	4	No	Raised	1,283	36.6	41	37.2	4.21	3.20
2/1/2005	Fall	3	Yes	Raised	1,971	36.6	31	36.0	1.35	1.57
2/10/2005	Fall	4	No	Raised	1,763	36.6	46	36.7	4.06	2.61
3/10/2005	Fall	4	No	Raised	1,216	36.6	27	36.5	3.93	2.22
3/13/2005	Fall	4	No	Raised	1,328	36.3	43	35.6	4.06	3.24
4/1/2005	Fall	4	No	Raised	1,949	57.1	50	62.3	3.49	2.57
9/11/2005	Winter	4	No	Lowered	1,437	35.6	14	38.9	2.22	0.97
10/4/2005	Winter	4	No	Raised	1,587	35.9	14	36.1	1.83	0.88
10/13/2005	Winter	4	No	Raised	1,577	35.7	21	36.6	2.33	1.33
2/15/2006	Fall	4	No	Raised	1,610	37.4	33	36.6	3.19	2.05
2/23/2006	Fall	4	No	Raised	1,503	37.2	38	36.6	2.68	2.53
1/21/2007	Fall	4	No	Raised	1,520	0.0	33	37.8	4.02	2.17
1/28/2007	Fall	4	Yes	Raised	1,987	37.6	18	37.8	3.65	0.91
2/5/2007	Fall	3	Yes	Raised	2,909	37.5	29	37.3	1.62	1.00
2/16/2007	Fall	4	No	Raised	1,782	37.9	34	38.5	3.51	1.91
3/2/2007	Fall	4	No	Raised	1,591	38.5	54	38.6	3.68	3.39
3/15/2007	Fall	4	No	Raised	953	37.6	26	37.6	4.29	2.73
3/20/2007	Fall	4	No	Raised	835	37.6	23	38.8	4.18	2.75

3/24/2007	Fall	4	No	Raised	944	37.7	23	38.0	4.24	2.44
Model	Employed	#Trials	Slope	Intercept	Р	₽²				
7/1/2006 -	6/30/2007	118	0.006653	0.00240145	<0.0001	0.420				
		# Traps	Traps		Releas	e Group	Recap	ture Group	_	
Date	Run	Sampling	Modified	RBDD Gates	N	FL (mm)	N	FL (mm)	%Q	%TE
1/23/2008	Fall	4	No	Raised	2,234	38.4	50	38.2	3.99	2.24
2/7/2008	Fall	4	Yes	Raised	2,324	38.1	60	37.9	2.19	2.58
2/14/2008	Fall	4	Mixed	Raised	1,993	38.4	83	38.8	3.40	4.16
2/20/2008	Fall	4	No	Raised	1,703	37.2	48	36.8	5.29	2.82
2/28/2008	Fall	3	No	Raised	2,080	37.6	63	38.3	3.45	3.03
Model	Employed	#Trials	Slope	Intercept	Р	R²				
7/1/2007 -	6/30/2008	123	0.00645	0.00303101	<0.0001	0.414				
		# Traps	Traps	<u></u>	Release	e Group	Recap	ture Group	_	
Date	Run	Sampling	Modified	RBDD Gates	N	FL (mm)	N	FL (mm)	%Q	%TE
1/23/2009	Fall	4	No	Raised	1,923	36.1	54	37.1	4.53	2.81
2/5/2009	Fall	4	No	Raised	1,868	36.8	58	37.4	4.65	3.10
Model	Employed	#Trials	Slope	Intercept	Р	R²				_
7/1/2008 -	6/30/2010	125	0.006332	0.00328530	<0.0001	0.425				
				-						
		# Traps	Traps	<u>. </u>	Release	e Group	Recap	ture Group		
Date	Run	# Traps Sampling	Traps Modified	RBDD Gates	Release N	e Group FL (mm)	Recap ^o N	ture Group FL (mm)	%Q	%TE
	Run Fall		-	RBDD Gates Raised			•		%Q 3.92	%TE 4.31
		Sampling	Modified		N	FL (mm)	N	FL (mm)		

2/11/2011	Fall	4	No	Raised	1,582	35.7	81	37.4	5.34	5.12
Model	Employed	#Trials	Slope	Intercept	Р	₽²				
7/1/2010 -	6/30/2012	129	0.007297	0.00123101	<0.0001	0.493				
		# Traps	Traps		Releas	e Group	Recap	ture Group		
Date	Run	Sampling	Modified	RBDD Gates	N	FL (mm)	N	FL (mm)	%Q	%TE
1/30/2012	Fall	4	No	Raised	1,319	36.3	46	36.1	4.08	3.49
2/4/2012	Fall	4	No	Raised	1,146	35.8	51	35.4	5.52	4.45
2/16/2012	Fall	4	No	Raised	1,465	35.7	73	35.0	5.36	4.98
2/28/2012	Fall	4	No	Raised	1,228	35.5	57	34.6	5.40	4.64
Model	Employed	#Trials	Slope	Intercept	Р	R²				
7/1/2012 -	6/30/2012	133	0.007676	0.00037735	< 0.0001	0.561				
		# Traps	Traps		Releas	e Group	Recap	ture Group		
Date	Run	# Traps Sampling	Traps Modified	RBDD Gates	Releas N	e Group FL (mm)	Recap	ture Group FL (mm)	%Q	%TE
Date 1/16/2013	Run Fall		-	RBDD Gates Raised					%Q 2.56	%TE 3.62
		Sampling	Modified		N	FL (mm)	N	FL (mm)		
1/16/2013	Fall	Sampling 4	Modified Yes	Raised	N 1,991	FL (mm) 35.6	N 72	FL (mm) 35.8	2.56	3.62
1/16/2013 1/23/2013	Fall Fall	Sampling 4 4	Modified Yes Yes	Raised Raised	N 1,991 1,965	FL (mm) 35.6 35.9	N 72 39	FL (mm) 35.8 35.3	2.56 2.61	3.62 1.98
1/16/2013 1/23/2013 1/30/2013	Fall Fall Fall	Sampling 4 4 4	Modified Yes Yes Yes	Raised Raised Raised	N 1,991 1,965 1,981	FL (mm) 35.6 35.9 36.3	N 72 39 44	FL (mm) 35.8 35.3 35.6	2.56 2.61 2.57	3.62 1.98 2.22
1/16/2013 1/23/2013 1/30/2013 2/3/2013	Fall Fall Fall Fall	Sampling 4 4 4 4	Modified Yes Yes Yes Yes Yes	Raised Raised Raised Raised	N 1,991 1,965 1,981 1,998	FL (mm) 35.6 35.9 36.3 36.5	N 72 39 44 42	FL (mm) 35.8 35.3 35.6 36.1	2.56 2.61 2.57 2.69	3.62 1.98 2.22 2.10
1/16/2013 1/23/2013 1/30/2013 2/3/2013 2/13/2013	Fall Fall Fall Fall Fall	Sampling 4 4 4 4 4	Modified Yes Yes Yes Yes Yes Yes Yes	Raised Raised Raised Raised Raised	N 1,991 1,965 1,981 1,998 2,079	FL (mm) 35.6 35.9 36.3 36.5 36.3	N 72 39 44 42 48	FL (mm) 35.8 35.3 35.6 36.1 36.2	2.56 2.61 2.57 2.69 2.62	3.62 1.98 2.22 2.10 2.31
1/16/2013 1/23/2013 1/30/2013 2/3/2013 2/13/2013 2/18/2013	Fall Fall Fall Fall Fall	\$\frac{4}{4}\$ 4 4 4 4 4 4 4	Modified Yes Yes Yes Yes Yes Yes Yes Yes	Raised Raised Raised Raised Raised Raised	N 1,991 1,965 1,981 1,998 2,079 2,156	FL (mm) 35.6 35.9 36.3 36.5 36.3 36.1	N 72 39 44 42 48 35	FL (mm) 35.8 35.3 35.6 36.1 36.2 36.8	2.56 2.61 2.57 2.69 2.62 2.89	3.62 1.98 2.22 2.10 2.31 1.62
1/16/2013 1/23/2013 1/30/2013 2/3/2013 2/13/2013 2/18/2013 2/22/2013	Fall Fall Fall Fall Fall Fall	\$\frac{4}{4} & 4 & 4 & 4 & 4 & 4 & 4 & 4 & 4 & 4 &	Modified Yes Yes Yes Yes Yes Yes Yes No	Raised Raised Raised Raised Raised Raised	N 1,991 1,965 1,981 1,998 2,079 2,156 2,439	FL (mm) 35.6 35.9 36.3 36.5 36.3 36.1 36.7 36.1 36.5	N 72 39 44 42 48 35 119	FL (mm) 35.8 35.3 35.6 36.1 36.2 36.8 36.6	2.56 2.61 2.57 2.69 2.62 2.89 6.52	3.62 1.98 2.22 2.10 2.31 1.62 4.88
1/16/2013 1/23/2013 1/30/2013 2/3/2013 2/13/2013 2/18/2013 2/22/2013 2/26/2013	Fall Fall Fall Fall Fall Fall Fall	\$\frac{4}{4} \\ 4\\ 4\\ 4\\ 4\\ 4\\ 4\\ 4\\ 4\\ 4\	Modified Yes Yes Yes Yes Yes Yes No No	Raised Raised Raised Raised Raised Raised Raised Raised Raised	N 1,991 1,965 1,981 1,998 2,079 2,156 2,439 1,400	FL (mm) 35.6 35.9 36.3 36.5 36.3 36.1 36.7 36.1	N 72 39 44 42 48 35 119 65	FL (mm) 35.8 35.3 35.6 36.1 36.2 36.8 36.6 37.3	2.56 2.61 2.57 2.69 2.62 2.89 6.52 6.87	3.62 1.98 2.22 2.10 2.31 1.62 4.88 4.64

^{7/1/2013 - 9/30/2013 142 0.007255 0.00150868 &}lt;0.0001 0.587

1 Denotes Coleman National Fish Hatchery Fall Chiprocoduction fish used during trial.

Table 3. Annual capture fork length summar@.ofiykiss by age and life-stage classification from the RBDD trap project between April 2002 through December 2012 by calenyelar (CY).

		Age Classifica			Life Stage Classification (%)						
	Fry	Sub-Yearling	Yearling	2+		Yolk- Silvery			Silvery-		
CY	<41 mm	41-138 mm	139-280 mm	>280 mm	CY	sac Fry	Fry	Parr	parr	Smolt	
2002	11.2	86.7	1.6	0.5	2002	0.0	6.3	54.4	37.2	2.1	
2003	8.1	89.5	2.3	0.0	2003	0.0	5.6	57.7	34.9	1.8	
2004	9.8	89.7	0.5	0.0	2004	0.0	4.6	60.2	34.7	0.5	
2005	3.5	93.2	3.1	0.2	2005	0.0	2.8	48.7	45.6	2.9	
2006	17.5	75.3	5.6	1.5	2006	0.2	9.2	78.9	9.2	2.4	
2007	6.5	91.2	1.7	0.6	2007	0.1	8.7	85.3	5.3	0.6	
2008	6.3	92.3	0.9	0.5	2008	0.1	8.2	79.4	12.0	0.4	
2009	9.0	87.7	2.1	1.2	2009	0.0	10.7	82.8	5.1	1.4	
2010	7.7	89.8	1.7	0.8	2010	0.3	9.7	87.4	1.7	1.0	
2011	4.6	89.7	5.0	0.6	2011	0.1	3.5	90.9	2.8	2.7	
2012	6.6	90.0	2.3	1.1	2012	0.2	5.9	88.2	4.2	1.5	
Mean	8.3	88.7	2.4	0.6	Mean	0.1	6.8	74.0	17.5	1.6	
SD	3.8	4.8	1.6	0.5	SD	0.1	2.6	15.5	16.8	0.9	

Table 4. Annual linear regression equations **%**5% confidence intervals (CI) for Log transformed juvenile (80-200 mr0) mykiss weight-length data sampled at the RBDD rotary traps from April 2002 through December 2001/2calendar year (CY).

			Slope	
CY	Weight-Length Equation	\mathbb{R}^2	Lower 95% CI Upper	r 95% CI
2002	Log ₀ (weight)=2.843(LogFL)-4.616	0.903	2.648	3.039
2003	Log ₀ (weight)=2.968(LogFL)-4.886	0.968	2.885	3.052
2004	Log ₀ (weight)=3.005(LogFL)-4.941	0.952	2.879	3.132
2005	Log ₀ (weight)=3.03(LogFL)-5.009	0.952	2.929	3.132
2006	$Log_0(weight)=3.052(LogFL)-5.085$	0.917	2.811	3.293
2007	Log ₀ (weight)=2.961(LogFL)-4.864	0.947	2.853	3.069
2008	Log ₀ (weight)=2.939(LogFL)-4.819	0.942	2.833	3.044
2009	Log ₀ (weight)=3.017(LogFL)-4.981	0.974	2.922	3.112
2010	Log ₀ (weight)=2.977(LogFL)-4.911	0.934	2.836	3.118
2011	Log ₀ (weight)=2.911(LogFL)-4.778	0.939	2.743	3.078
2012	Log ₀ (weight)=2.858(LogFL)-4.662	0.903	2.746	2.970
Mean	Log ₀ (weight)=2.946(LogFL)-4.840	0.942	2.913	2.979

Table 5a. RBDD rotary trap fall Chinook totaluah effort and passage estimates (sum of weekly values), lower and upper 90% confidenterivals (CI), ratio of fry to presmolt/smolt passage and ratio of estimated pass(Figst) and interpolated passage (Interp) for brood year (BY) 2002-2012.

BY	Effort	Total	Low 90%C	Up 90%	CI I	Fry S	molt	Ensterp I
2002	0.76	17,038,417	857,106	47,315,257	0.86	0.14	0.54	0.46
2003	0.81	27,736,868	8,839,840	50,653,446	0.85	0.15	0.74	0.26
2004	0.85	14,108,238	5,079,300	24,967,671	0.56	0.44	0.70	0.30
2005	0.56	18,210,294	3,500,275	39,096,017	0.64	0.36	0.40	0.60
2006	0.90	16,107,651	6,522,666	26,414,402	0.63	0.37	0.85	0.15
2007	0.88	12,131,603	6,130,892	18,170,520	0.79	0.21	0.84	0.16
2008	0.79	9,115,547	4,381,560	13,849,709	0.73	0.27	0.81	0.19
2009	0.84	8,532,377	3,064,273	14,052,588	0.81	0.19	0.56	0.44
2010	0.75	8,842,481	4,727,816	13,252,907	0.71	0.29	0.79	0.21
2011	0.87	6,271,261	3,431,940	9,125,109	0.71	0.29	0.82	0.18
2012	0.85	24,429,420	16,028,521	33,112,943	0.87	0.13	0.91	0.09
Mean	0.81	14,774,923			0.74	0.26	0.72	0.28
SD	0.09	6,825,382			0.10	0.10	0.16	0.16
CV	11.7%	46.2%			13.9%	40.3%	22.0%	57.4%

Table 5b. RBDD rotary trap late-fall Chinookltathnual effort and passage estimates (sum of weekly values), lower and upper 90% confidentervals (CI), ratio of fry to pre-smolt/smolt passage and ratio of estimated pages (Est) and interpolated passage (Interp) for brood year (BY) 2002-2012.

BY	Effort	Total	Low 90%CI	Up 90%	CI	Fry S	Smolt	Ensterp I
2002	0.57	2,559,519	659,986	4,953,910	0.17	0.83	0.52	0.48
2003	0.76	346,058	78,407	911,270	0.57	0.43	0.56	0.44
2004	0.88	147,160	74,930	220,231	0.17	0.83	0.91	0.09
2005	0.73	143,362	41,800	333,415	0.35	0.65	0.71	0.29
2006	0.70	460,268	125,197	902,089	0.62	0.38	0.44	0.56
2007	0.90	535,619	271,079	800,447	0.27	0.73	0.86	0.14
2008	0.89	91,995	46,660	138,310	0.11	0.89	0.89	0.11
2009	0.72	219,824	97,294	342,652	0.13	0.87	0.73	0.27
2010	0.86	183,439	61,775	305,937	0.62	0.38	0.61	0.39
2011	0.77	97,040	28,738	165,997	0.72	0.28	0.53	0.47
2012	0.89	140,534	42,673	249,500	0.48	0.52	0.80	0.20
Mean	0.79	447,711			0.38	0.62	0.69	0.31
SD	0.10	715,999			0.23	0.23	0.16	0.16
CV	13.2%	159.9%			58.8%	36.5%	23.8%	52.5%

Table 5c. RBDD rotary trap winter Chinook **tartari**ual effort and passage estimates (sum of weekly values), lower and upper 90% co**nfide** intervals (CI), ratio of fry to pre-smolt/smolt passage and ratio of estimated **pages** (Est) and interpolated passage (Interp) for brood year (BY) 2002-2012.

BY	Effort	Total	Low 90%C	I Up 90%	CI	Fry S	molt	Ensterp I
2002	0.64	7,119,041	2,541,407	12,353,367	0.90	0.10	0.58	0.42
2003	0.81	5,221,016	3,202,609	7,260,798	0.85	0.15	0.86	0.14
2004	0.84	3,434,683	1,998,468	4,874,794	0.90	0.10	0.82	0.18
2005	0.64	8,363,106	4,558,069	12,277,233	0.90	0.10	0.89	0.11
2006	0.83	6,687,079	3,801,539	9,575,937	0.87	0.13	0.76	0.24
2007	0.89	1,440,563	931,113	1,953,688	0.80	0.20	0.92	0.08
2008	0.87	1,244,990	776,634	1,714,013	0.85	0.15	0.77	0.23
2009	0.75	4,402,322	2,495,734	6,311,739	0.81	0.19	0.74	0.26
2010	0.81	1,285,389	817,207	1,756,987	0.68	0.32	0.92	0.08
2011	0.82	848,976	576,177	1,122,022	0.75	0.25	0.88	0.12
2012	0.89	1,349,819	904,552	1,795,106	0.53	0.47	0.92	0.08
Mean	0.80	3,763,362			0.80	0.20	0.82	0.18
SD	0.09	2,753,256			0.11	0.11	0.11	0.11
CV_	10.9%	73.2%			13.9%	57.5%	12.8%	59.6%

Table 5d. RBDD rotary trap spring Chinook **tortal**ual effort and passage estimates (sum of weekly values), lower and upper 90% co**nfide**intervals (CI), ratio of fry to pre-smolt/smolt passage and ratio of estimated **pages** (Est) and interpolated passage (Interp) for brood year (BY) 2002-2012.

 BY	Effort	Total	Low 90%C	Up 90%	6 CAry	Smolt	Est	Interp
2002	0.75	277,477	110,951	494,590	0.57	0.43	0.59	0.41
2003	0.81	626,915	249,225	1,053,421	0.80	0.20	0.67	0.33
2004	0.85	430,951	174,174	710,419	0.36	0.64	0.78	0.22
2005	0.57	616,040	131,328	1,382,036	0.69	0.30	0.58	0.42
2006	0.89	421,436	239,470	603,952	0.41	0.59	0.80	0.20
2007	0.89	369,536	229,766	510,868	0.91	0.09	0.99	0.01
2008	0.85	164,673	66,515	262,959	0.24	0.76	0.62	0.38
2009	0.79	438,405	176,952	700,959	0.50	0.50	0.51	0.49
2010	0.77	158,966	62,563	261,105	0.56	0.44	0.67	0.33
2011	0.86	184,290	101,443	272,769	0.48	0.52	0.85	0.15
2012	0.86	320,897	173,312	469,137	0.42	0.58	0.74	0.26
Mean	0.81	364,508			0.54	0.46	0.71	0.29
SD	0.09	164,135			0.20	0.20	0.14	0.14
 CV	11.3%	45.0%			36.4%	43.0%	19.7%	47.6%

Table 5e. RBDD rotary tropmykiss total annual effort and passage estimates (sum of weekly values), lower and upper 90% confidencerinals (CI), and ratio of estimated passage (Est) and interpolated passage (Interp) afterndar year (CY) 2002-2012.

CY	Effort	Total	Low 90%CI	Up 90%	6 CEIst	Interp
2002 ¹	0.53	124,436	27,224	244,701	0.53	0.47
2003	0.76	139,008	54,885	243,927	0.78	0.22
2004	0.83	151,694	86,857	218,132	0.95	0.05
2005	0.83	85,614	32,251	152,568	0.76	0.24
2006	0.59	83,801	20,603	169,712	0.44	0.56
2007	0.91	139,424	73,827	205,647	0.89	0.11
2008	0.89	131,013	69,331	193,584	0.88	0.12
2009	0.76	129,581	62,350	197,795	0.83	0.17
2010	0.85	100,997	47,050	155,692	0.74	0.26
2011	0.76	56,798	23,494	89,369	0.76	0.24
2012	0.86	136,621	78,804	194,892	0.96	0.04
Mean	0.78	116,272			0.78	0.22
SD	0.12	29,912			0.16	0.16
CV	15.6%	25.7%			20.9%	72.2%

Incomplete year; sampling began in April 2002.

Table 6a. Fall Chinook fry-equivalent productistimates, lower and upper 90% confidence inter(als), estimates of adults upstream of RBDD (Adult Estimate), estimated fertraleale sex ratios, estimated females, estimatesentiale fecundity, calculated juveniles per estimated female (recripites female) and egg-to-fry survival estimates (EDT/Fbrood year (BY) for Chinook sampled at RBDD rotary traps between December 2002September 2013.

oampioa	dampied at 1833 fetally trape settled in Secondary Description 2016.											
	FRY EQ	Lower	Upper	Adult	Sex	Ratio	Estimated		Recruits per			
BY	Passage	90% CI	90% CI	Estimate	(F:	M)	Females	Fecundity	Female	ETF		
2002	18,683,720	1,216,244	51,024,926	458,772	0.46	0.54	211,035	5,407	89	1.6%		
2003	30,624,209	10,162,712	55,109,506	140,724	0.57	0.44	79,509	5,407	385	7.1%		
2004	18,421,457	6,224,790	33,728,746	64,276	0.48	0.52	31,045	5,407	593	11.0%		
2005	22,739,315	4,235,720	49,182,045	80,294	0.47	0.53	37,738	5,407	603	11.1%		
2006	20,276,322	8,670,090	32,604,760	78,692	0.54	0.46	42,730	5,407	475	8.8%		
2007	13,907,856	7,041,759	20,838,463	31,592	0.54	0.46	16,996	5,407	818	15.1%		
2008	10,817,397	5,117,059	16,517,847	36,104	0.46	0.54	16,644	5,407	650	12.0%		
2009	9,674,829	3,678,373	15,723,368	12,908	0.51	0.49	6,531	5,407	1,481	27.4%		
2010	10,620,144	5,637,617	15,895,197	29,321	0.24	0.76	7,008	5,407	1,515	28.0%		
2011	7,554,574	4,171,332	10,960,125	31,931	0.29	0.71	9,260	5,407	816	15.1%		
2012	26,567,379	17,219,525	36,197,837	65,664	0.50	0.50	32,635	5,407	814	15.1%		
Mean	17,262,473	6,670,475	30,707,529	93,662	0.46	0.54	44,648	•	749	13.9%		
CV	43.2%	64.0%	51.7%	134.7%			132.4%		57.2%	57.2%		

¹ Sex ratios based on RBDD fish ladder data bet**2/ee3** and 2007 and CNFH data between 2008 and 240M2 rage, in italics, input for 2002 due to lack of available data. ² Female fecundity estimates based on average váltoes CNFH fall Chinook spawning data collected/bet 2008 and 2012.

Table 6b. Late-fall Chinook fry-equivalent protition estimates, lower and upper 90% confidence rivrates (CI), estimates of adults upstream of RBDD (Adult Estimate), estimated fertoaleale sex ratios, estimated females, estimatesenfalle fecundity, calculated juveniles per estimated female, and engine survival estimates (ETF) by brood year (PAYC) Ininook sampled at RBDD rotary traps between April 2002 and March 2013.

	totally diagnostic print 2002 and material 2010.											
	FRY EQ	Lower	Upper	Adult	Sex F		Estimated		Recruits per			
BY	Passage	90% CI	90% CI	Estimate	(F: I	M)	Females	Fecundity	Female	ETF		
2002	4,041,505	1,063,720	7,808,619	36,220	0.46	0.54	16,661	4,662	243	5.2%		
2003	451,230	133,225	1,067,819	5,513	0.46	0.54	2,536	4,662	178	3.8%		
2004	233,106	124,245	342,837	8,924	0.46	0.54	4,105	4,662	57	1.2%		
2005	209,066	70,548	441,133	9,610	0.46	0.54	4,421	4,662	47	1.0%		
2006	582,956	186,984	1,086,699	7,770	0.46	0.54	3,574	4,662	163	3.5%		
2007	809,272	426,272	1,192,625	13,939	0.46	0.54	6,412	4,662	126	2.7%		
2008	149,049	80,500	218,597	3,747	0.46	0.54	1,724	4,662	86	1.9%		
2009	353,003	159,726	546,546	3,792	0.46	0.54	1,744	4,662	202	4.3%		
2010	232,279	89,343	376,286	3,961	0.46	0.54	1,822	4,662	127	2.7%		
2011	116,188	38,688	194,400	3,777	0.46	0.54	1,737	4,662	67	1.4%		
2012	191,672	69,229	325,189	2,931	0.46	0.54	1,348	4,662	142	3.0%		
Mean	669,939	222,044	1,236,432	9,108			4,190	-	131	2.8%		
CV	169.8%	134.4%	178.7%	105.5%			105.5%		48.1%	48.1%		

¹ Sex ratio value of (0.46:0.54) is equivalent to alwerage ratio for fall Chinook between 2003 a0d2used in Table 6a.
² Female fecundity estimates based on average values CNFH late-fall Chinook spawning data collebted ween 2008 and 2012.

Table 6c. Winter Chinook fry-equivalent productestimates, lower and upper 90% confidence in the confid upstream of RBDD (Adult Estimate), estimated fertoaleale sex ratios, estimated females, estimatesenfalle fecundity, calculated juveniles per estimated female (recripites female) and egg-to-fry survival estimates (EDT/Fbrood year (BY) for Chinook sampled at RBDD rotary traps between July 2002Jane 2013.

	FRY EQ	Lower	Upper	Adult	Sex I	Ratio	Estimated	-	Recruits per	
BY	Passage	90% CI	90% CI	Estimate	(F:	M)	Females	Fecundity	Female	ETF
2002	7,635,469	2,811,132	13,144,325	7337	0.77	0.23	5,670	4,923	1,347	27.4%
2003	5,781,519	3,525,098	8,073,129	8133	0.64	0.36	5,179	4,854	1,116	23.0%
2004	3,677,989	2,129,297	5,232,037	8635	0.37	0.63	3,185	5,515	1,155	20.9%
2005	8,943,194	4,791,726	13,277,637	15730	0.56	0.44	8,807	5,500	1,015	18.5%
2006	7,298,838	4,150,323	10,453,765	17205	0.50	0.50	8,626	5,484	846	15.4%
2007	1,637,804	1,062,780	2,218,745	2488	0.61	0.39	1,517	5,112	1,080	21.1%
2008	1,371,739	858,933	1,885,141	2850	0.51	0.49	1,443	5,424	951	17.5%
2009	4,972,954	2,790,092	7,160,098	4537	0.60	0.40	2,702	5,519	1,840	33.3%
2010	1,572,628	969,016	2,181,572	1533	0.53	0.47	813	5,161	1,934	37.5%
2011	996,621	671,779	1,321,708	824	0.51	0.49	424	4,832	2,351	48.6%
2012	1,789,259	1,157,240	2,421,277	2581	0.58	0.42	1,491	4,518	1,200	26.6%
Mean	4,152,547	2,265,220	6,124,494	6,532	0.56	0.44	3,623	5,167	1,349	26.4%
CV	70.1%	64.0%	74.9%	85.7%	17.9%	22.9%	83.4%	6.7%	35.5%	37.9%

Annual sex ratio values based on annual carcarseys estimates of female recoveries.

Female fecundity estimates based on annual values LSNFH winter Chinook spawning data collected then 2002 and 2012.

Table 6d. Spring Chinook fry-equivalent produncestimates, lower and upper 90% confidence inates (CI), estimates of adults upstream of RBDD (Adult Estimate), estimated fertcaleale sex ratios, estimated females, estimates of adults calculated juveniles per estimated female (recruites female) and egg-to-fry survival estimates (EDI) for Chinook sampled at RBDD rotary traps between October 162200d September 30, 2013.

campica	FRY EQ	Lower	Upper	Adult	•	Ratio	Estimated	Recruits per			
BY	Passage	90% CI	90% CI	Estimate	1		Females	Fecundity	Female	ETF	
2002	360,352	142,134	657,043	608	0.46	0.54	280	5,078	1,288	25.4%	
2003	714,086	293,095	1,187,827	319	0.46	0.54	147	5,078	4,866	95.8%	
2004	624,079	255,886	1,029,162	575	0.46	0.54	265	5,078	2,359	46.5%	
2005	747,026	146,488	1,695,236	189	0.46	0.54	87	5,078	8,592	169.2%	
2006	594,511	328,845	860,757	353	0.46	0.54	162	5,078	3,661	72.1%	
2007	392,451	242,563	544,184	767	0.46	0.54	353	5,078	1,112	21.9%	
2008	251,795	96,737	406,863	305	0.46	0.54	140	5,078	1,795	35.3%	
2009	591,549	238,710	945,904	314	0.46	0.54	144	5,078	4,095	80.7%	
2010	207,793	80,320	344,475	208	0.46	0.54	96	5,078	2,172	42.8%	
2011	251,444	130,051	382,077	167	0.46	0.54	77	5,078	3,273	64.5%	
2012	451,705	238,187	665,825	868	0.46	0.54	399	5,078	1,131	22.3%	
Mean	471,527	199,365	792,668	425	-	•	195		3,122	61.5%	
CV	40.9%	41.7%	51.5%	56.8%			56.8%		70.8%	70.8%	

Sex ratio value of (0.46:0.54) is equivalent to always a ratio for fall Chinook between 2003 a0d(22) used in Table 6a.

² Female fecundity estimates based on average of the provided of the provide

Table 7. Green Sturgeon annual capture, catchupit volume (CPUV) and total length summaries for sturgeon captured by RBDDryothaps between calendar year (CY) 2002 and 2012.

		CPUV	Min TL	Max TL	Mean	Median
CY	Captures	fish/ac-ft	(mm)	(mm)	(mm)	(mm)
2002	35	0.3	23	52	28.8	27.5
2003	360	1.9	22	188	27.8	27
2004	266	1.0	21	58	30.5	29
2005	271	1.1	24	65	28.9	27
2006	193	0.8	21	79	30.5	28
2007	19	0.1	25	49	29.6	27
2008	0	0.0	-	-	-	-
2009	32	0.2	24	47	28.0	26
2010	70	0.5	20	36	27.1	27
2011	3701	20.1	18	86	27.4	27
2012	288	1.4	21	41	27.2	27
Ave	475.9	2.5	21.9	70.1	28.6	27.3
SD	1077.4	5.9	2.1	44.4	1.3	8.0
CV	226.4%	236.3%	9.7%	63.3%	4.5%	2.9%

Table 8a. Unidentified Lamprey ammocoetes an **napt**ure, catch per unit volume (CPUV) and total length summaries for ammocoet**esure**d by RBDD rotary traps between water year (WY) 2003 and 2013.

	,	CPUV	Min TL	Max TL	Mean	Median
WY	Captures	Fish/ac-ft	(mm)	(mm)	(mm)	(mm)
2003	908	7.30	14	144	98	100
2004	925	6.80	27	191	105	108
2005	1415	11.65	22	159	104	108
2006	657	4.45	52	186	112	115
2007	556	5.16	29	155	105	111
2008	385	3.64	41	146	101	108
2009	593	5.53	41	150	106	112
2010	935	11.45	45	166	111	114
2011	859	7.07	30	186	111	117
2012	455	5.11	27	155	100	104
2013	632	6.45	25	160	103	107
Mean	756.4	6.8	32.1	163.5	105.1	109.5
SD	291.3	2.6	11.3	16.8	4.7	5.0
CV	38.5%	38.5%	35.1%	10.3%	4.5%	4.6%

Table 8b. Pacific Lamprey macrothalmia and authitual capture, catch per unit volume (CPUV) and total length summaries for matratoria captured by RBDD rotary traps between water year (WY) 2003 and 2013.

		CPUV	Min TL	Max TL	Mean	Median
WY	Captures	Fish/ac-ft	(mm)	(mm)	(mm)	(mm)
2003	204	2.16	100	693	261	131
2004	478	3.91	96	630	149	125
2005	4645	45.00	72	665	137	126
2006	417	5.62	98	700	136	125
2007	3107	34.08	96	660	150	128
2008	5252	40.29	78	580	139	128
2009	2938	81.24	91	834	132	124
2010	699	32.30	80	819	136	125
2011	2747	68.18	92	620	140	129
2012	3464	112.76	86	500	136	127
2013	1734	25.63	88	617	131	127
Mean	2335.0	41.0	88.8	665.3	149.7	126.8
SD	1759.4	34.7	9.0	97.1	37.3	2.1
CV	75.3%	84.5%	10.2%	14.6%	24.9%	1.6%

Table 9a. Summary of fall Chinook abiotic samphelicions at RBDD rotary traps during dates of captby brood year (BY).

-	Dat	tes of Captu	re	H ₂ 0 Te	emperati	ure°(F)		Discharge (CF		Turbidity (NTU)		
BY	Initial	Final	Days	Min	Max	Ave	Min	Max	Ave	Min	Max	Ave
2002	4-Dec	30-Aug	269	47	61	55	6,390	86,500	17,471	0.5	240.2	19.6
2003	9-Dec	15-Aug	250	46	62	55	7,380	92,800	18,707	2.0	413.5	21.8
2004	8-Dec	29-Aug	264	46	63	56	5,390	76,200	13,315	1.9	626.5	24.6
2005	3-Dec	29-Aug	269	47	61	53	6,450	118,000	27,279	1.6	731.7	22.5
2006	10-Dec	26-Aug	259	46	62	55	6,030	45,400	10,628	1.6	90.0	8.0
2007	7-Dec	2-Sep	270	44	62	55	5,210	44,600	10,127	1.5	233.3	11.1
2008	5-Dec	4-Sep	273	45	64	56	4,160	33,000	9,297	2.1	129.8	12.0
2009	10-Dec	21-Aug	254	45	61	54	5,260	95,100	17,531	1.3	162.6	10.3
2010	7-Dec	29-Aug	265	45	61	54	5,260	95,100	17,331	1.3	162.6	10.2
2011	10-Dec	2-Sep	267	45	65	55	4,800	35,200	10,281	1.4	180.6	8.8
2012	2-Dec	23-Aug	264	44	64	56	5,330	70,400	11,323	1.5	315.5	9.9
Mean	7-Dec	27-Aug	264	45	62	55	5,605	72,027	14,844	1.5	298.7	14.4
SD			7	1.1	1.4	8.0	890	28,600	5,442	0.4	209.6	6.3
CV			3%	2%	2%	1%	16%	40%	37%	28%	70%	44%

Table 9b. Summary of late-fall Chinook abioticpslemonditions at RBDD rotary traps during datesapfture by brood year (BY).

	Dat	es of Captu	re	H ₂ 0 Te	emperati	ure⁰(F)		Discharge (CF	Tı	Turbidity (NTU)		
BY	Initial	Final	Days	Min	Max	Ave	Min	Max	Ave	Min	Max	Ave
2002	19-Apr	14-Jan	270	47	62	57	6,176	86,500	12,981	0.4	59.7	11.3
2003	3-Apr	6-Mar	338	46	61	55	6,310	92,800	16,650	0.9	413.5	20.9
2004	2-Apr	21-Jan	294	46	62	57	5,170	57,000	10,983	1.4	470.0	8.0
2005	2-Apr	22-Jan	295	48	63	57	6,050	118,000	17,431	1.6	731.7	24.4
2006	1-Apr	13-Jan	287	46	61	55	6,610	80,900	15,374	2.0	178.0	8.8
2007	4-Apr	9-Jan	280	46	62	57	5,490	38,600	10,035	1.3	198.0	5.7
2008	2-Apr	2-Mar	334	45	64	56	4,160	33,000	8,775	1.5	129.8	6.9
2009	3-Apr	1-Mar	332	46	64	57	3,920	60,400	9,855	1.9	250.6	14.2
2010	1-Apr	12-Jan	286	47	62	56	5,900	50,600	11,831	1.1	220.3	7.3
2011	1-Apr	27-Jan	301	45	61	55	5,570	57,400	11,888	2.0	68.5	5.5
2012	2-Apr	11-Jan	284	46	62	56	5,536	67,520	12,580	1.4	272.0	11.3
Mean	4-Apr	29-Jan	300	46	62	56	5,536	67,520	12,580	1.4	272.0	11.3
SD			24	0.9	1.0	0.7	849	25,109	2,829	0.5	198.7	6.2
CV			8%	2%	2%	1%	15%	37%	22%	34%	73%	55%

Table 9c. Summary of winter Chinook abiotic sampheditions at RBDD rotary traps during dates of twae by brood year (BY).

	Dat	es of Captu	re	H ₂ 0 Te	emperati	ure⁰(F)		Discharge (CF	Tı	Turbidity (NTU)		
BY	Initial	Final	Days	Min	Max	Ave	Min	Max	Ave	Min	Max	Ave
2002	4-Jul	8-Apr	278	47	61	55	6,176	86,500	14,081	0.4	240.2	13.5
2003	16-Jul	17-Mar	245	46	61	54	6,310	92,800	16,809	0.9	413.5	22.8
2004	22-Jul	25-Mar	246	46	62	55	5,170	57,000	9,817	1.4	470.0	12.1
2005	25-Jul	17-Feb	207	48	61	55	6,450	118,000	19,174	1.6	731.7	19.7
2006	16-Jul	10-Mar	237	46	59	54	6,030	45,400	9,788	1.6	90.0	7.2
2007	18-Jul	4-Apr	261	44	62	54	5,210	44,600	9,318	1.3	233.3	11.3
2008	30-Jul	24-Apr	268	45	64	55	4,160	33,000	7,647	1.5	129.8	8.2
2009	26-Jul	30-Mar	247	46	64	55	3,920	60,400	9,303	1.9	250.6	15.0
2010	18-Jul	7-Apr	263	45	61	54	5,260	95,100	14,941	1.1	162.6	8.6
2011	12-Aug	31-Mar	232	45	60	53	4,800	35,200	8,646	1.7	180.6	7.0
2012	23-Jul	19-Apr	270	46	61	55	5,349	66,800	11,952	1.3	290.2	12.5
Mean	22-Jul	28-Mar	250	46	61	55	5,349	66,800	11,952	1.3	290.2	12.5
SD			20	1.1	1.5	8.0	843	27,776	3,767	0.4	185.4	5.1
CV			8%	2%	2%	1%	16%	42%	32%	31%	64%	41%

Table 9d. Summary of spring Chinook abiotic samphelitions at RBDD rotary traps during dates options by brood year (BY).

	Da	tes of Captu	re	H ₂ 0 Te	emperati	ure⁰(F)		Discharge (CF	S)	Turbidity (NTU)		
BY	Initial	Final	Days	Min	Max	Ave	Min	Max	Ave	Min	Max	Ave
2002	16-Oct	29-May	225	47	61	54	6,176	86,500	16,877	0.4	240.2	19.1
2003	16-Oct	11-Jun	239	46	62	54	6,310	92,800	17,267	0.9	413.5	23.0
2004	16-Oct	3-Jun	230	46	63	54	5,170	76,200	11,612	1.4	626.5	27.6
2005	16-Oct	3-Jun	230	47	61	52	6,450	118,000	28,158	1.6	731.7	25.3
2006	16-Oct	26-May	222	46	62	53	6,030	45,400	8,630	1.6	90.0	8.3
2007	16-Oct	12-Jun	240	44	61	53	5,210	44,600	8,823	1.3	233.3	11.4
2008	16-Oct	7-Jun	234	45	64	54	4,160	33,000	7,841	1.7	129.8	10.1
2009	16-Oct	25-May	221	46	62	54	3,920	60,400	9,495	1.9	250.6	17.1
2010	16-Oct	12-Jun	239	45	61	53	5,260	95,100	16,656	1.3	162.6	9.9
2011	16-Oct	27-May	224	45	65	53	4,800	35,200	8,344	1.7	180.6	8.8
2012	16-Oct	23-Jun	250	46	62	53	5,349	68,720	13,370	1.4	305.9	16.0
Mean	16-Oct	4-Jun	232	46	62	53	5,349	68,720	13,370	1.4	305.9	16.0
SD			9	1.0	1.4	0.6	843	27,696	6,116	0.4	205.5	7.0
CV			4%	2%	2%	1%	16%	40%	46%	30%	67%	43%

Table 9e. Summary of mykiss abiotic sample conditions at RBDD rotary trapsingudates of capture by calendar year (CY).

	Dat	es of Captu	re	H ₂ 0 Te	emperati	ure⁰(F)	D	ischarge (CF	S)	Turbidity (NTU)		
CY	Initial	Final	Days	Min	Max	Ave	Min	Max	Ave	Min	Max	Ave
2002 ¹	-	-	-	-	-	-	-	_	-	_	-	_
2003	19-Jan	30-Dec	345	46	61	56	6,310	56,800	13,677	0.9	240.2	16.4
2004	6-Jan	17-Dec	346	46	62	56	5,170	92,800	14,613	1.4	413.5	9.3
2005	1-Jan	29-Dec	362	46	63	56	5,890	94,700	12,661	1.6	626.5	20.1
2006	3-Jan	30-Dec	361	47	61	54	6,610	82,900	20,803	2.0	190.5	11.4
2007	16-Jan	27-Dec	345	46	62	56	5,510	45,400	9,596	1.3	74.5	6.4
2008	6-Jan	28-Dec	357	44	64	56	4,610	44,600	9,478	1.5	233.3	9.0
2009	12-Jan	25-Dec	347	45	64	57	4,020	33,000	8,775	1.9	129.8	10.3
2010	15-Jan	12-Dec	331	47	62	56	5,150	60,400	11,194	1.1	250.6	12.4
2011	1-Jan	30-Dec	363	45	61	55	5,260	95,100	13,833	1.3	162.6	7.2
2012	17-Jan	14-Dec	332	45	65	56	4,800	70,400	10,557	1.2	315.5	11.0
Mean	10-Jan	23-Dec	349	46	63	56	5,333	67,610	12,519	1.4	263.7	11.4
SD			12	0.9	1.3	8.0	783	22,986	3,551	0.3	159.1	4.1
CV			3%	2%	2%	1%	15%	34%	28%	24%	60%	37%

Sampling did not begin until mid-April of 2002 athits year not included in analyses.

Table 9f. Summary of Green Sturgeon abiotic sammelitions at RBDD rotary traps during dates of twee by calendar year (CY).

	Date	es of Captur	e	-	emperati	ure⁰(F)_		ischarge (CF	(S)	Turbidity (NTU)		
CY	Initial	Final	Days	Min	Max	Ave	Min	Max	Ave	Min	Max	Ave
2002	7-May	16-Jul	70	55	60	58	9,317	15,680	13,038	0.9	16.3	3.5
2003	13-Jun	11-Nov	151	52	61	58	6,950	16,000	10,802	0.9	48.6	6.5
2004	4-May	29-Jul	86	55	60	58	9,560	16,700	14,210	3.0	18.3	4.9
2005	7-May	13-Aug	98	54	61	58	10,200	76,200	18,614	2.3	626.5	26.4
2006	10-Jun	25-Aug	76	56	59	57	12,800	15,600	14,579	3.4	13.9	5.7
2007	11-May	24-Jul	74	55	61	58	9,790	17,000	12,905	1.7	50.4	4.5
2008	-	-	0	-	-	-	-	-	-	-	-	-
2009	11-May	16-Jul	66	58	64	61	9,460	13,700	11,226	4.1	34.4	13.5
2010	26-May	29-Aug	95	55	61	58	9,150	18,300	13,143	1.6	22.0	5.4
2011	16-May	27-Aug	103	52	61	58	10,400	24,800	14,059	3.6	23.5	6.8
2012	1-May	26-Jun	56	55	61	58	8,763	21,398	12,258	2.2	85.4	7.7
Mean	17-May	12-Aug	88	55	61	58	9,639	23,538	13,483	2.4	93.9	8.5
SD			27	1.7	1.2	0.9	1,464	18,782	2,181	1.1	188.4	6.9
CV			31%	3%	2%	2%	15%	80%	16%	47%	201%	81%

Table 9g. Summary of Lampsepp. abiotic sample conditions at RBDD rotary trapsingudates of capture by water year (WY).

	Date	es of Capture		H₂0 Te	emperati	ure⁰(F)		ischarge (CF		Turbidity (NTU)		
WY	Initial	Final	Days	Min	Max	Ave	Min	Max	Ave	Min	Max	Ave
2003	1-Oct	27-Sep	361	47	61	56	6,176	86,500	15,033	0.4	240.2	15.1
2004	1-Oct	29-Sep	364	46	62	55	6,310	92,800	15,528	0.9	413.5	16.3
2005	2-Oct	29-Sep	362	46	63	56	5,170	76,200	11,800	1.4	626.5	18.6
2006	1-Oct	29-Sep	363	47	61	54	6,450	118,000	22,724	1.6	731.7	17.9
2007	1-Oct	29-Sep	363	46	62	55	6,030	45,400	9,832	1.6	90.0	7.3
2008	1-Oct	29-Sep	364	44	63	56	5,210	44,600	9,342	1.3	233.3	8.8
2009	1-Oct	29-Sep	363	45	64	57	4,160	33,000	8,791	1.6	129.8	10.5
2010	1-Oct	30-Sep	364	46	62	56	3,920	60,400	10,241	1.1	250.6	12.1
2011	3-Oct	30-Sep	362	45	61	55	5,260	95,100	15,022	1.3	162.6	8.4
2012	3-Oct	27-Sep	360	45	65	55	4,800	35,200	9,753	1.2	180.6	7.1
2013	5-Oct	28-Sep	358	44	64	56	5,330	70,400	10,479	1.1	315.5	8.5
Mean	2-Oct	29-Sep	362	46	63	56	5,347	68,873	12,595	1.2	306.8	11.9
SD			2	1.1	1.3	0.7	843	27,701	4,177	0.3	205.5	4.4
CV			1%	2%	2%	1%	16%	40%	33%	29%	67%	37%

Figures

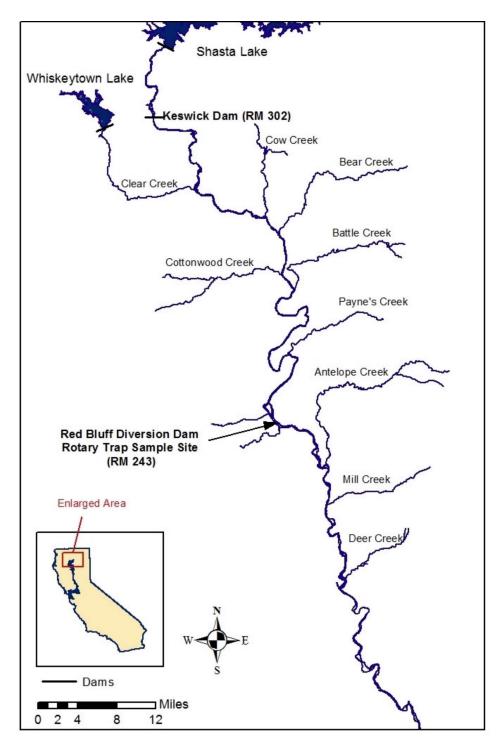


Figure 1.Location of Red Bluff Diversion Dam rotary trap stemsite on the Sacramento River, California (RM 243).

Red Bluff Diversion Dam Complex East Fish Ladder Rotary-Screw Traps Bypass Outfall Structure **East River Margin Habitat** Ŝ 9 Sacramento River Mid-channel Habitat ω თ 9 West River Margin Habitat Tehama-Colusa Red Bluff Research Pumping Plant Canal Head-works West Fish Ladder

Figure 2. Rotary-screw trap sampling transecteat Bluff Diversion Dam Site (RM 243) on the Saonton Reiver, California.

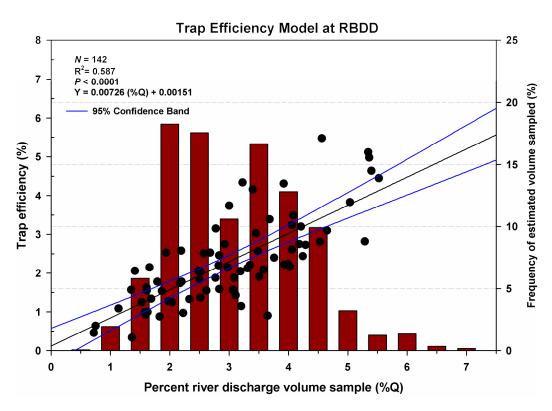


Figure 3. Trap efficiency model for combined diameter rotary traps at Red Bluff Diversion D(aRM 243), Sacramento River, CA. Mark-recapture trial (= 142) were used to estimate trap efficiencies to by ram indicates percentage of time traps sampled various levels (half percent bins) of river dischedules and September 2013.

BY 2002-2012 Fall Chinook Capture Fork Length Summaries

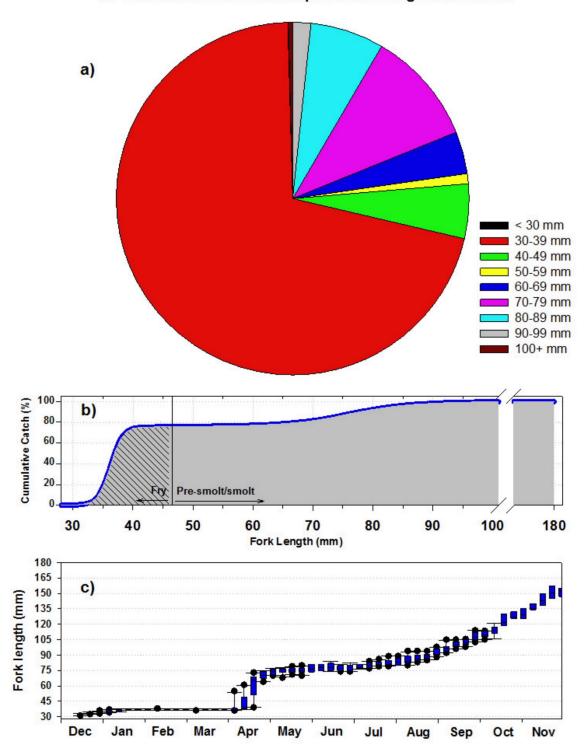


Figure 4. Fall Chinook fork length (a) capturoportions, (b) cumulative capture size curve, and (c) average weekly median boxplotsator thinook sampled by rotary traps at RBDD between December 2002 and September 2013.

BY 2002- 2012 Late-Fall Chinook Capture Fork Length Summaries

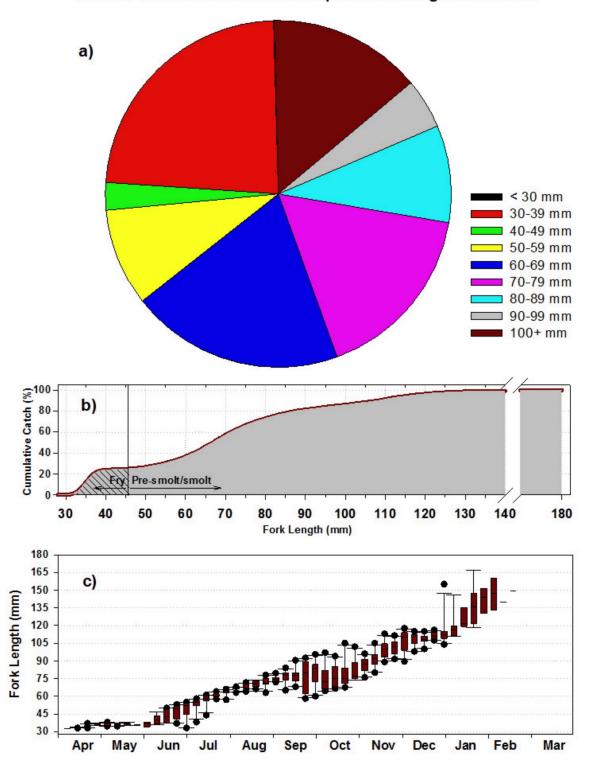


Figure 5. Late-fall Chinook fork length (a) preproportions, (b) cumulative capture size curve, and (c) average weekly median box to the test that chinook sampled by rotary traps at RBDD between April 2002 and March 32

BY 2002-2012 Winter Chinook Capture Fork Length Summaries

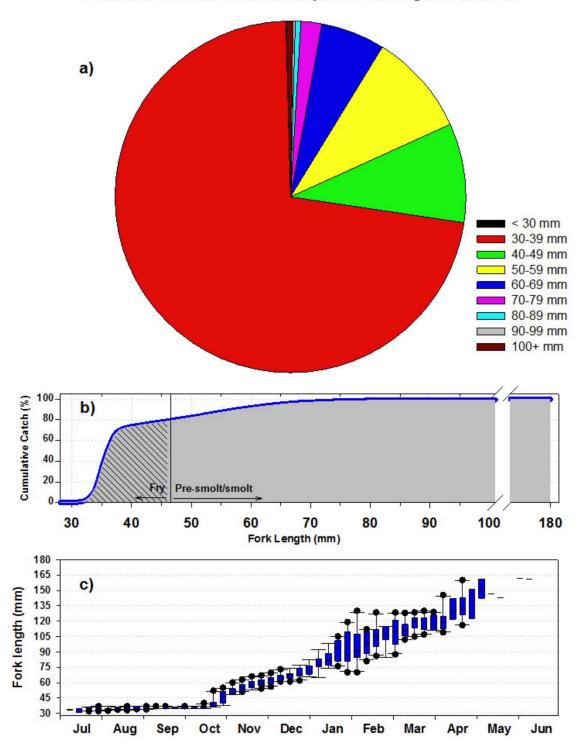


Figure 6. Winter Chinook fork length (a) capture portions, (b) cumulative capture size curve, and (c) average weekly median box to two inter Chinook sampled by rotary traps at RBDD between July 2002 and June 201

BY 2002-2012 Spring Chinook Capture Fork Length Summaries

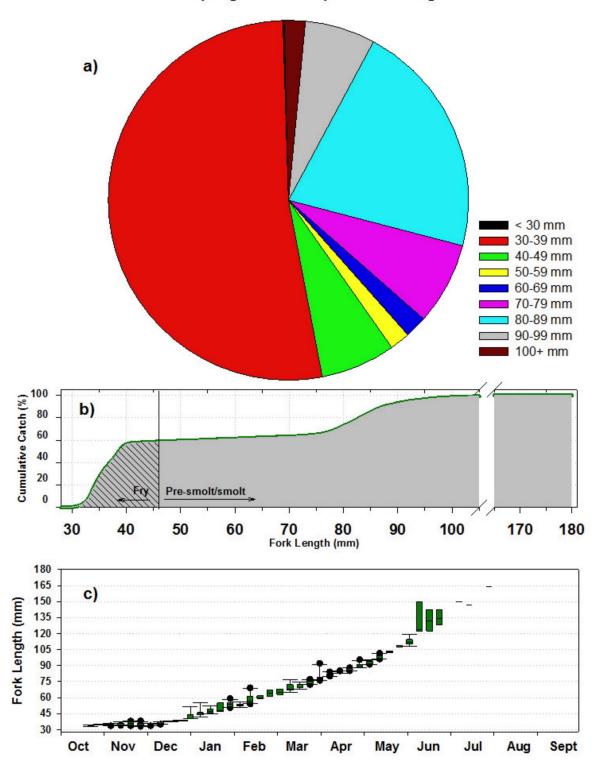


Figure 7. Spring Chinook fork length (a) captureportions, (b) cumulative capture size curve, and (c) average weekly median box total box traps at RBDD between October 2002 and Srepter 2013.

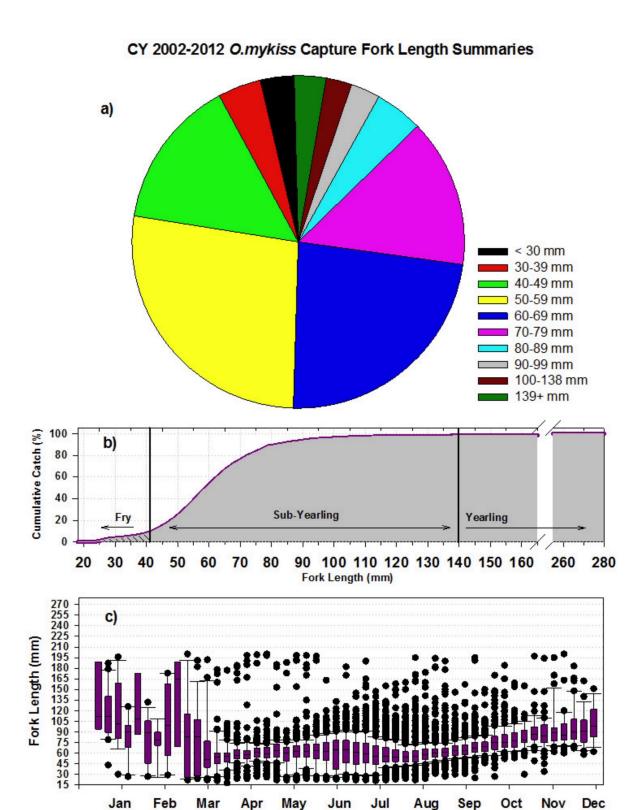


Figure 8.0. mykiss fork length (a) capture proportions, (b) cumulætivapture size curve, and (c) average weekly median boxplotsOfomykiss sampled by rotary traps at RBDD between April 2002 and December 2012.

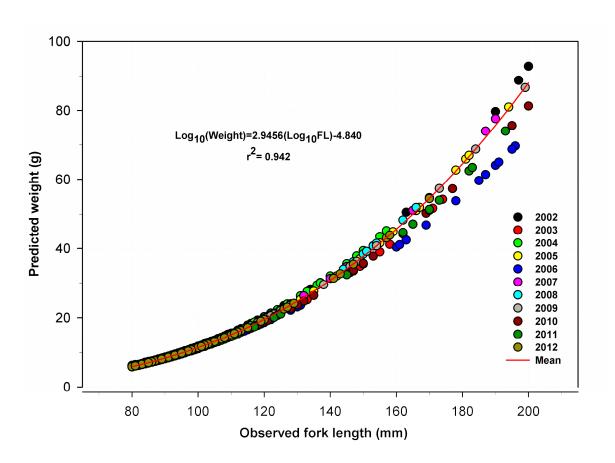


Figure 9. Predicted weight (g) for mykiss with measured fork lengths (FL) between 80 and 2000 using annual weight-length regression equation.

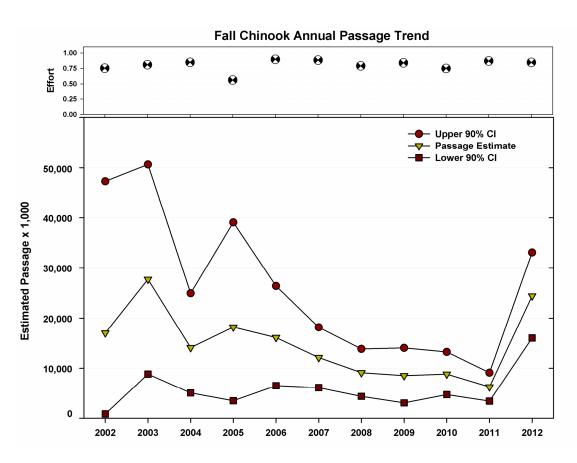


Figure 10. RBDD rotary trap fall Chinook ansaumlple effort and passage estimates with 90% centite intervals (CI) for the period December 2002 through September 2013

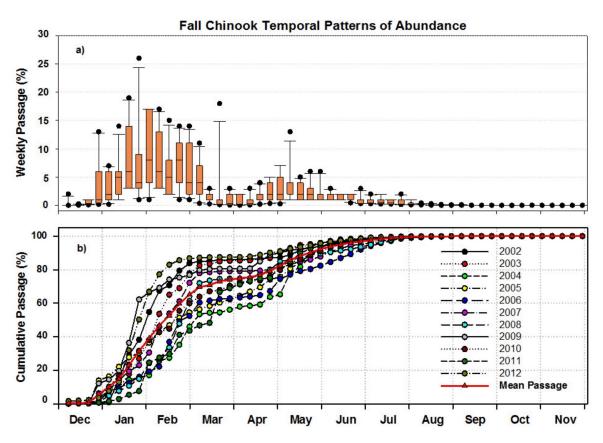


Figure 11. RBDD rotary trap fall Chinook (a) looks of weekly passage estimates relative to artificitial passage estimates and (b) cumulative weekly passage with 11-year mean pastraged line for the period December 2002 through the benefit of th

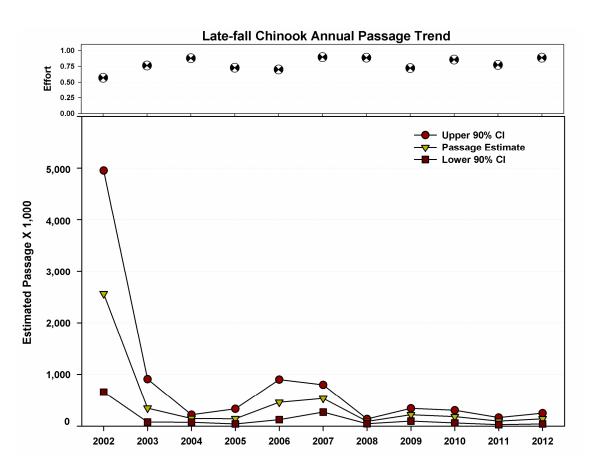


Figure 12. RBDD rotary trap late-fall Chinookuahsample effort and passage estimates with 90% dence intervals (CI) for the period April 2002 through March 2013.

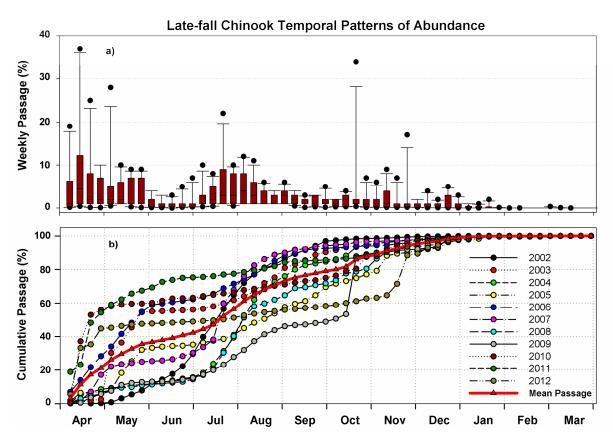


Figure 13. RBDD rotary trap late-fall Chino box plots of weekly passage estimates relativertoual total passage estimates and (b) cumulative weekly passage with 11-year massage trend line for the period April 2002 through March 2013.

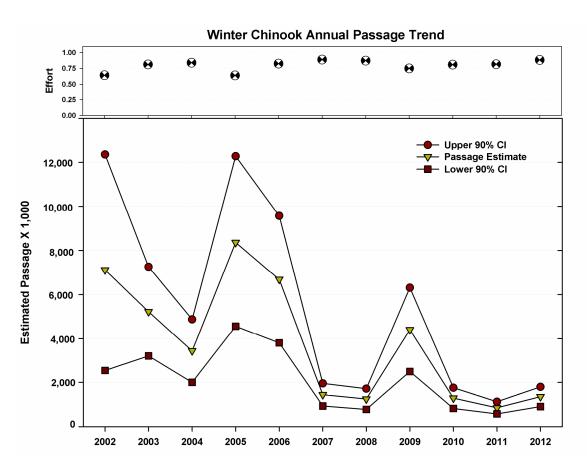


Figure 14. RBDD rotary trap winter Chinook ahsaumple effort and passage estimates with 90% idente intervals (CI) for the period July 2002 through June 2013.

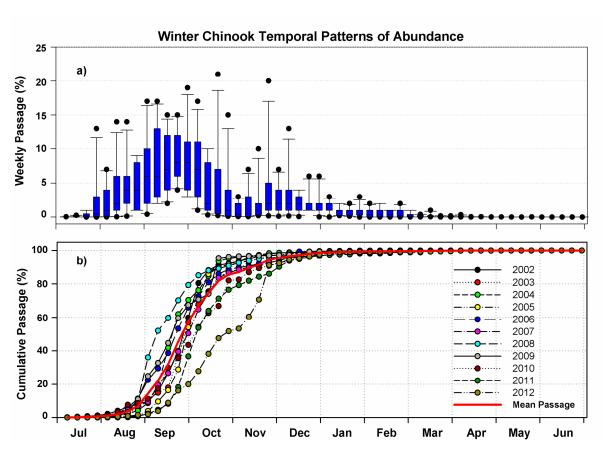


Figure 15. RBDD rotary trap winter Chinook (ax) blots of weekly passage estimates relative town total passage estimates and (b) cumulative weekly passage with 11-year means age trend line for the period July 2002 through 2013.

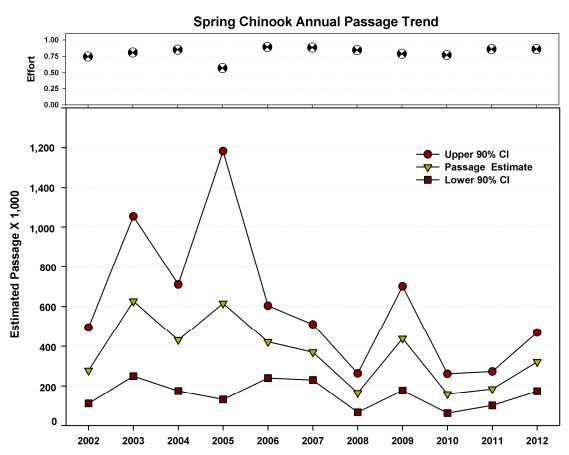


Figure 16. RBDD rotary trap spring Chinook ansurable effort and passage estimates with 90% centite intervals (CI) for the period October 2002 through September 2013.

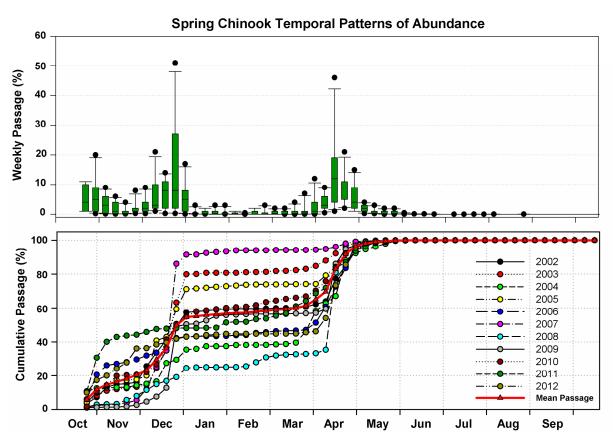


Figure 17. RBDD rotary trap spring Chinooko(xq) lbts of weekly passage estimates relative to wart notal passage estimates and (b) cumulative weekly passage with 11-year means appears trend line for the period October 2002 through the period october 2013.

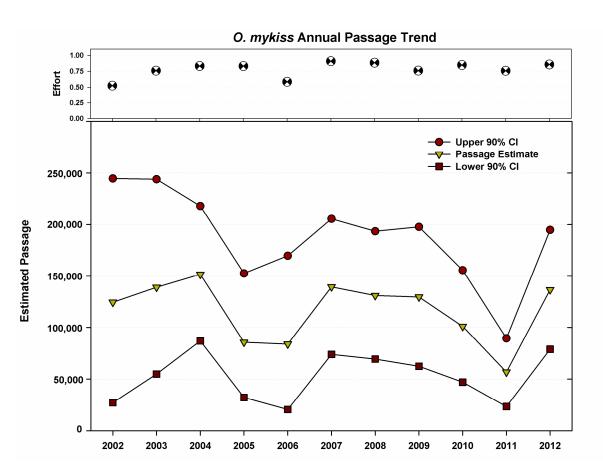


Figure 18. RBDD rotary trapmykiss annual sample effort and passage estimates with sonfidence intervals (CI) for the period April 2002 through December 2012.

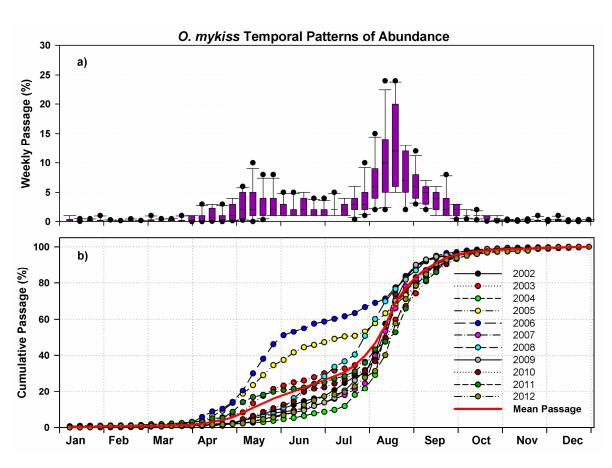


Figure 19. RBDD rotary trapmykiss (a) boxplots of weekly passage estimates relativænnual total passage estimates and (b) cumulative weekly passage with 11-year mean pastrægel line for the period April 2002 through Dedærn 2012.

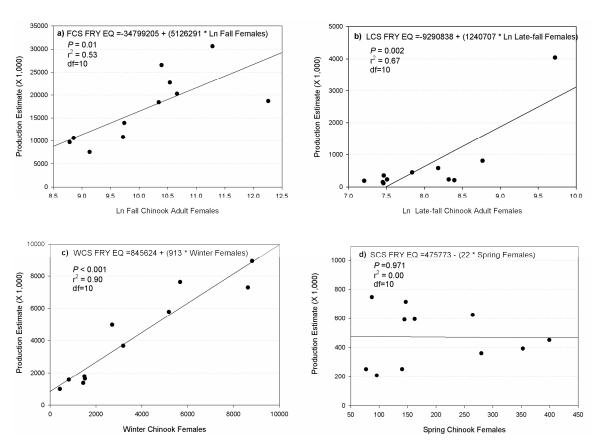


Figure 20. Relationships between a) fall, b) fatle-c) winter, and d) spring Chinook fry-equivalent duction estimates and estimated number of female adult Chinook salmont was of RBDD between 2002 and 2012. Note: fall late-fall adult females were natural log transformed due to extraordinars capement values estimated for the year 2002.

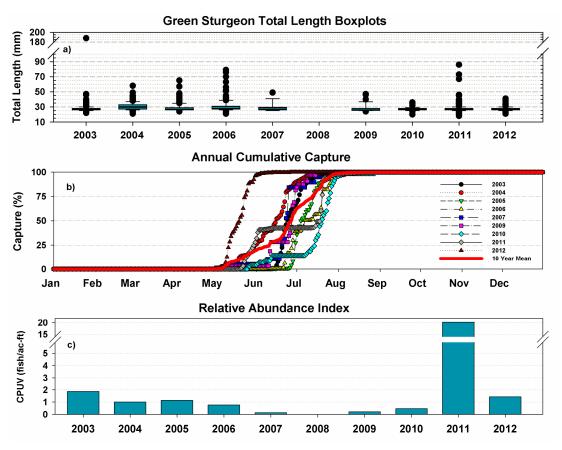


Figure 21. Green sturgeon a) annual total lengibiture boxplots, b) annual cumulative capture trenvish 10-year mean trend line, and c) relative abundance indices. All displitured by rotary trap at RBDD (RM 243) on the department River, CA between 2003 and 2012. Data from 2002 exclude the family sis due to limited effort and USBR Crown Etody resulting in incomparable sampling regimes and results.

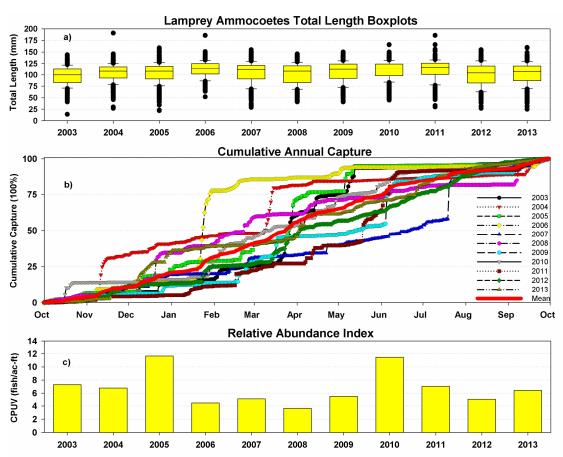


Figure 22. Unidentified lamprey ammocoetes a) **Ittet**agth distribution box plots, b) cumulative an **h c**apture trends, and c) relative abundance indices from rotary trap samples ected between October 1, 2002 and September 2003 by water year from the Sacramento River, CA at the RBDD (RM 243).

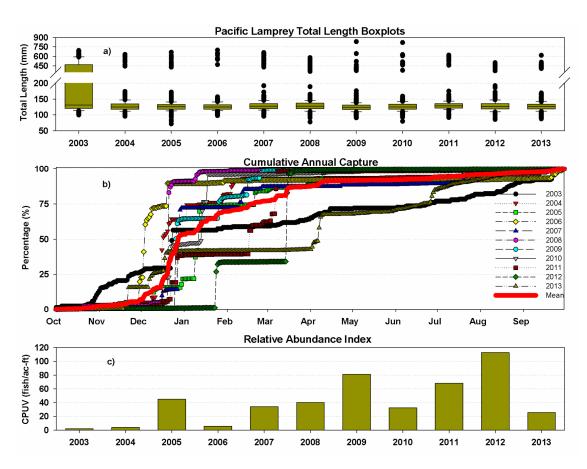


Figure 23. Pacific Lamprey (macropthalmia anudtsa)da) total length distribution box plots, b) noulative annual capture trends, and c) relative abundance indices from rotary tsamples collected between October 1, 2002 and Septem 10, 2013 by water year from the Sacramento River, CA at the RBDD (F3M) 2

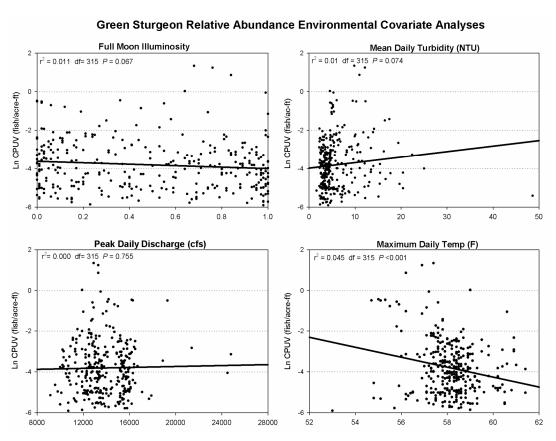


Figure 24. Regression analysis results of nlatuga(Ln) Green Sturgeon catch per unit volumeU(C)Pand a) full moon illuminosity, b) mean daily turbidity, c) peak datischarge and d) maximum daily temperatures at RBIAD fish captured by rotary trap at RBDD (RM 243) on the Upper SacraonRinter, CA between 2003 and 2012. Data from 2002 ded from analysis due to limited effort and USBR Crown Flow study. Items in incomparable sampling regimes and results

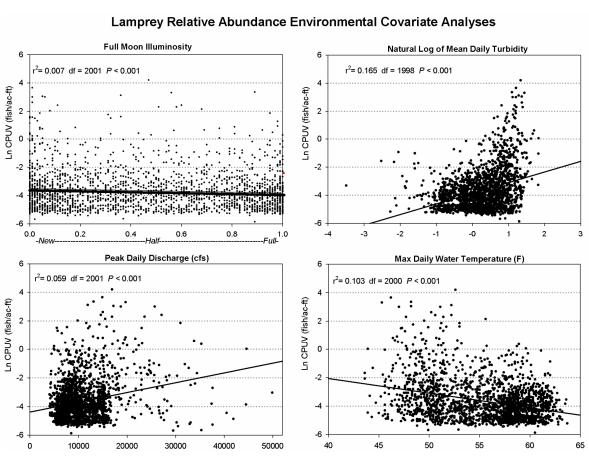


Figure 25. Regression analysis results of nathoga(Ln) Lampresypp. catch per unit volume (CPUV) and a) full moormilhosity, b) Ln mean daily turbidity, c) peak daily discharge a)nmaximum daily temperatures at RBDD. Allotaphtured by rotary trap at RBDD (RM 243) on the Upper Sacramento River, CA:elentwater year 2003 and 2013.

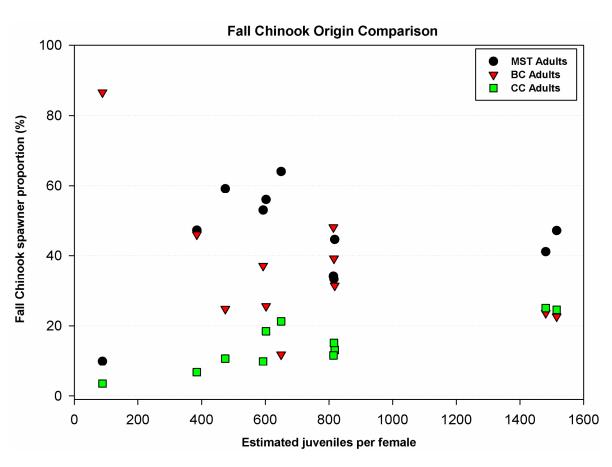


Figure 26. Comparison of estimated juveniles **posed** per estimated number of females in relation of fall Chinook spawners in the mainstem Sacramento River (MST) LeBateek (BC), and Clear Creek (CC) between Jean 2012.

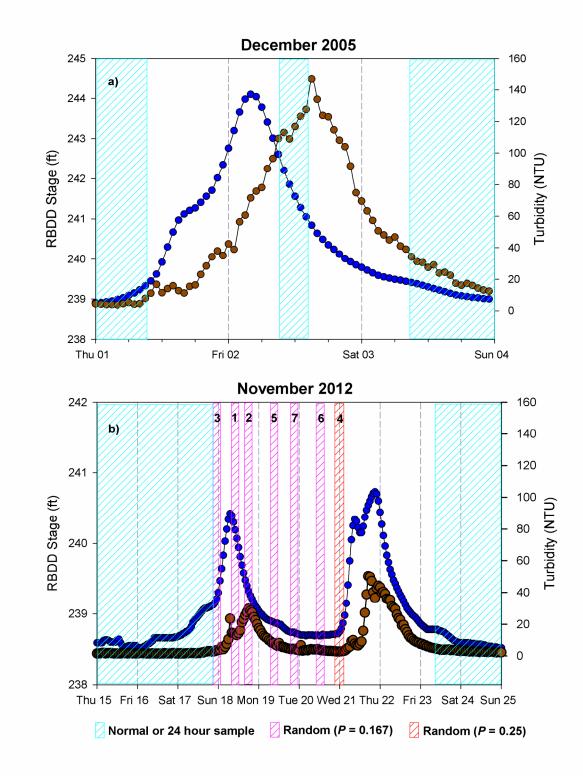


Figure 27. Timing comparison of RBDD staged(iseharge level) and turbidity measurements along with sample collection timessform events on a) December 1-4, 2005 and b) November 15-25, 2012. Numerals wishimple period boxes in figure b indicate rank of standardized Chinook passage statement (1) to least (7).

APPENDIX 1

Appendix 1: List of Tables

Table
A1.Summary of RBDD rotary trap annual effort, falhobk fry (<46 mm FL) passage estimates and lower and upper 90% confidence inter(CI), by brood year for the period December 2002 through September 2013
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Fall Chinook

Table A1. Summary of RBDD rotary trap annual teffed Chinook fry (<46 mm FL) passage estimates and lower and upper 90% confied intervals (CI), by brood year for the period December 2002 through September 2013.

шо ролов 2 ос		Estimated Fry	"	
Brood Year	Effort	Passage	Low 90% CI	Up 90% CI
2002	0.76	14,687,984	348,386	42,027,818
2003	0.81	23,612,094	6,953,966	44,283,689
2004	0.85	7,946,496	3,449,094	12,447,378
2005	0.56	11,740,225	2,452,034	24,687,255
2006	0.90	10,152,406	3,458,524	17,567,355
2007	0.88	9,594,099	4,834,813	14,353,810
2008	0.79	6,684,332	3,335,617	10,033,164
2009	0.84	6,900,302	2,190,210	11,662,489
2010	0.75	6,302,961	3,432,017	9,502,694
2011	0.87	4,437,956	2,380,436	6,498,878
2012	0.85	21,375,192	14,332,396	28,700,826

Table A2. Summary of RBDD rotary trap annualtefall Chinook pre-smolt/smolt (>45 mm FL) passage estimates and lower and upperconfidence intervals (CI), by brood year for the period December 2002 through tempor 2013.

		Estimated	"	
Brood Year	Effort	Smolt Passage	Low 90% CI	Up 90% CI
2002	0.76	2,350,433	505,837	5,318,021
2003	0.81	4,124,773	1,879,521	6,393,281
2004	0.85	6,161,742	1,626,946	12,527,167
2005	0.56	6,470,030	1,041,939	14,426,210
2006	0.90	5,955,245	3,056,683	8,855,302
2007	0.88	2,537,504	1,291,848	3,821,912
2008	0.79	2,431,215	1,034,851	3,827,754
2009	0.84	1,632,074	868,002	2,396,298
2010	0.75	2,539,519	1,288,830	3,850,851
2011	0.87	1,833,305	1,029,403	2,637,509
2012	0.85	3,054,227	1,692,494	4,416,322

Late-Fall Chinook

Table A3. Summary of RBDD rotary trap annual teffate-fall Chinook fry (<46 mm FL) passage estimates and lower and upper 90% decorde intervals (CI), by brood year for the period April 2002 through March 2013.

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		Estimated Fry		_
Brood Year	Effort	Passage	Low 90% CI	Up 90% CI
2002	0.57	442,393	84,832	901,368
2003	0.76	196,271	4,562	683,458
2004	0.88	24,382	8,802	40,591
2005	0.73	50,274	5,723	175,598
2006	0.70	284,999	41,006	634,496
2007	0.90	144,688	54,397	235,201
2008	0.89	10,489	4,347	17,813
2009	0.72	29,568	13,126	46,360
2010	0.86	113,667	26,705	200,935
2011	0.77	69,686	18,487	120,996
2012	0.89	67,479	9,925	136,431

Table A4. Summary of RBDD rotary trap annual telfate-fall Chinook presmolt/smolt (>45 mm FL) passage estimates and loamer upper 90% confidence intervals (CI), by brood year for the period Apard through March 2013.

, , , , , ,	<u> </u>	<u> </u>		
· · · · · · · · · · · · · · · · · · ·		Estimated		
Brood Year	Effort	Smolt Passage	Low 90% CI	Up 90% CI
2002	0.57	2,117,122	569,453	4,093,545
2003	0.76	149,976	72,089	230,841
2004	0.88	122,779	64,498	181,783
2005	0.73	93,407	35,067	160,738
2006	0.70	175,269	82,005	273,572
2007	0.90	390,932	213,642	568,595
2008	0.89	81,506	41,983	121,166
2009	0.72	190,256	83,201	297,652
2010	0.86	69,771	33,929	106,575
2011	0.77	27,354	9,535	45,914
2012	0.89	73,055	32,567	113,633
	2002 2003 2004 2005 2006 2007 2008 2009 2010 2011	2002 0.57 2003 0.76 2004 0.88 2005 0.73 2006 0.70 2007 0.90 2008 0.89 2009 0.72 2010 0.86 2011 0.77	Brood Year Effort Smolt Passage 2002 0.57 2,117,122 2003 0.76 149,976 2004 0.88 122,779 2005 0.73 93,407 2006 0.70 175,269 2007 0.90 390,932 2008 0.89 81,506 2009 0.72 190,256 2010 0.86 69,771 2011 0.77 27,354	Brood Year Effort Smolt Passage Low 90% CI 2002 0.57 2,117,122 569,453 2003 0.76 149,976 72,089 2004 0.88 122,779 64,498 2005 0.73 93,407 35,067 2006 0.70 175,269 82,005 2007 0.90 390,932 213,642 2008 0.89 81,506 41,983 2009 0.72 190,256 83,201 2010 0.86 69,771 33,929 2011 0.77 27,354 9,535

Winter Chinook

Table A5. Summary of RBDD rotary trap annual teffiointer Chinook fry (<46 mm FL) passage estimates and lower and upper 90% confiedentervals (CI), by brood year for the period July 2002 through June 2013.

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_	_	Estimated Fry		
Brood Year	Effort	Passage	Low 90% CI	Up 90% CI
2002	0.64	6,381,286	2,156,758	11,217,962
2003	0.81	4,420,296	2,743,637	6,096,955
2004	0.84	3,087,102	1,812,619	4,361,584
2005	0.64	7,533,380	4,225,130	10,841,630
2006	0.83	5,813,140	3,307,323	8,318,957
2007	0.89	1,158,791	744,804	1,572,817
2008	0.87	1,063,919	662,381	1,465,748
2009	0.75	3,587,134	2,076,422	5,098,125
2010	0.81	875,049	603,549	1,146,644
2011	0.82	638,056	441,983	834,289
2012	0.89	722,048	545,751	898,345

Table A6. Summary of RBDD rotary trap annual teffiointer Chinook pre-smolt/smolt (>45 mm FL) passage estimates and lower and upper confidence intervals (CI), by brood year for the period July 2002 through June 200

***************************************		Estimated		
Brood Year	Effort	Smolt Passage	Low 90% CI	Up 90% CI
2002	0.64	737,755	373,538	1,149,079
2003	0.81	800,719	453,256	1,169,559
2004	0.84	347,581	179,502	519,265
2005	0.64	829,302	324,860	1,442,763
2006	0.83	873,940	487,244	1,264,701
2007	0.89	281,773	180,254	387,123
2008	0.87	181,071	110,592	252,089
2009	0.75	815,188	410,512	1,222,586
2010	0.81	410,341	210,252	613,810
2011	0.82	210,920	130,861	291,312
2012	0.89	627,771	354,764	900,897

Spring Chinook

Table A7. Summary of RBDD rotary trap annualteffpring Chinook fry (<46 mm FL) passage estimates and lower and upper 90% conficientervals (CI), by brood year for the period October 2002 through September 2013.

			Estimated Fry	_	
Broo	od Year	Effort	Passage	Low 90% CI	Up 90% CI
2	002	0.75	159,084	67,900	255,023
2	003	0.81	502,386	189,371	857,899
2	004	0.85	155,053	59,655	250,451
2	005	0.57	427,719	111,396	925,898
2	006	0.89	174,186	114,642	233,907
2	007	0.89	336,714	212,765	460,712
2	800	0.85	40,213	26,016	54,448
2	009	0.79	219,627	91,683	347,845
2	010	0.77	89,213	39,829	138,597
2	011	0.86	88,355	63,469	113,274
2	012	0.86	134,028	82,843	185,271

Table A8. Summary of RBDD rotary trap annual test pring Chinook pre-smolt/smolt (>45 mm FL) passage estimates and lower and upper confidence intervals (CI), by brood year for the period October 2002 through Sepber 2013.

		Estimated		
Brood Year	Effort	Smolt Passage	Low 90% CI	Up 90% CI
2002	0.75	118,393	43,022	239,870
2003	0.81	124,529	59,434	197,777
2004	0.85	275,898	113,564	460,990
2005	0.57	187,828	19,676	460,441
2006	0.89	247,250	123,621	371,968
2007	0.89	32,787	15,894	51,271
2008	0.85	124,460	40,130	208,954
2009	0.79	218,778	83,930	354,607
2010	0.77	69,753	21,938	123,577
2011	0.86	95,935	37,782	159,702
2012	0.86	186,869	89,566	284,936

Table A9. River Lampre ampetra ayreşiannual capture, catch per unit volume (CPUV) and total length summaries for River Lampa ured by RBDD rotary traps between water year (WY) 2003 and 2013.

	,	CPUV	Min TL	Max TL	Mean	Median
WY	Catch	Fish/ac-ft	(mm)	(mm)	(mm)	(mm)
2003	0	0.00	-	-	=	-
2004	1	0.01	102	102	102	-
2005	0	0.00	-	-	-	-
2006	0	0.00	-	-	-	-
2007	0	0.00	-	-	-	-
2008	0	0.00	-	-	-	-
2009	0	0.00	-	-	-	-
2010	1	0.01	110	110	110	-
2011	26	0.23	99	151	121	121
2012	4	0.02	128	168	144	140
2013	0	0.00	-	-	-	-
Mean	2.9	0.02	109.8	132.8	119.3	130.5
SD	7.8	0.07	13.0	31.8	18.2	13.4
CV	266.5%	279.2%	11.9%	24.0%	15.3%	10.3%

Table A10. Pacific Brook Lamprlesympetra pacifica, annual capture, catch per unit volume (CPUV) and total length summaries for RaBifiook Lamprey captured by RBDD rotary traps between water year (WY) 2003 and 2013.

		CPUV	Min TL	Max TL	Mean	Median
WY	Catch	Fish/ac-ft	(mm)	(mm)	(mm)	(mm)
2003	6	0.06	98	132	116	114.5
2004	1	0.01	159	159	159	-
2005	0	0.00	-	-	-	-
2006	0	0.00	-	-	-	-
2007	0	0.00	-	-	-	-
2008	0	0.00	-	-	-	-
2009	0	0.00	-	-	-	-
2010	1	0.02	120	120	120	120
2011	1	0.01	147	147	147	147
2012	6	0.04	112	156	138	142
2013	21	0.12	110	148	124	122
Mean	3.3	0.02	124.3	143.7	134.0	129.1
SD	6.3	0.04	23.6	14.9	16.9	14.4
CV	192.8%	159.7%	19.0%	10.4%	12.6%	11.2%

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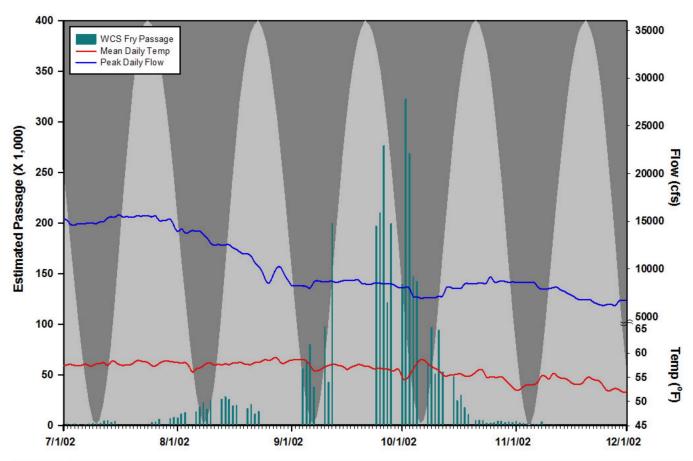


Figure A1. Brood Year 2002 winter Chinook fry passage with moon illuminosity indicated by back ground shading (peak of light gray equals full moon), mean daily water temperatures (red), and peak daily flows (blue) at Red Bluff Diversion Dam.

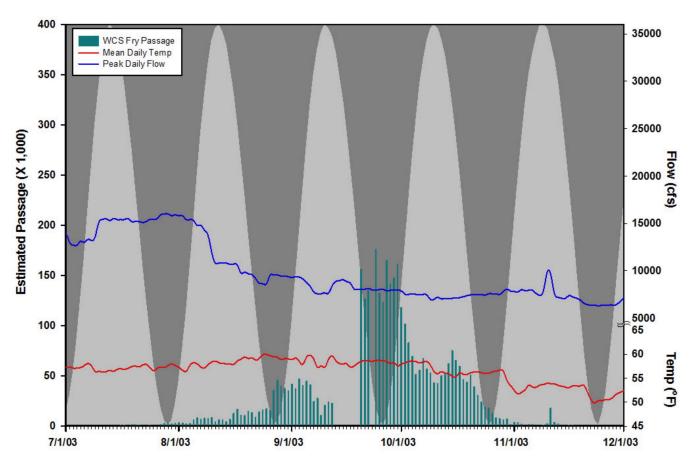


Figure A2. Brood Year 2003 winter Chinook fry passage with moon illuminosity indicated by back ground shading (peak of light gray equals full moon), mean daily water temperatures (red), and peak daily flows (blue) at Red Bluff Diversion Dam.

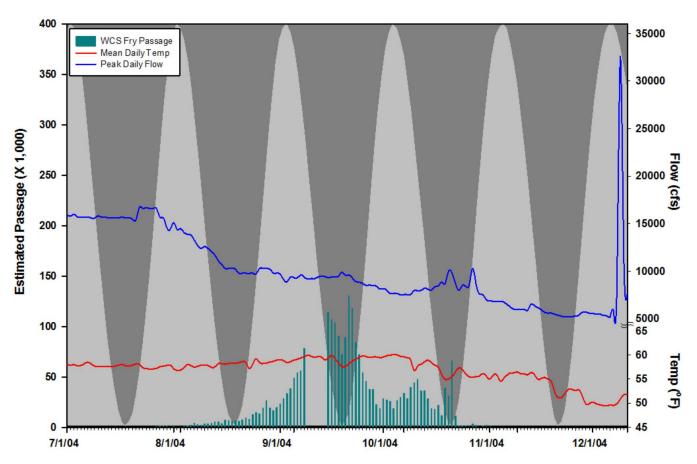


Figure A3. Brood Year 2004 winter Chinook fry passage with moon illuminosity indicated by back ground shading (peak of light gray equals full moon), mean daily water temperatures (red), and peak daily flows (blue) at Red Bluff Diversion Dam.

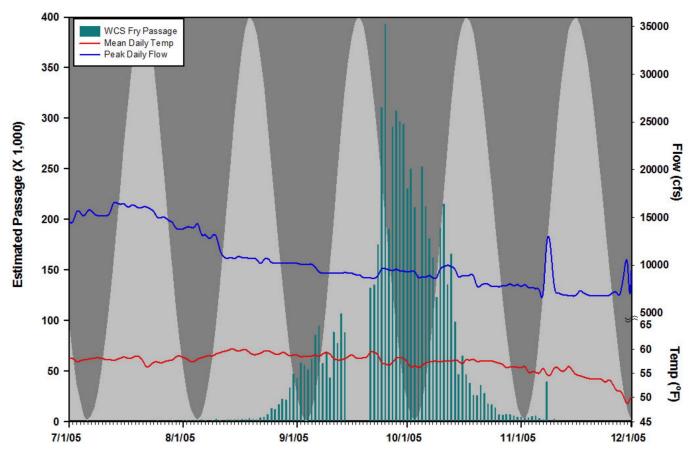


Figure A4. Brood Year 2005 winter Chinook fry passage with moon illuminosity indicated by back ground shading (peak of light gray equals full moon), mean daily water temperatures (red), and peak daily flows (blue) at Red Bluff Diversion Dam.

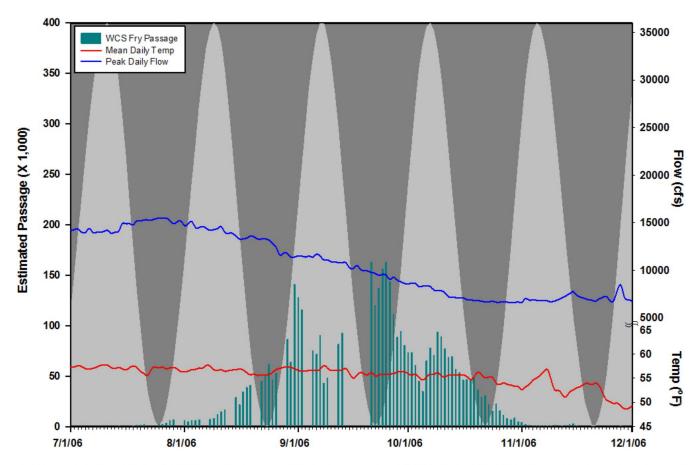


Figure A5. Brood Year 2006 winter Chinook fry passage with moon illuminosity indicated by back ground shading (peak of light gray equals full moon), mean daily water temperatures (red), and peak daily flows (blue) at Red Bluff Diversion Dam.

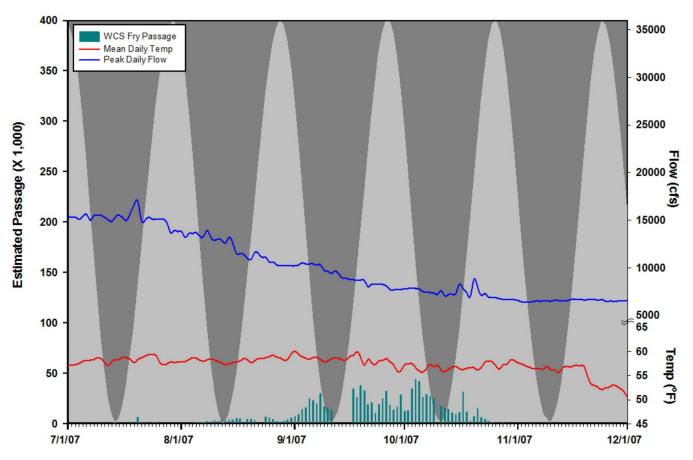


Figure A6. Brood Year 2007 winter Chinook fry passage with moon illuminosity indicated by back ground shading (peak of light gray equals full moon), mean daily water temperatures (red), and peak daily flows (blue) at Red Bluff Diversion Dam.

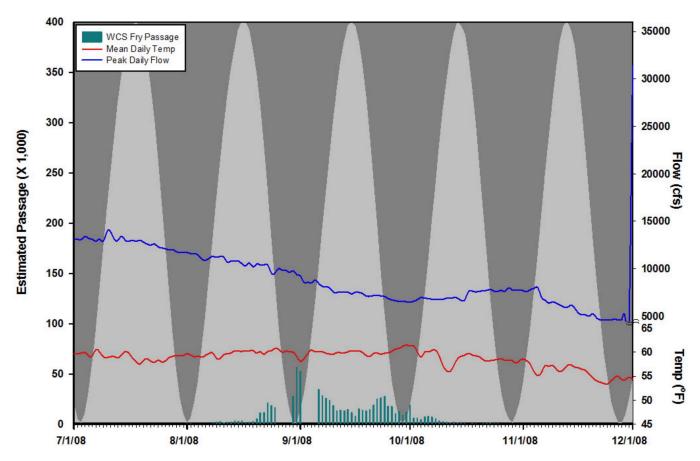


Figure A7. Brood Year 2008 winter Chinook fry passage with moon illuminosity indicated by back ground shading (peak of light gray equals full moon), mean daily water temperatures (red), and peak daily flows (blue) at Red Bluff Diversion Dam.

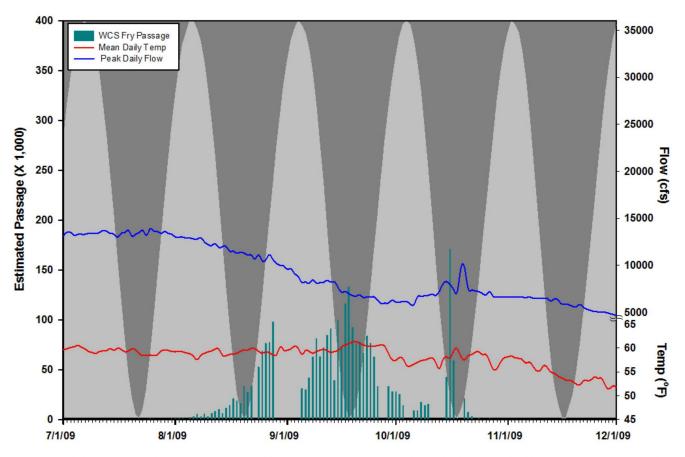


Figure A8. Brood Year 2009 winter Chinook fry passage with moon illuminosity indicated by back ground shading (peak of light gray equals full moon), mean daily water temperatures (red), and peak daily flows (blue) at Red Bluff Diversion Dam.

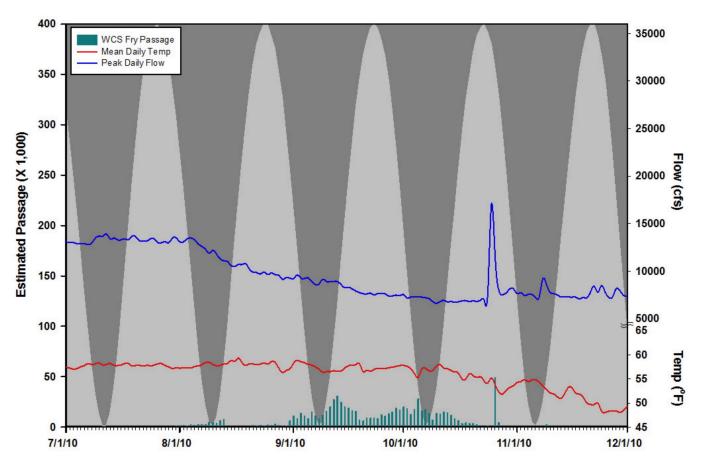


Figure A9. Brood Year 2010 winter Chinook fry passage with moon illuminosity indicated by back ground shading (peak of light gray equals full moon), mean daily water temperatures (red), and peak daily flows (blue) at Red Bluff Diversion Dam.

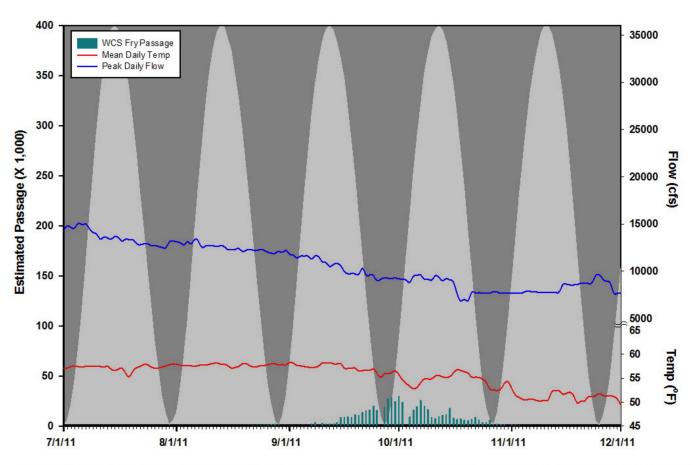


Figure A10. Brood Year 2011 winter Chinook fry passage with moon illuminosity indicated by back ground shading (peak of light gray equals full moon), mean daily water temperatures (red), and peak daily flows (blue) at Red Bluff Diversion Dam.

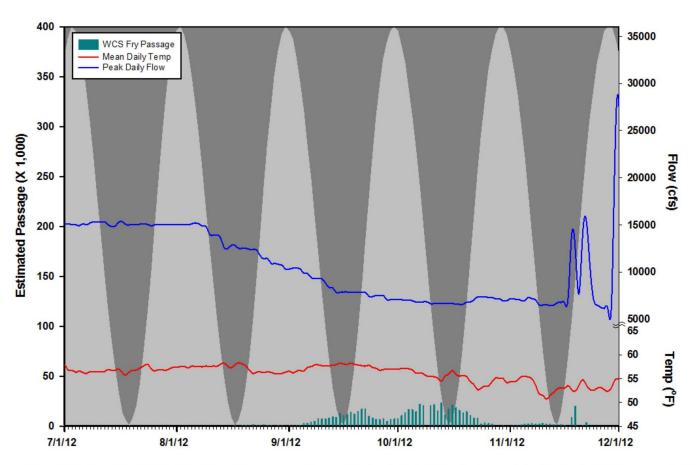


Figure A11. Brood Year 2012 winter Chinook fry passage with moon illuminosity indicated by back ground shading (peak of light gray equals full moon), mean daily water temperatures (red), and peak daily flows (blue) at Red Bluff Diversion Dam.

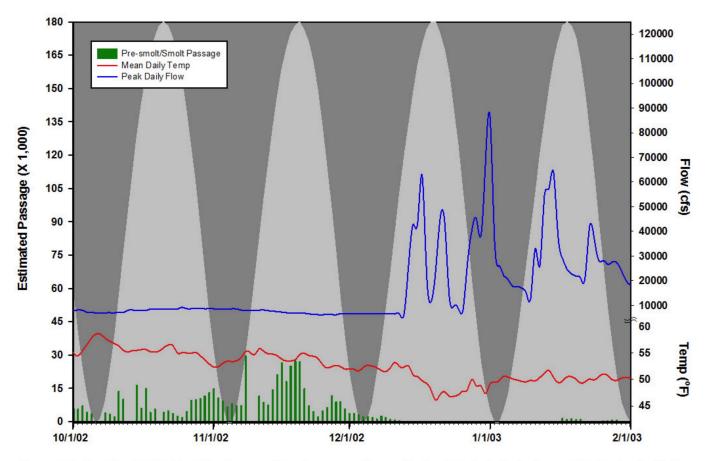


Figure A12. Brood Year 2002 winter Chinook pre-smolt/smolt passage with moon illuminosity indicated by back ground shading (peak of light gray equals full moon), mean daily water temperatures (red), and peak daily flows (blue) at Red Bluff Diversion Dam.

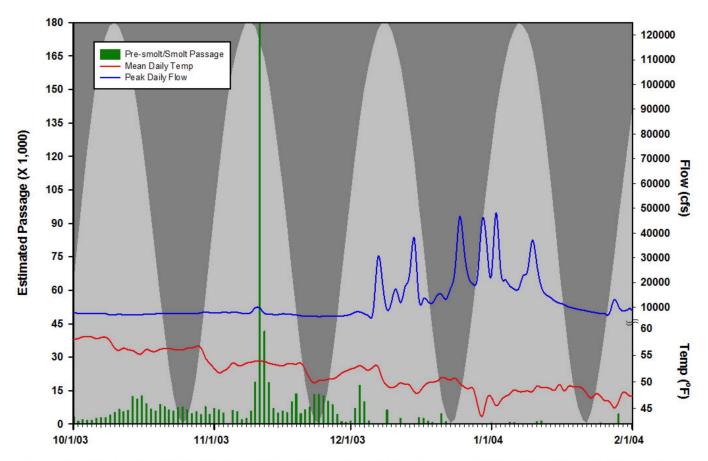


Figure A13. Brood Year 2003 winter Chinook pre-smolt/smolt passage with moon illuminosity indicated by back ground shading (peak of light gray equals full moon), mean daily water temperatures (red), and peak daily flows (blue) at Red Bluff Diversion Dam.

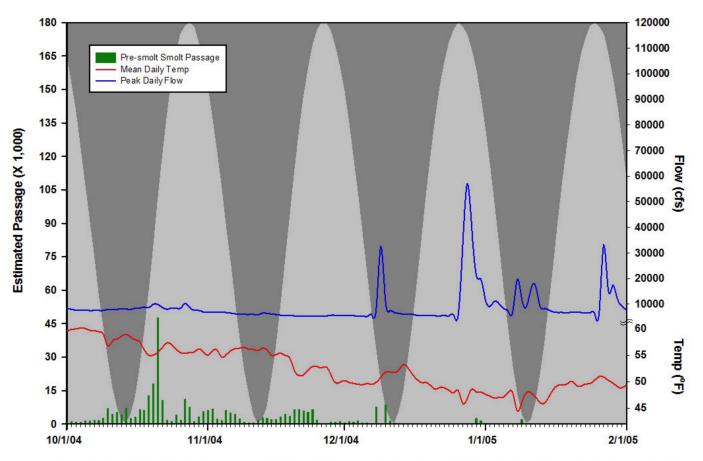


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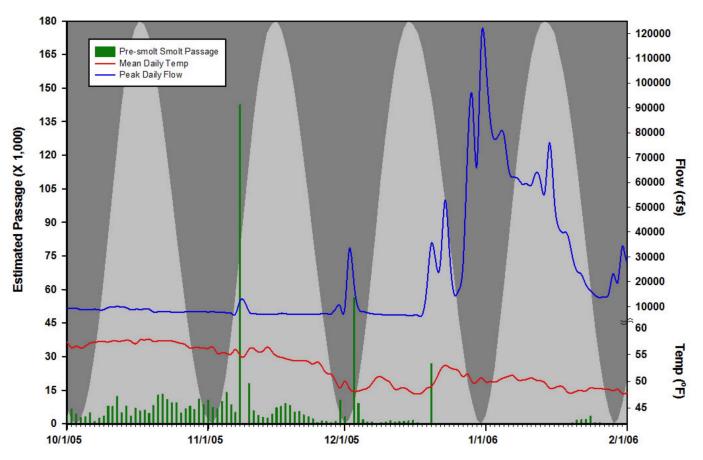


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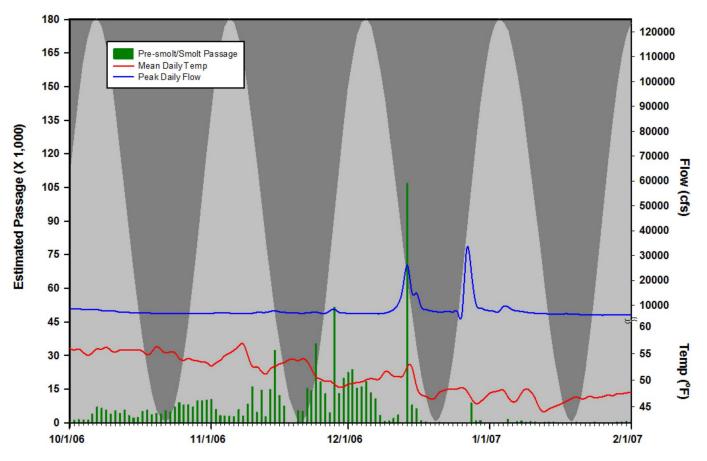


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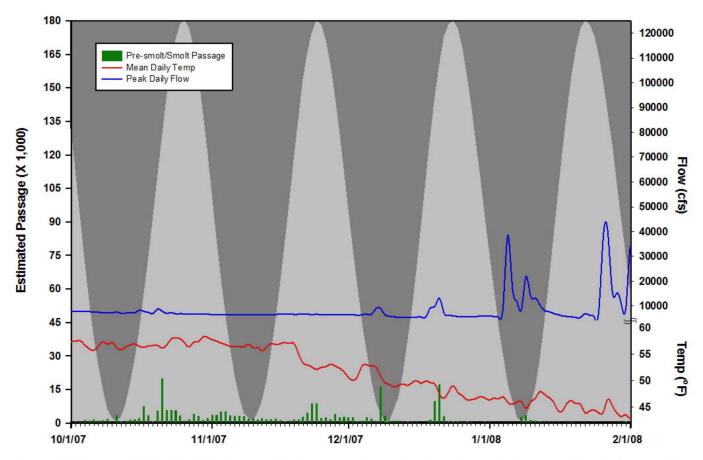


Figure A17. Brood Year 2007 winter Chinook pre-smolt/smolt passage with moon illuminosity indicated by back ground shading (peak of light gray equals full moon), mean daily water temperatures (red), and peak daily flows (blue) at Red Bluff Diversion Dam.

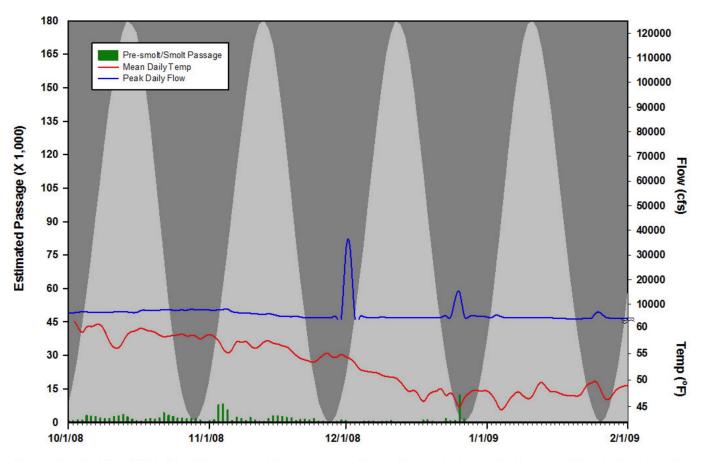


Figure A18. Brood Year 2008 winter Chinook pre-smolt/smolt passage with moon illuminosity indicated by back ground shading (peak of light gray equals full moon), mean daily water temperatures (red), and peak daily flows (blue) at Red Bluff Diversion Dam.

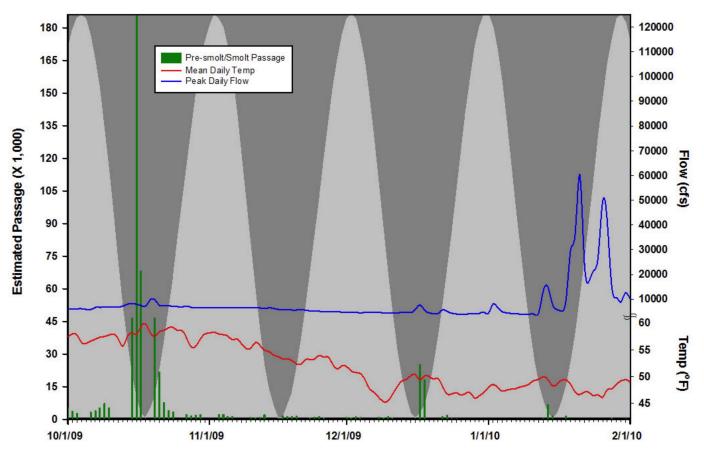


Figure A19. Brood Year 2009 winter Chinook pre-smolt/smolt passage with moon illuminosity indicated by back ground shading (peak of light gray equals full moon), mean daily water temperatures (red), and peak daily flows (blue) at Red Bluff Diversion Dam.

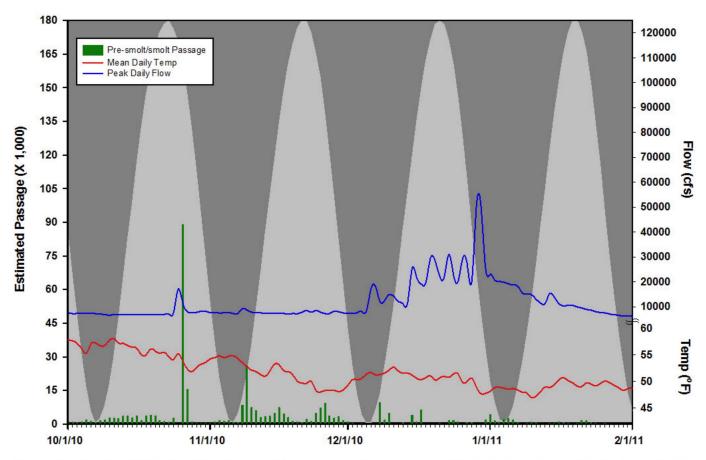


Figure A20. Brood Year 2010 winter Chinook pre-smolt/smolt passage with moon illuminosity indicated by back ground shading (peak of light gray equals full moon), mean daily water temperatures (red), and peak daily flows (blue) at Red Bluff Diversion Dam.

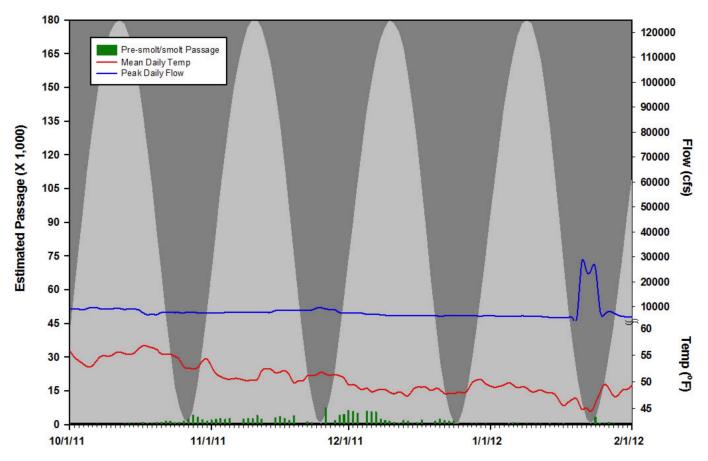


Figure A21. Brood Year 2011 winter Chinook pre-smolt/smolt passage with moon illuminosity indicated by back ground shading (peak of light gray equals full moon), mean daily water temperatures (red), and peak daily flows (blue) at Red Bluff Diversion Dam.

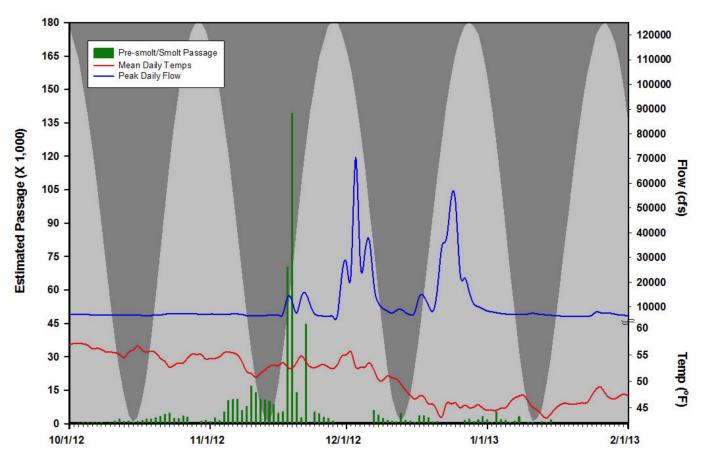


Figure A22. Brood Year 2012 winter Chinook pre-smolt/smolt passage with moon illuminosity indicated by back ground shading (peak of light gray equals full moon), mean daily water temperatures (red), and peak daily flows (blue) at Red Bluff Diversion Dam.